

Assessing the importance of seed priming for seed germination in Muscari.

IMAN ROHOLLAHI¹, Narjes Labbaf¹, and Amir Mohammad NAJI¹

¹Shahed University

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Abstract

Muscari with absorbing color is a widespread and commercial species. As a pot and bedding bulb flower has excellent adaptability and vigorous growth in late winter and early spring. In our experiment, the quantity and uniformity of Muscari propagation by seed and the importance of seed priming will be measured. The selected *M. neglectum* populations which already were gathered in our previews research and *M. armeniacum* were evaluated under separated sulfuric acid (5, 15, and 20 min) and hot water (5, 15, and 20 min) scarification plus 15, 30, and 45 days stratification in a completely randomized design, four replications were planted in separate containers. Seeds of *M. armeniacum* only germinate (70%) by 15 min sulfuric acid plus 45-day stratification treatments. Based on the analysis of variance and mean comparison data that only were reported in *M. neglectum*, their seeds optimally germinated by 5 and 15 min sulfuric acid treatment plus 45-day stratification. In conclusion, *M. neglectum* seeds germination is tuned to take place well in mid-winter under natural conditions; on the other hand, 45 days of lengthy constant stratification and 5 min sulfuric acid priming will accelerate *M. neglectum* seeds germination.

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Muscari with absorbing color is a widespread and commercial species. As a pot and bedding bulb flower has excellent adaptability and vigorous growth in late winter and early spring. In our experiment, the quantity and uniformity of *Muscari* propagation by seed and the importance of seed priming will be measured. The selected *M. neglectum* populations which already were gathered in our previous research and *M. armeniacum* were evaluated under separated sulfuric acid (5, 15, and 20 min) and hot water (5, 15, and 20 min) scarification plus 15, 30, and 45 days stratification in a completely randomized design, four replications were planted in separate containers. Seeds of *M. armeniacum* only germinate (70%) by 15 min sulfuric acid plus 45-day stratification treatments. Based on the analysis of variance and mean comparison data that only were reported in *M. neglectum*, their seeds optimally germinated by 5 and 15 min sulfuric acid treatment plus 45-day stratification. In conclusion, *M. neglectum* seeds germination is tuned to take place well in mid-winter under natural conditions; on the other hand, 45 days of lengthy constant stratification and 5 min sulfuric acid priming will accelerate *M. neglectum* seeds germination.

Keywords: Germination, Hot water, Uniformity, Sulfuric acid, Stratification,

1. Introduction:

The demand and endeavor for breeding *Muscari* species with great bulbous plants are increase because of its excellent horticultural specifics, economic and pharmaceutical importance (Pehlivan and Özler, 2003). Among the noteworthy species in the *Muscari* genus is '*M. neglectum* Guss. ex Ten.' that forcing bulbs as cut flowers or pots is very probably, also it is beneficial for gardens and

1 landscape (Lost and Regained, 1993). *Muscari* flower colors are infinitely varied ,and it's worth
2 noting that its natural pigment has many particularly advantageous compounds which may support
3 human well-being (Fawzi Mahomoodally *et al.*, 2021). *Muscari armeniacum* with absorbing property
4 is a popular and commercial species. It is appropriate as pot, bedding, garden, and woodland owing
5 to greater adaptability, vigorous growth, and salient blue colors in spring (Grey-Wilson C, Mathew
6 B, 1981). *M. neglectum* growth generally in the whole Mediterranean but it distributed into Russia
7 and Iran (Davis, P.H. & Stuart, 1984). *Muscari* as the endemic flower generally grows in the central
8 area of the Zagros mountainous region of Iran (Candido *et al.*, 2017; Ghahreman, 1984). Labbaf,
9 Rohollahi, and Naji, (2020) investigation showed pretty high values of genetic variation and
10 morphological characteristics within the population and low genetic diversity level among nine
11 genotypes of *M. neglectum*, belonging to Iran's nine distribution regions.

12 *Muscari* produces many seeds which has a short juvenile period, so their propagation can occur
13 from seeds. Other advantage of seed propagation is that, unlike vegetatively propagated resources,
14 the plants grown from seed are virus-free (Kamenetsky and Okubo, 2012). Therefore, despite their
15 seed propagation significance, there are still problems with their germination. *Muscari* seeds usually
16 germinate better within their native environments. They often take a long time to germinate which is
17 not favorable for propagation. *Muscari* seeds have a hard coating with low water permeability which
18 indicate morphophysiological dormancy with a dormancy period of about 7–8 months (Herrmann,
19 Gabriele, and Durka Walter, 2006; Baskin and Baskin, 2014). In natural environments, seed
20 dormancy induces inactivation and prevents seed germination on a maternal level, allowing for
21 greater dispersal and passage of harmful environmental conditions (Sohindji *et al.*, 2020). Final seed
22 germination in *M. neglectum* was found to be remarkably suppressed by white light (photoinhibition).
23 Therefore, seed dormancy, long germination period, and low germination rate are major problems for

Muscari species (Doussi and Thanos, 2002; Baskin and Baskin, 2014; Candido *et al.*, 2017). However, before sowing, dormant seeds should be handled. Pre-sowing treatments like cold stratification and chemical or mechanical scarification could minimize seed hardness and increase germination rates in different seeds plant (Baskin and Baskin, 2014; Dürr *et al.*, 2015; Martínez-Díaz *et al.*, 2018; Sohindji *et al.*, 2020).

