

DC-T cell axis is an effective target for treating non-small cell lung cancer

Shuangcui Wang¹, Guan Zhang¹, qian cui¹, Yanjie Yang¹, Dong Wang¹, Aqing Liu¹, Ying Xia¹, Wentao Li¹, Yunhe Liu¹, and Jianchun Yu¹

¹First Teaching Hospital of Tianjin University of Traditional Chinese Medicine

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Abstract

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Shuangcui Wang^{1,2}, Guan Zhang^{1,2}, Qian Cui^{1,2}, Yanjie Yang^{1,2}, Dong Wang^{1,2}, Aqing Liu^{1,2}, Ying Xia^{1,2}, Wentao Li¹, Yunhe Liu¹, Jianchun Yu^{1*}

1. Department of Oncology, First Teaching Hospital of Tianjin University of Traditional Chinese Medicine, Tianjin, China

2. National Clinical Research Center for Chinese Medicine Acupuncture and Moxibustion, Tianjin, China

Shuangcui Wang, E-mail:2778761272@qq.com

Guan Zhang, E-mail:1483950415@qq.com

Qian Cui, E-mail:cuiqian1025@163.com

Yanjie Yang, E-mail:1026790005@qq.com

Dong Wang, E-mail: 1632016659@qq.com

Aqing Liu, E-mail:8liuxiangyu8@163.com

Ying Xia, E-mail:1151364753@qq.com

Wentao Li, E-mail:v23liwentao@163.com

Yunhe Liu, E-mail:yunhe654321@126.com

Jianchun Yu, E-mail:yujianchun2000@163.com

Corresponding authors: Jianchun Yu

Correspondence: Jianchun Yu, Department of Oncology, First Teaching Hospital of Tianjin University of Traditional Chinese Medicine, Tianjin University of Traditional Chinese Medicine, Tianjin, China.

*Corresponding author.

Abstract: The DC-T cell axis is a bridge connecting innate immunity and adaptive immunity. The initial immune response against tumors is mainly induced by mature antigen-presenting dendritic cells (DC). Enhancing the crosstalk between DC and T cells could improve the immune response to non-small cell lung cancer. This article reviews the interaction between DC-T cells in the treatment of non-small cell lung cancer and how this interaction impacts the treatment outcome.

Keywords: non-small cell lung cancer; dendritic cells; T cells; Artificial DC; Chimeric Antigen Receptor T-Cell Immunotherapy.

Preface:

Lung cancer is a common cause of cancer-related death worldwide^[1-4], and the incidence and mortality rate of lung cancer are projected to rise in the next few decades. Therefore, lung cancer remains a public health concern^[5]. Histopathologically, lung cancer is broadly classified into two types: small cell lung cancer (SCLC) and non-small cell lung cancer (NSCLC). The NSCLC accounts for about 85% of lung cancer cases, adenocarcinoma, squamous cell carcinoma, and large cell carcinoma are the main histological subtypes. NSCLC is the most common histologic type, with a 5-year overall survival rate (OS) of approximately 15%^[6-8]. Therefore, it is particularly important to identify more critical factors influencing efficacy of NSCLC treatment.

Recently, immunotherapy based on immune checkpoint inhibitors (ICIs) alone or in combination with chemotherapy has greatly improved cancer treatment outcomes. However, only a few patients (15%) respond to ICIs, which has generated considerable urgency in identifying immune factors affecting its efficacy^[9-11].

Recent advances in understanding tumor immunity have led to the rapid development of cell-based immunotherapy strategies targeting antigen-presenting cells (APC), T cells, and tumor cells. Among them, the DC-T cell axis has been the focus of recent aims for developing anticancer therapies. DC can mediate the initiation of adaptive immune responses of T cells and directly kill tumor cells^[12-13]. The DC-T cell axis is a bridge connecting innate immunity and adaptive immunity, playing a pivotal role in the development of specific immunity. Therefore, focusing on the DC-T cell axis provides a comprehensive understanding of the interaction of DC and T cell in the treatment of NSCLC. Additionally, the work explores new therapeutic opportunities to overcome the immune tolerance of NSCLC and improve the prognosis of NSCLC patients.

1. Crosstalk between DC and T cells

1.1 T cells are the key to the occurrence and development of tumor immune response

T cells derived from lymphoid stem cells in bone marrow and undergo differentiation and maturation in the thymus, and then distributed to immune organs and tissues throughout the body through lymphatic and blood circulation. T cells mediate cellular immunity and coordinate the whole immune response^[14-15]. CD4⁺ or CD8⁺ T cells are the two most important types of T cells^[16-17] studied in the development of adaptive immunity. The most important feature of adaptive immunity is characterized by specificity via stimulation of specific antigens from tumor or pathogens. The CD4⁺naive T cells (CD4⁺ T_n) can develop and differentiate into CD4⁺ T cells under stimulation by antigens presented by DCs. Of them, Th1, Th2, Th17, and regulatory T cells (Treg) have been extensively studied^[18-20]. Th1 mainly secretes IL-2, IL-12, IFN- γ , and TNF- α , promoting the proliferation of Th1, and plays critical roles in the cellular immunity. Furthermore, these cytokines inhibit the proliferation of Th2^[21-22]. IFN- γ activates and enhances the phagocytotic property of macrophages and promotes the production of IgG^[23-24]. IL-2, IFN- γ and IL-12 can enhance the killing ability of NK cells^[25]. IL-2 and IFN- γ synergistically stimulate the proliferation and differentiation of cytotoxic T cells (CTL). On the other hand, TNF directly induces apoptosis of target cells and also promotes inflammatory response^[26-27]. Th2 mainly secretes IL-4, IL-5, IL-6, IL-10, and IL-13, which promotes the proliferation of Th2 cells and participates in the activation of B cells. In addition, these cytokines induce humoral immunity and inhibit the proliferation of Th1^[28-30]. Th17 induces innate immunity and inflammation by secretion of IL-17 (IL-17A to IL-17F), IL-21, IL-22, IL-26, TNF- α , and

other cytokines. Moreover, Th17 induces the occurrence and development of immunopathological damage, especially autoimmune diseases [31-33]. Treg cells exert the immunosuppression by inhibiting the activation and proliferation of CD4⁺ and CD8⁺ T cells and function of DC [34], negatively regulating the immune response. The act is performed mainly in two ways: Inhibiting the ability of the APC to activate the T cells mainly through the molecules CTLA-4, LAG-3 on the surface of Treg to interact with the CD80, CD86, and MHC on the DC surface, respectively; Suppressing effector T cells by secreting TGF- β and IL-10 and IL-35 [34]. Generally, cytokines secreted by CD4⁺ T cells regulate the function of immune cells, monitor the development, differentiation, and function of other immune cells such as CD8⁺ T cells, B cells, NK cells, DC. Therefore, CD4⁺ T cells play a key and central role in the immune system. Naïve CD8⁺ T cells (CD8⁺ Tn) can develop and differentiate into CD8⁺ T cells upon stimulation by antigens presented by DC. Consequently, CD8⁺ T cells can differentiate into CTL that kill pathogen-infected cells and tumor cells through cytolytic molecules, such as perforin and granzyme, and are essential for a cell-mediated antitumor immune response [35-37]. Alternatively, CTLs induce tumor cell apoptosis by promoting the binding of FasL to Fas on the surface of tumor cells [38-39] (Fig. 1).

1.2 DC is the initial factor inducing adaptive tumor immunity

DCs were first reported in skin by Langerhans in 1868. Steinman and Cohn later described DC in peripheral lymphoid organs in 1973. DCs form an essential link between innate immunity and adaptive immunity. More importantly, DCs induce the occurrence of adaptive immunity [40].