M. comosum seeds germinated slowly and steadily at a cool temperature range (5–15°C). It was seriously inhibited in the dark, over 15°C (Doussi and Thanos, 2002). Time, uniformity, and synchronization index are important not only for seed technologists and physiologists, but also for ecologists, since they can be used to measure and forecast the dynamics of germination phase (Ranal and De Santana, 2006).

The current research aims to fill this information gap by exploring how to enhance the germination of selected populations of *M. neglectum* and *Muscari armeniacum*. Germination parameters for native species, especially rare and endemic species, are critical for population survival via the sexual reproduction. (Bu *et al.*, 2008). The specific goals had achieved: (1) morphological diversity and germinability investigation; (2) to increase germination rate and uniformity, reduce germination period; by (3) introducing laboratory method for breaking seed dormancy.

2. Material and methods

2.1. Plant Materials

The study was conducted with *M. neglectum* seeds resulting from endemic population samples (Table 1) that showed the most morphological characteristics as well as higher flowering percentage in labbaf, Rohollahi and Naji (2020) investigation. furthermore, their Genetic diversity investigation

indicates that these populations are in the same cluster. Bulbs were screened based on circumference (6.7 cm), (De Hertogh and Le Nard, 1993) and selected bulbs were cultivated in 20 cm diameter potting and moved to the greenhouse after cold treatment (15 weeks at 9 °C) in September 2017. In April 2018, seeds from *M. neglectum* populations were obtained at the same time. *M. armeniacum* seeds were bought from ‘www.plant-world-seeds.com’.

2.2. Germination

This study was organized in two separate factorial experiments base on a completely randomized design (CRD) with 4 replicates, and each treatment contained 25 seeds. The first and second experiment treatments were acid plus cold stratification and hot water plus cold stratification, respectively. Before providing any treatment, the seeds were sterilized by soaking for 5 min in a 2 percent NaOCl solution and then rinsing three times with distilled water. Chemical scarification with sulfuric acid (70 percent v/v) for 0, 5, 15, and 20 min is used in the first experiment. Seeds were rinsed with distilled water after scarification treatments and put on double filter paper disks (Whatman No.1) in 9-cm glass Petri dishes moistened with 7 mL sterilized distilled water and stratified at 4°C for 0, 15, 30 and 45 days. They transferred to the germinator (± 0.5 °C– darkness) at the optimum values of 10 °C (Doussi and Thanos, 2002). In the second experiment, the seeds were scarified with hot water (70°C) for 0, 5, 15, and 20 min. The number of germinated seeds was counted every day. A seed was considered as germinated once its radicle had protruded about 2mm from the seed coat (ISTA, 2017). The seed count was concluded when no additional seeds germinated. The following formula was used to measure germination rate and germination percentage (GR and GP), respectively (Maguire, 1962; Olmez, Gokturk and Temel, 2007)

$$GP = n/v \times 100 \quad (1)$$

Whereas n: the number of seeds that have germinated and v: the number of viable seeds that have been initiated,

$$GR = [(n_1 \times t_1) + (n_2 \times t_2) + (n_i \times t_i)] / T \quad (2)$$

Where n_i represents the number of days in each counting, t_i represents the number of germinated seeds in each counting, and T represents the cumulative number of germinated seeds.

The mean germination time (MGT) and mean germination rate (MDR) were determined in this study according to the methods defined by Dorneles, Ranal and Santana, (2005). In usual, during germination activity, more or less deviation from normality can be fined in the context of time. Other drawback of this assessment is that for comparisons, the mean germination period of samples or treatments should be the same, also variances and standard deviations depending on the mean's magnitude (Ranal and De Santana, 2006). The variation coefficient of germination time (CVt) was recommended to calculate the uniformity or instability of germination compared to the mean germination time (Dorneles, Ranal, and Santana, 2005). GV “germination value” was mentioned as $GV = PV \times MDG$, where PV: peak value of germination, and MDG: mean daily germination which means the number of seeds that germinate per day (Czabator, 1962). PV stands for the highest cumulative germination percentage divided by the number of days it took to get there. This investigation was approved out by adopting a set of instructions including uncertainty (U) and synchronization index (Z). Seed germination as a synchronized characteristic can be quantified by synchronization index (Z) (Ranal and De Santana, 2006). The value of U depends on one seed germination and means that U determines the distribution degree of germination via time, and the synchronization index determines germination overlapping degree (Ranal and De Santana, 2006; Dorneles, Ranal, and Santana, 2005). When at least two seeds can germinate, one behind the other means $Z=0$ and, $Z= 1$ indicates all seeds germination simultaneously happen. Treatment effects were

determined by analysis of variance according to the general linear model procedure in Statistical Analysis System (SAS Institute, Cary, NC, USA). Mean comparisons were performed with Duncan's multiple range test at the 1% level of significance. Because of *M. armeniacum* seed germination results, analysis of variance and mean comparison data only reported in *M. neglectum*.

3. Results

Stratification, chemical, and hot water scarification were performed on a selected population of *M. neglectum* and *M. armeniacum* (Table 1) to increase germination rate, reduce germination period, and breaking seed dormancy. The variance analysis table showed that sulfuric acid, hot water, and stratification treatments significantly modified the germination characteristic of *Muscari neglectum* seeds (Table 2). Moreover, sulfuric acid and stratification interaction, as well as hot water and stratification interaction, had a significant effect on germination characteristics.