DC represents a complex heterogeneous population of antigen-presenting cell with significant phenotypic heterogeneity and functional plasticity. DCs can be divided into two main subpopulations: conventional DC (cDC) and plasmacytoid DC (pDC). DCs are derived from bone marrow and migrate through blood circulation to lymphoid tissues such as lymph nodes and spleen and non-lymphoid tissues such as skin, lung, and intestine [41]. cDCs are further subdivided into type 1 cDC and type 2 cDC [42-43]. cDC1 can cross-present antigens and trigger an antitumor immune response and, thus, often referred to as cross-presenting DCs. cDC1 presents tumor antigen to CD8⁺ Tn cells via the MHCI, activating the initial immune response [44-45]. After activating CD8⁺ Tn, CD8⁺ Tn gradually develop and differentiate into immune memory cells, such as Tscm, Tcm, Tem, and Tte, inducing the occurrence of recall response by re-stimulation of past antigen encounters. The quick responses to the secondary stimulation of antigen form a large number of specific immune cells in a short time and effectively kill tumor cells [46-47]. It is worth noting that among several antigen-presenting cells, only DC can stimulate the initial immune response and induce Tn to produce an immune response targeting “new” antigens. It is speculated that the damage of DC is closely related to refractory tumors. cDC1 plays a key role in lung immunity [48]. On the other hand, cDC2 is a major DC subpopulation in different human tissues and organs. Besides, cDC2 presents tumor antigens to CD4⁺ Tn cells via the MHCII and activates the initial immune response of CD4⁺ Tn. Like CD8⁺ Tn, after activation, CD4⁺ Tn also gradually develop and differentiate into immune memory cells such as Tscm, Tcm, Tem, and Tte, forming a memory immune response to coordinate the immune response and homeostasis [49]. In addition, pDC are key players in antiviral immunity. The pDCs can produce high levels of type I interferon (IFN), promoting innate and adaptive immunity [50] (Figure 1). The IFN is considered the key natural immune defense of the body against pathogen infection. Furthermore, IFN inhibits the proliferation of tumor cells and regulates immunity [51-52].

1.3 DC interacts with T cells at the immune synapse

The activation of T cells by DC is mainly through the interaction between DC and T cells, and the formation of immune synapses is the most important feature of this process. The immune synapse is a special structure formed at the cell-cell contact interface when APC and T cells interact [53-54]. In this process, DC provides three key signals.

The initial signal is generated when T cells recognize homologous peptide antigens presented on MHC I or MHC II on the surface of DC through their T cell receptor (TCR) [55-58]. MHC I molecules bind to CD8⁺ Tn cells, and MHC II molecules bind to CD4⁺ Tn cells to present specific antigens to the Tn [59].

MHC, antigen and TCR interaction triggers the activation of T cells, initiating downstream signals via the immune receptor tyrosine-based activation motif (ITAM)^[60]. Subsequently, in addition to the MHC-antigen-TCR complex, a second costimulatory signal is required to initiate and maintain T cell activation and proliferation. The second signal can be divided into costimulatory molecules and co inhibitory molecules according to different effects^[61-62]. The key costimulatory molecules involved in T cell activation include CD28 (binding to CD80/86 on DC), ICOS (binding to icosl on DC), OX40 (binding to OX40L on DC), and CD40L (binding to CD40 on DC), all the key signaling molecules are responsible for T cell activation, differentiation, and survival. The costimulatory signals cooperate with MHC-antigen -TCR complexes to enhance T cell activation. For instance, the costimulatory molecule CD28 is an important signaling molecule that determines the quality and quantity of T cell immune response^[63-64]. In addition, after CD28 activation, PI3K/Akt and other signals are induced to promote the proliferation and development of Tn. Consequently, Tn-Tscm-Tcm-Tem-Tte forms the initial immune response and memory immune response, enabling the body to respond to both the “old” antigens generated by tumor recurrence and the “new” antigens generated by tumor variation^[65-66]. After CD28 activating, followed by activating PI3K/Akt^[67], NF- κ B^[68] and other signaling pathway, Tn and Tm then complete their development, differentiation, and proliferation, forming effector T cells to kill tumors and other pathogens. Contrarily, T lymphocyte-associated protein 4 (CTLA-4) competes with CD28 for binding to CD80/86 on DC while PD-1 binds to programmed cell death ligand 1 (PD-L1) on the surface of DC, inhibiting the activation signals^[69-70]. Positive and negative costimulatory molecules interact to effectively start, moderately effect, and timely terminate the immune response^[71]. The third signal in the form of cytokines secreted by DC is triggered once T cells receive MHC antigen TCR complex signals and sufficient costimulation^[72-73]. Combined with T cells, DC can secrete plenty of IL-12 and IL-18. In addition, DCs are dominant partners of T cells and necessary for activating the proliferation of T cells, inducing CTL production, dominating priming of Th1 type immune response, and facilitating tumor clearance^[74-76].