The germination time courses at stratification and the effect of sulfuric acid and hot water scarification on germination aspects for *M. neglectum* (Table 1) are illustrated in Fig. (1-a and b). Seeds of *M. neglectum* optimally germinated by 5 and 15 min sulfuric acid treatment after 45 days cold stratification, while the germination percentage was sharply suppressed after 20min sulfuric acid treatment (0%) (Fig1-a). The final germination percentage for 5 or 15 min sulfuric and 45-day stratification treatment upon transfer to the optimum condition is about 95% (Fig1-a). Almost none of the seeds of *M. neglectum* germinate under control (without stratification) conditions, except those of seeds within 15 min sulfuric acid treatment (30% germination) (Fig. 1-a). The average number of *M. neglectum* seeds germination is reduced 13%, 38%, and 65% by hot water 0, 5, and 15 min respectively plus 45-day stratification, except those of seeds, within 20 min hot water treatment (Fig.

1 1-b) which destroyed the embryo. Therefore 45- day stratification plus 5 or 15 min sulfuric acid can
2 only accelerate the germination percentage, and approximately all seeds were germinated after 15
3 days (Fig. 1-a). Seeds of *M. armeniacum* only germinate (70%) by 15 min sulfuric acid treatment,
4 plus 45-day stratification, while germination percentage was wholly suppressed after hot water
5 treatment with and without stratification (Fig. 1-b); therefore, analysis of variance and mean
6 comparison data only reported in *M. neglectum*.

7 The weighted mean of germination time is used to measure the mean germination time (MGT).
8 MGT significantly decreased under 15, 30, and 45- days stratification respectively (Fig 2 and 3 – a).
9 Our results reveal the MGT decreased 88% under 45-day stratification plus 5 and 15 min sulphuric
10 acid compared to 15-day stratification indicating that 45-day stratification substantially improved
11 germination for *M. neglectum* seeds. On the other hand, the MGR increased significantly compared
12 to control under 45-day stratification (Figure 2 – b). The highest MGR significantly achieved by 5
13 min sulphuric acid and 45-day stratification (Figure 2 – b). Furthermore, 15 min hot water plus 45-
14 day stratification shows the highest MGR, which means there is the shortest mean germination time
15 during these treatments (Figure 3 – a and b).

16 To evaluate the germination value (GV), we report that 5 min sulfuric and 45-day stratification
17 had the highest GV (Figure 2-a), which shows no significant differences among control, 5, and 15
18 min sulfuric acid treatments. Our research indicates that hot water significantly decreases GV,
19 specifically after 15 min hot water treatment (Fig 3-a) plus 15, and 30 days stratification. Moreover,
20 there is no significant difference among control, 5 and 15 min hot water plus 45-day stratification
21 (Fig 3-c). GV increases in 45-day stratification groups compared to control (without stratification)
22 (Fig 2 and 3-c).

To determine germination variability, we evaluate the variation coefficient of the germination time (CVt). The highest CVt was achieved by the interaction of 15 (73%) and 20 (72%) min sulfuric acid and 30-day stratification; respectively therefore higher germination variability can be interpreted from these treatments (Fig. 2-d). Because of low germination in hot water treatments (Fig1-b), CVt was significantly suppressed by all hot water treatments (Fig. 3- d).

The high synchronization index (Z) value shows low variation, and the zero Z index indicates high diversity. Hence, uncertainty (U) correlated by distributing the relative frequency of germination.

Low U values represent frequencies with few peaks indicating more concentrated germination over time. Figure (2 - e and f) display no difference in z index and U among sulfuric acid treatments in different stratification, except 5 min sulfuric acid plus 45-day stratification (0.1) which significantly reveal the highest Z index. This observation indicates that 5 min sulfuric acid plus 45-day stratification has the highest germination overlapping (Fig 2-f). Uncertainty (U) increased in 15-day stratification plus sulfuric acid treatment groups (Fig 2 - e), which implies a wide range of relative germination frequencies and a low Z index. (Fig 2 –e and f). Figure (3 – e) shows U significantly decreased under hot water treatments. Thus, hot water 15 min plus 45-day stratification reveals the highest Z index (0.12), which means more seeds germinated at the same time (Figure 3- f).

4. Discussion

Our results indicate that *M. neglectum* seeds germinate 95 % after 45-day stratification and 5 or 15 min sulfuric acid treatments in optimum temperature (Fig1-a). Seeds of *M. armeniacum* only germinate 69% by 15 min sulfuric acid treatment plus 45-day stratification. Doussi and Thanos (2002) indicated that *Muscari* seed germination took place in a relatively small range of cold temperatures

1 (10 or 15°C) and at an astonishingly slow rate; also, no primary dormancy was reported. Cold
2 stratification and chemical or mechanical scarification seem to have decrease seed hardness and
3 increased low or irregular germination and emergence rates. (Baskin and Baskin, 2014; Dürr *et al.*,
4 2015; Martínez-Díaz *et al.*, 2018). According to Doussi and Thanos (2002), chipping *M. comosum*
5 seeds resulted in only a small improvement in germination rate without primary dormancy; however,
6 our findings explain the 45-day stratification impact *Muscari* embryos' post-maturing growth
7 requirements. Many Liliaceous species have previously been recorded to have tiny, linear, and
8 underdeveloped embryos. (Baskin, C.C. and Baskin, 1998). Our results conclude that 5 min sulfuric
9 acid plus 45-day stratification treatment on *M. neglectum* seed decreases germination time to 15 days
10 in optimum temperature. Seeds of different plants require 4–28 weeks of stratification to break their
11 dormancy, depending on the species (Tang *et al.*, 2019). Stratification plays an important role as a
12 stimulator that helps to break dormancy. The oxygen demand of the embryo is best met at lower
13 temperatures because more oxygen is soluble in water. (Leo, 2013). Hence, by - enzyme activity and
14 generating amino acids needed for embryo use during development, stratification was successful in
15 shortening the seed dormancy time (Saffari *et al.*, 2021) However, there is a report that indicates the
16 chipping without stratification increased the germination rate, and around 100% germination occurred
17 24 days after plantation of *M. neglectum* (Doussi and Thanos, 2002).

18 Our experiment MGT and MGR results indicate that 45-day stratification plus 5 min sulphuric
19 acid remarkably improved the germination quality of *M. neglectum* seeds. The number of seeds which
20 germinated during the data collection periods is used as a weight. (Ranal and De Santana, 2006).
21 Using of the weighted mean is needed in this situation, as it considers that a various number of seeds
22 germinate. GV increases in 45-day stratification plus hot water and sulfuric acid (5 and 15 min)
23 compared to control (without stratification) (Fig 2 and 3-c). This result suggests that GV is a measure

of germination speed and totality, and their interaction, as defined by Brown and Mayer (1988). On the other hand, our data show that 15 and 45 days stratifications significantly decreased CVt, as a consequence, these treatments can be perceived as having a higher germination uniformity.

We found that 5 min sulfuric acid plus 45-day stratification significantly increased synchronization index (Z) (0.1) which has the highest germination overlapping or low uncertainty (U) (Fig 2-e and f). This observation, consistent to this treatments' germination course time in figure (1 - a). We recommended that 5 min sulfuric acid plus 45-day stratification can be used as the most effective and easily applicable pretreatment for the shortest time to complete germination in *M. neglectum* populations. On the other hand, *M. armeniacum* seeds show 70% germination only by 15 min sulfuric acid plus 45-day stratification treatment.

5. Conclusion

Our finding demonstrates that *M. neglectum* populations (Table 1) have significant diversity under stratification and seed priming. Selected populations that were collected from “Golpaygan” and “Isfahan” provinces (Table 1) showed the highest morphological characteristic (Labbaf, Rohollahi and Naji, 2020). We conclude that genetic and environmental aspects can influence morphological details and germination characteristics. (Rohollahi *et al.*, 2015). Our observations indicate essential germination features which help discriminate between excellent and poor treatments. The germination estimation limits were used to help by understanding and interpretation during comparisons.

In summary, our results indicated, *M. armeniacum* seeds only germinate by 15 min sulfuric acid treatment plus 45-day stratification and *M. neglectum* seeds should receive 5 min sulfuric acid plus 45 days stratification for rapid, uniform, and valuable germination. Based on our experiment

1 results, field germination of top morphological Iranian selected population of *M.neglectum* seeds is
2 tuned to take place well in mid-winter and provided a lengthy constant stratification for embryo
3 development and some scarification.

4 **6. Acknowledgements**

5 We gratefully acknowledge the critical comments of Dr. Heshmat Omid, Shahed University.

6 **7. Declaration of competing interest**

7 The authors declare that they have no conflict of interest

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3 Table1. In various provinces of Iran, accessions of *M. neglectum* were collected.

<i>Muscari</i> Species	Country/Province/CS	Latitude	Longitude	Altitude (m)
<i>M. neglectum</i>	IRAN/Isfahan/ Golpaygan A	50° 19' 25"	33° 25' 55"	1851
<i>M. neglectum</i>	IRAN/Isfahan/ Golpaygan B	50° 17' 71"	33° 28' 23"	1823
<i>M. neglectum</i>	IRAN/Isfahan/ Golpaygan C	50° 20' 31"	33° 28' 42"	1811
<i>Muscari</i> Species	Country/Province	Website		
<i>M. armeniacum</i>	United Kingdom's	www.plant-world-seeds.com		