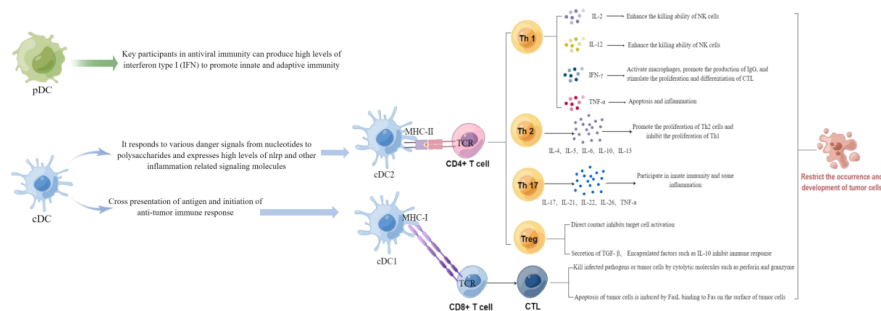


Fig 1: The schema chart of a complex interaction between DC, T cells, and tumor cells

Note: DC promotes immunity by presenting antigens to T cells and providing immunomodulatory signals through cell-cell contact and cytokines. DC can recognize, ingest and process tumor antigens. After processing and presentation, antigens are presented to CD4⁺ T cells or CD8⁺ T cells via MHC class I or class II molecules on the surface of DC, initiating T cell activation. Mature DC highly expresses costimulatory molecules such as CD80, CD86, and CD40, providing a second signal for the full activation of T cells. DCs secrete several cytokines, including IL-2/IL-12/IFN- γ , which further induce the proliferation and differentiation of activated T cells to complete the initiation of the immune response. Unlike activating memory T cells, the activation of naive T cells is more dependent on the presence of DC stimulation signals. Therefore, DC is the most potent mature APC, unique in its ability to prime naive T cells.

2. Function of DC and T cells in NSCLC

2.1T cells affect prognosis and efficacy of immunotherapy in NSCLC patients

The number of circulating lymphocytes, including the proportion of and the absolute number of lymphocytes, is an important indicator of the efficacy of the body's immune function and the degree of stress^[77]. Interaction

and influence each other exist between the various cells in the immune system, the appropriate ratio for immune cells in the body is an important guarantee for their function. Peterson et al explored the prognostic relevance of Treg cells in 64 patients with stage NSCLC and found that a higher proportion of Treg cells in the tumor significantly increased the risk of recurrence. Immunohistochemistry has detected Treg cells in 51% of NSCLC patients. Therefore, Treg cells can be identified as an important risk factor for postoperative recurrence of NSCLC^[78]. In addition, Liu et al reported that a high Foxp3⁺Treg/CD8⁺ T cell ratio predicted poor response to platinum-based chemotherapy in advanced NSCLC. Given this apparent functional relevance of Treg cells in NSCLC, targeting Treg cells might be an interesting approach for immunotherapy of lung cancer^[79].