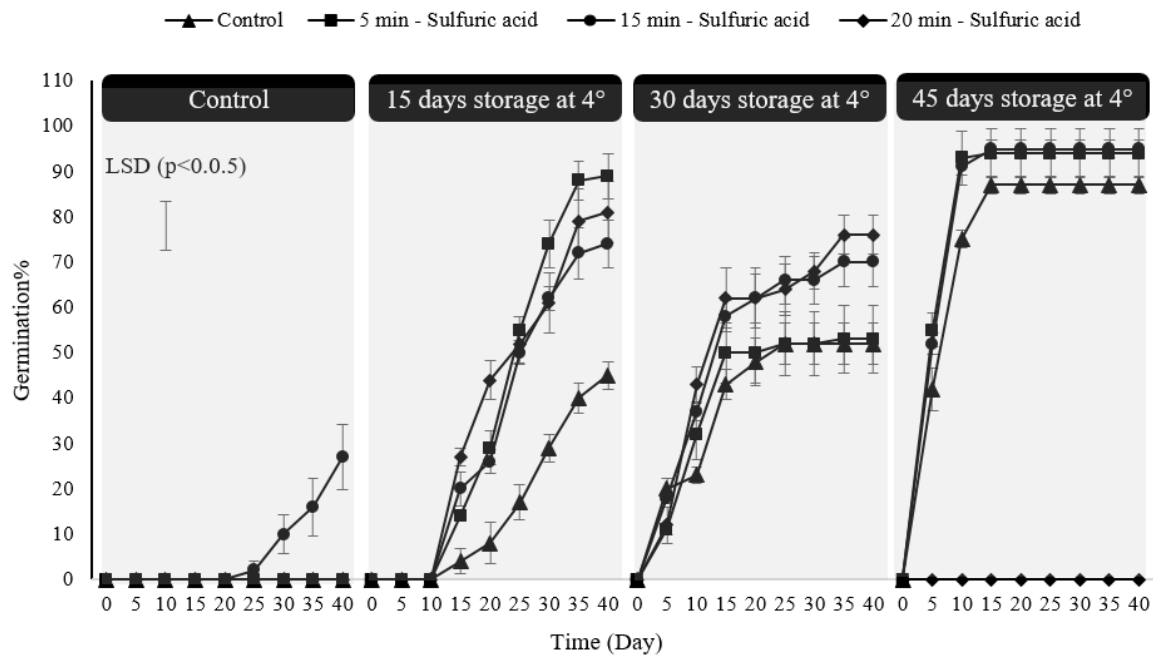
4 CS: Collecting Site

5

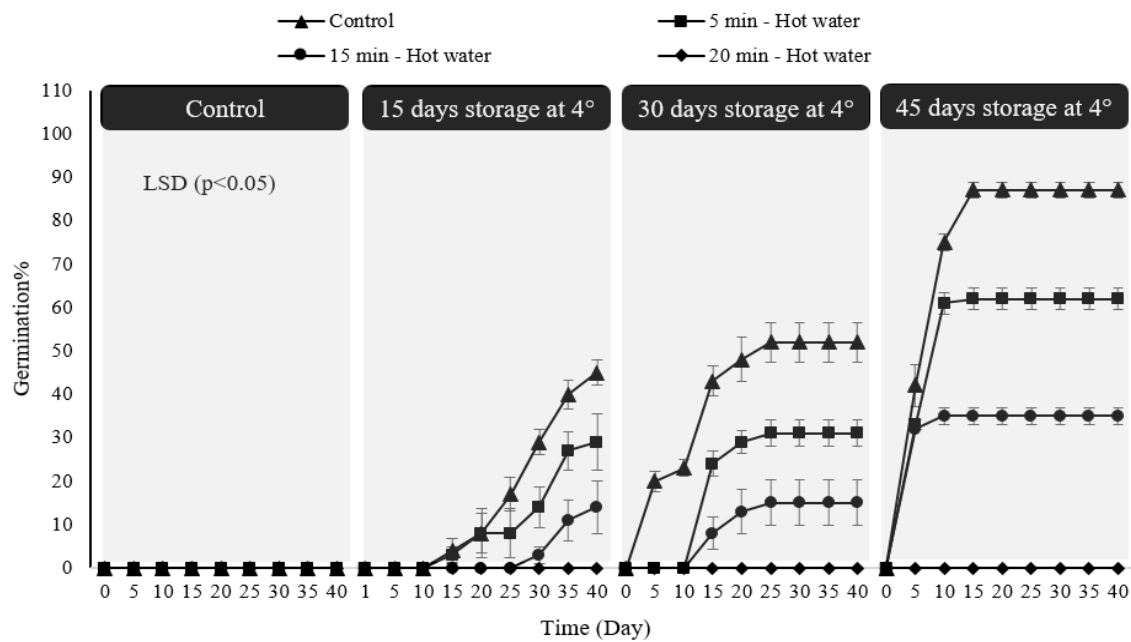
Table1. Variance analysis of Sulfuric acid and hot treatments on germination characteristic of *Muscari neglectum* seeds.

Source of variation	Df	Mean Sq						
		Ge	MGT	MGR	CV	U	Z	GV
Sulfuric-Acid	3	2431.58**	316.27**	0.01**	677.92**	5.04**	0.004**	3.539**
Stratification	3	15254.9**	1101.17**	0.05**	9867.82**	26.22**	0.016**	17.980**
Sulfuric-Acid × Stratification	9	2955.8**	314.55**	0.007**	908.74**	3.90**	0.002**	3.513**
Error	48	80.08	2.70	0.00009	43.71	0.10	0.0003	0.07
CV %	-	16.9	13.9	14	19.5	13.5	3.2	11.7
Hot Water	3	3.90**	1.62**	0.04**	47.62**	0.29**	0.03**	0.014**
Stratification	3	3.70**	2.24**	0.12**	49.36**	0.25**	0.02**	0.050**
Hot Water × Stratification	9	0.47**	0.26**	0.02**	7.73**	0.04**	0.05**	0.006**
Error	48	0.02	0.01	0.0001	0.60	0.003	0.002	0.00007
CV %	-	16.8	15.8	2.3	20.8	12.2	8.1	19.8

Table 2.



(a)



(b)

Fig 1. Time courses of seed germination and final seed germination in chemical scarification plus cold stratification treatment (a) Scarification with hot water and cold stratification, for *Muscari neglectum* (b).

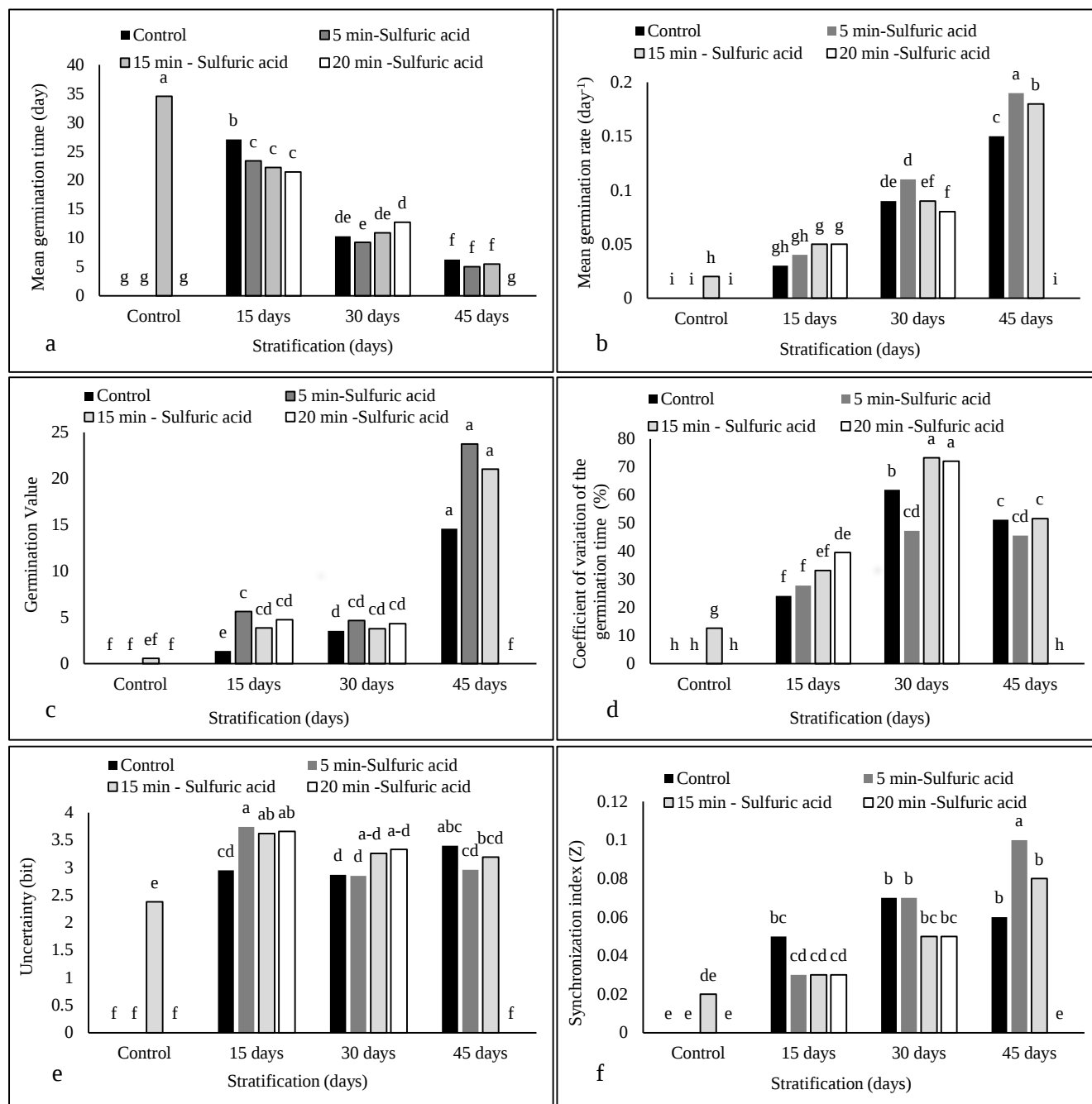


Fig 2. Stratification and sulfuric acid interaction on Mean germination time (MGT) (a) Mean germination rate (MGR) (b) germination value (GV) (c) Coefficient of variation of the germination time (CVt) (d) Uncertainty (U) (e) Synchronization index (Z) (f) in *Muscari neglectum*. Bars with different letters within each preservative and each group are significantly different in the least squares means test.