Clinically, the absolute number of lymphocytes is much more important than the proportion. Tumors can significantly inhibit the proliferation of bone marrow cells during the developmental period, significantly decreasing the absolute number of various immune cells, including lymphocytes^[80-81]. At the same time, one of the main consequences of chemotherapy and radiotherapy is damaging the bone marrow^[82]. As a result, it decreases the absolute number of lymphocytes that is closely related to the prognosis and efficacy of immunotherapy^[83-84]. Numerous studies have shown that low lymphocyte absolute count (ALC) is an independent negative prognostic factor for advanced malignant tumors^[85]. Oh et al found that the baseline absolute lymphocyte count level is an independent prognostic factor of rectal adenocarcinoma patients receiving preoperative radiotherapy and chemotherapy in different treatment periods and follow-up periods. In one study, the absolute lymphocyte count decreased after preoperative radiotherapy and chemotherapy, reaching the lowest level at 1 month but gradually increased after the end of radiotherapy and chemotherapy^[86]. Furthermore, Xia Ying et al. found that the absolute numbers but not the relative numbers of lymphocyte subsets CD3⁺, CD4⁺, CD8⁺, B, and NK cells were significantly damaged in NSCLC patients, and were closely related to the disease progression and progression-free survival period. And they were served as biomarkers for disease prognosis and efficacy. The work indicated that only paying attention to the proportion of lymphocyte subsets in clinic is easy to lead to clinical misjudgment^[87].

T cells can also effectively reduce NSCLC by increasing their activity. IFN- γ promotes the activity of T cells by inducing the presentation of antigen-presenting cells (APCs) and upregulating the expression of MHC I / peptide complex on APCs to reactivate and induce the proliferation of antigen-specific CD8⁺ T cells. The activated tumor antigen-specific CD8⁺ T cells can recognize and secrete cytotoxic molecules to kill tumor cells^[88]. Wu et al showed that Fuzheng anticancer formula effectively enhances the secretion of IFN- γ by CD8⁺ T cells, which is critical in clearing tumor cells^[89]. CTLA-4 and PD-L/PD-IL are the most studied checkpoint pathways, and they inhibit T cell activity in different ways^[90-91]. CTLA-4 regulates T cell proliferation early in immune response^[92], whereas PD-1 suppresses T cells later in immune response^[93]. The immune checkpoint inhibitors including ipilimumab and tremelimumab (CTLA-4 monoclonal antibody), nivolumab and pembrolizumab (PD-1 monoclonal antibody), atezolizumab and durvalumab (PD-L1 monoclonal antibody) have opened a new era in the treatment of advanced NSCLC. Nivolumab and pembrolizumab have been approved by FDA and EMA for the second-line treatment of NSCLC^[94-95].

2.2 DC plays an important role in NSCLC

As a professional antigen-presenting cell, DC plays a vital role in NSCLC prognosis^[96]. The proportion of DCs positively correlates with the progression-free survival of NSCLC. The expression of certain genes, such as TOP2A and TLR3, is related to DC infiltration in NSCLC. Based on the recent findings related to gene expression characteristics and DC infiltration in NSCLC patients, effective strategies for patients with refractory cancer can be designed^[97]. The plasmacytoid dendritic cells (pDCs) and myeloid dendritic cells (mDC) are the main DCs in NSCLC^[98]. Zahran et al demonstrated the possible impact of pDC and mDC on the treatment outcome of NSCLC. Zahran et al reported that higher levels of pDCs in the peripheral blood of NSCLC patients were related to a good NSCLC prognosis, lower tumor stage and longer mean overall survival time (OS). However, mDCs are the opposite^[99-100]. Bianchi et al. reported that the overall survival positively correlated with the proportion of TLR3-CD1-3⁺ dendritic cells and the corresponding activation

of CD8⁺T cells ^[101]. The maturation rate of DC is significantly lower in healthy individuals than in NSCLC patients. The main manifestations of poor tumor prognosis are as follows: lack of functional DCs in the lung tumor lesions; recruitment of pDC to the surrounding lung tumor tissues; Lung tumor-induced regulatory DC; underexpression of DC effector molecules in the lung tumors; secretion of immunosuppressive molecules by infiltrating DCs in the Lung cancer tissue ^[102].

3. Development trend of clinical immunotherapy based on DC and T cells

Currently, immune checkpoint inhibitors have been widely used in clinical cancer treatments and are an important treatment for advanced NSCLC. CAR-T has emerged as a genuine modality for treating blood tumors. However, their low efficacy in treating solid tumors limits their application. Although it has been reported, it is still in the exploratory stage ^[103-104]. In combination with the clinical application of immunotherapy and advances made in understanding immune cells, especially DC, the trend of next-generation immunotherapy is moving towards developing artificial DC and CAR-T that can be used to treat solid tumors. Both can override the limitation of low immunity in tumor patients.