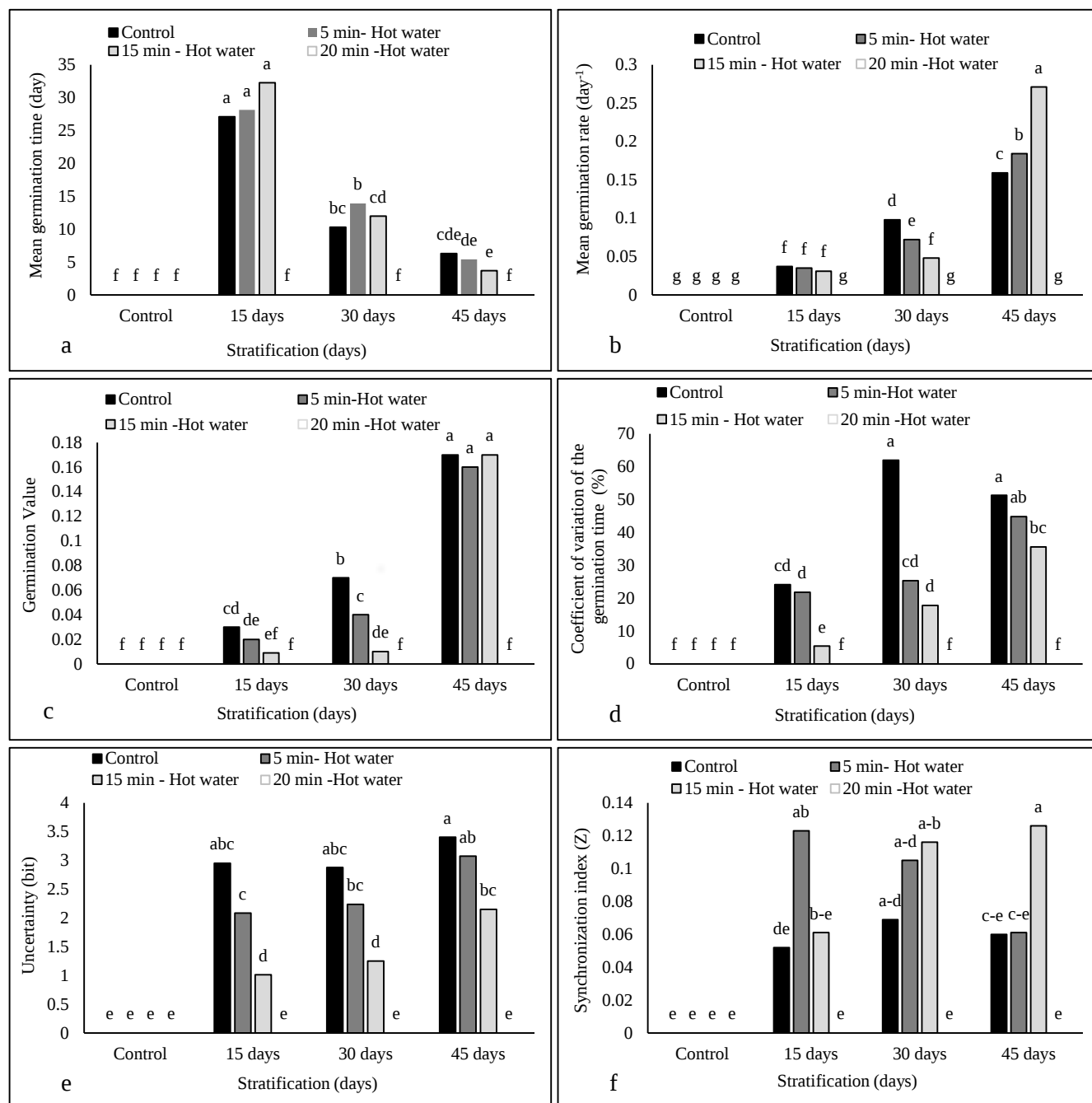


Fig 3. Stratification and hot water interaction on Mean germination time (MGT) (a) Mean germination rate (MGR) (b) germination value (GV) (c) Coefficient of variation of the germination time (CVt) (d) Uncertainty (U) (e) Synchronization index (Z) (f) in *Muscari neglectum*. Bars with different letters within each preservative and each group are significantly different in the least squares means test.

8. Data Accessibility

Sampling locations and germination data of genotypes: Dryad doi: 10.5061/dryad.p5hqbzks5

9. References

Baskin, C.C. and Baskin, J. M. (1998) *No Title Seeds: Ecology, Biogeography, and Evolution of Dormancy and Germination*. San Diego: CA: Academic Press.

Baskin, C. C. and Baskin, J. M. (2014) ‘Seeds: Ecology, Biogeography, and Evolution of Dormancy and Germination’, pp. 1–1586. doi: 10.1016/C2013-0-00597-X.

Bu, H. *et al.* (2008) ‘Community-wide germination strategies in an alpine meadow on the eastern Qinghai-Tibet plateau: Phylogenetic and life-history correlates’, *Plant Ecology*, 195(1), pp. 87–98. doi: 10.1007/s11258-007-9301-1.

Candido, V. *et al.* (2017) ‘Seed-propagated *Muscari comosum* (L.) Mill.: Effects of sowing date and growing conditions’, *Plant Biosystems - An International Journal Dealing with all Aspects of Plant Biology*, 151(3), pp. 484–492. doi: 10.1080/11263504.2016.1194337.

Czabator, F. J. (1962) ‘Germination value: an index combining speed and completeness of pine seed germination’, *Forest science*, 8(4), pp. 386–396.

Davis, P.H. & Stuart, D. . (1984) *Muscari Miller*. In: *davis, P.H. (Ed.) Flora of Turkey and the east Aegean Island*. Edinburgh: Edinburgh University Press.

Dorneles, M. C., Ranal, M. A. and Santana, D. G. (2005) ‘Germinação de diásporos recém-colhidos de *Myracrodruon urundeuva* Allemão (Anacardiaceae) ocorrente no cerrado do Brasil Central’, *Revista Brasileira de Botânica*, 28(2), pp. 399–408. doi: 10.1590/S0100-84042005000200018.