3.1 Engineered DC is the latest treatment strategy for NSCLC

Artificial APCs (aAPCs), which can be readily prepared from “off-the-shelf” components, are promising alternatives to custom-made autologous APCs that can effectively stimulate antigen-specific T cells in vitro ^[105-106]. Several aAPCs for in vitro activation and expansion of antigen-specific T cells have been successfully developed ^[107]. For example, cell-based aAPCs have been studied in several different cell lines, including fly black stomach cells ^[108], NIH3T3 mouse fibroblasts ^[109], and K562 human erythroid leukemia cells ^[110]. These cells were generated using retroviral or lentiviral transduction to introduce molecules that provide TCR, costimulatory, and adhesion molecules necessary for synaptic formation. Given that these cells can be stored for a long time, they can be easily obtained from source companies and distributors^[111]. Although cellular aAPCs can induce a high expansion rate of CD8⁺ T cells, their performance of specific control on the expression level of T cell activation signals is limited. In addition, non-tumor antigens and other stimulatory or inhibitory molecules may be present in cellular aAPCs.

The cell-free aAPCs have been developed to better define the transmission of different signals and avoid the use of allogeneic cells^[112]. Compared to cellular aAPCs, acellular aAPCs allow for more stringent control over the signals delivered and are attractive tools because of their relatively easy preparation through micro latex, polyethylene glycol, magnetic beads, and lipid-based vesicles ^[113]. In general, the aAPCs approach has focussed on the induction of CD8⁺ CTLs through MHC I stimulation because these cells are capable of antigen-specific tumor cell lysis ^[114-115]. Other immune cells, such as CD4⁺ T helper cells, mediate anticancer immune responses by activating CTLs. Artificial APCs comprising various sizes, shapes, surface ligand distribution, and ligand mobility have been developed, and these properties affect the level of T cell activation. Various cell-free aAPCs structures reflect different attempts to simulate different aspects of natural DC ^[112].

It has been shown that DCs usually exhibit poor maturity in the tumor microenvironment and are less effective in presenting tumor antigens^[116]. Therefore, targeting delivery of antigens and adjuvants to cells in vivo is an important method for developing DC vaccines. Sun et al developed intelligent artificial DC cells (iDCs) comprising nanoparticles loaded with a photothermal agent (IR-797) and coated with mature DC cell membranes. The DC cell membrane on the surface of iDCs preserves the ability to present antigens and prime T cells. The iDCs injected into mice enter lymph nodes and stimulate T cell activation and proliferation. Cancer-specific T cells are activated in a TCR-dependent manner and kill tumor cells upon TCR binding of antigens presented via MHCs. Alternately, activated T cells secrete cytokines (TNF- α), reducing the expression of heat shock proteins (HSPs) in tumor cells, thus, enhancing the sensitivity of tumor cells to heat stress. Subsequently, mild photothermal treatment can kill the remaining tumor cells (42-45°C). At the same time, low temperature photoheat can also induce the death of immunogenic cells, thereby activating the body's own DC cells and restarting the tumor immune cycle. As a new precise antitumor nanosystem, iDCs combine the advantages of DC cell immunotherapy and photothermal therapy. Consequently, iDCs effectively

enhance the antitumor immune response of the body, improve the efficacy of tumor treatment, and provide a new strategy for immunosensitized low-temperature photothermal therapy^[117]. Concurrently, Suarez et al developed an Artificial Immune Modulation nanoparticle (AIM-np) technology, a customizable “off-the-shelf” technology that can be used for synthesizing APCs to guide antigen-specific natural CD8⁺ T cells. AIM-np provides a controlled method for antigen presentation and T cell costimulation by directly binding to antigen-specific T cells. This method uses proprietary nanoparticles combined with a proprietary manufacturing process to enrich and expand antigen-specific CD8⁺ T-cell products with consistent purity, identity, and composition required for effective and durable anti-tumor response. AIM-np consists of superparamagnetic iron oxide nanoparticles as the main structure, which is decorated with two humanized signal proteins. HLA-A2-IgG4 hinge dimer molecules are conjugated with core nanoparticles to deliver signal 1 (antigen presentation). In addition, together with humanized anti-CD28 antibodies, the conjugated HLA-A2-IgG4 hinge dimer molecules deliver signal 2 (costimulation). Subsequently, AIM-np acts as a synthetic APC, directly engaging target T cells through naturally occurring signaling mechanisms. Signal 1 is transmitted by a peptide-loaded HLA class I dimerization fusion protein that presents antigenic peptides to cognate T cell receptors. On the other hand, signal 2 is delivered by monoclonal antibodies against CD28 receptors, which deliver costimulatory signals, also known as “danger signals”, to induce antigen-specific T cell activation and proliferation^[118].