Doussi, M. A. and Thanos, C. A. (2002) ‘Ecophysiology of seed germination in Mediterranean geophytes. 1. *Muscari* spp’, *Seed Science Research*, 12(3), pp. 193–201. doi: 10.1079/ssr2002111.

Dürr, C. *et al.* (2015) ‘Ranges of critical temperature and water potential values for the germination of species worldwide: Contribution to a seed trait database’, *Agricultural and Forest Meteorology*, 200, pp. 222–232. doi: 10.1016/j.agrformet.2014.09.024.

Fawzi Mahomoodally, M. *et al.* (2021) ‘A comprehensive evaluation of the chemical profiles and biological properties of six geophytes from Turkey: Sources of bioactive compounds for novel nutraceuticals’, *Food Research International*, 140(December 2020), p. 110068. doi: 10.1016/j.foodres.2020.110068.

Ghahreman, A. (1984) *Color Atlas of Iranian Plants*. 5th edn. Tehran: Institute of Forestries and Grasslands, Botany Division.

Grey-Wilson C, Mathew B, B. M. (1981) *Bulbs: the bulbous plants of Europe and their allies*. Collins, London. Available at: <https://www.amazon.com/Bulbs-Christopher-Grey-Wilson/dp/000219211X>.

Herrmann, N., Gabriele, W. and Durka walter (2006) ‘20.nick.etal.2006.pdf’, pp. 81–101.

De Hertogh, A. and Le Nard, M. (Eds) (1993) *The Physiology of Flower Bulbs*. Amsterdam, Netherlands.: Elsevier Science.

- 1 ISTA (2017) *International Rules for Seed Testing*,. Bassersdorf, Switzerland.
- 2 Kamenetsky, R. and Okubo, H. (2012) *Ornamental Geophytes*. CRC Press. doi: 10.1201/b12881.
- 3 Labbaf, N., Rohollahi, I. and Naji, A. M. (2020) ‘Genetic diversity and population structure of wild
4 Persian grape hyacinths (*Muscari neglectum* Guss. ex Ten.) assessed by morphological and
5 molecular markers’, *Genetic Resources and Crop Evolution*, 67(6), pp. 1481–1492. doi:
6 10.1007/s10722-020-00922-7.
- 7 Leo, J. (2013) *The Effect of Cold Stratification on Germination in 28 Cultural Relict Plant Species*.
- 8 Lost, P. and Regained, P. (1993) ‘The Physiology of Flower Induction -’, *The physiology of flower
9 bulbs*, pp. 7–20.
- 10 Maguire, J. D. (1962) ‘Speed of germination—Aid in selection and evaluation for seedling
11 emergence and vigor’, *Crop science*, 2(2), pp. 176–177.
- 12 Martínez-Díaz, E. *et al.* (2018) ‘Germination and morpho-phenological traits of *Allium*
13 *melananthum*, a rare species from south-eastern Spain’, *Flora: Morphology, Distribution,
14 Functional Ecology of Plants*, 249(August), pp. 16–23. doi: 10.1016/j.flora.2018.09.005.
- 15 Olmez, Z., Gokturk, A. and Temel, F. (2007) ‘Effects of cold stratification, sulphuric acid,
16 submersion in hot and tap water pretreatments on germination of bladder-senna (*Colutea armena*
17 Boiss. & Huet.) seeds’, *Seed Science and Technology*, 35(2), pp. 266–271. doi:
18 10.15258/sst.2007.35.2.02.
- 19 Pehlivan, S. and Özler, H. (2003) ‘Pollen morphology of some species of *Muscari* Miller
20 (*Liliaceae-Hyacinthaceae*) from Turkey’, *Flora*, pp. 200–210. doi: 10.1078/0367-2530-00092.
- 21 Ranal, M. A. and De Santana, D. G. (2006) ‘How and why to measure the germination process?’,
22 *Revista Brasileira de Botanica*, 29(1), pp. 1–11. doi: 10.1590/S0100-84042006000100002.
- 23 Rohollahi, I. *et al.* (2015) ‘Evaluation of seedling emergence and relative DNA content under dry
24 soil conditions of wild *Festuca arundinacea* populations collected in Iran’, *Grassland Science*,
25 61(1), pp. 6–14. doi: 10.1111/grs.12074.
- 26 Saffari, P. *et al.* (2021) ‘Effect of treatments on seed dormancy breaking, seedling growth, and
27 seedling antioxidant potential of *Agrimonia eupatoria* L.’, *Journal of Applied Research on
28 Medicinal and Aromatic Plants*, 20(October), p. 100282. doi: 10.1016/j.jarmap.2020.100282.
- 29 Sohindji, F. S. *et al.* (2020) ‘Understanding molecular mechanisms of seed dormancy for improved
30 germination in traditional leafy vegetables: An overview’, *Agronomy*, 10(1). doi:
31 10.3390/agronomy10010057.
- 32 Tang, Y. *et al.* (2019) ‘Dormancy-breaking and germination requirements for seeds of *Sorbus*
33 *alnifolia* (Siebold & Zucc.) K.Koch (*Rosaceae*), a mesic forest tree with high ornamental potential’,
34 *Forests*, 10(4). doi: 10.3390/f10040319.

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