The loss of cell surface functional “arms” during DC antigen presentation weakens the interaction between DCs and T cells, disrupting T cell induction^[119]. Modification of DCs to enhance their antigen presentation is the most common strategy for T cell activation. “Engineering” or modifying the cell surfaces with synthetic ligands or receptors provides a novel strategy for regulating the interaction between different cell types^[120-121]. Lectins are carbohydrate-binding proteins that play an important role in promoting intercellular recognition and adhesion due to their specificity and bond stability^[122]. A recent study reported that mannose-modified tumor antigens greatly enhance the ability of DCs to recognize and bind antigens^[123]. Therefore, researchers believe that DCs engineered with glycopolymers can specifically attach to T cells through carbohydrate lectin binding, enhancing the stability of DC-T cell binding while promoting T cell activation. Therefore, adding appropriate synthetic glycopolymers to the cell surface is a valuable and effective way to design and optimize cell vaccines^[124].

3.2 Chimeric antigen receptor T cells (CAR-T) therapy is another new strategy for the treatment of NSCLC

CAR-T cells are genetically engineered T cells that express synthetic CAR vectors. On the basis of self proliferation, CAR-T cells specifically recognize and bind antigens (such as CD19) on tumor cells and specifically kill tumor cells^[125-126]. Since 2017, Kymriah (CTL019) and Yescarta (KTE-C19) CAR-T therapies have successively been approved for use. Furthermore, CAR-T therapy has emerged as a novel treatment strategy with promising results against blood tumors^[127]. However, when the research further turned its attention to solid tumors, which account for 90% of all tumors, CAR-T therapy did not get satisfactory results. The particularity of solid tumors themselves and their microenvironment brought great challenges to CAR-T cell therapy^[128].

Unlike CD19, tumor cells in solid tumors generally express multiple targets abnormally, and these abnormally expressed antigens are also expressed in normal tissues. For example, the current research targets for glioma include prostate-specific antigen (PSMA), carcinoembryonic antigen (CEA), human epidermal growth factor receptor 2 (HER2), epithelial cell adhesion molecule (EpCAM), and Mesothelin^[129-130]. Cancer-associated fibroblasts (CAFs) are one of the most abundant and critical components in the tumor microenvironment^[131], constituting the tumor stromal layer that releases certain inhibitory cytokines. Immunosuppressive cells such as Treg cells, bone marrow-derived suppressive cells, and M2 type macrophages secrete TGF- β , IL-10, and other cytokines to negatively regulate CAR-T cell immune response. In addition, solid tumor cells lose cytokine receptors and escape immune cell surveillance. CAR-T cells cannot effectively respond to chemotaxis secreted by tumor cells, which inhibits their homing ability. However, the high expression of immunosuppressive receptors in solid tumors inhibits the effective activation of CAR-T cells and lowers the

efficacy of CAR T-cell therapy ^[132-134].

As a new strategy for treating NSCLC, CAR-T therapy has made considerable breakthroughs and entered a rapid development stage^[135-136]. Thus far, more CAR-T studies have focused primarily on NSCLC ^[137-138]. The most common target antigens of NSCLC include human epidermal growth factor receptor (EGFR), mesothelin, mucin 1 (MUC1), PD-L1^[139], carcinoembryonic antigen (CEA)^[140], and HER2 ^[141]. Studies have shown that T cells redirected to EphA2 by EphA2-specific CAR have effective antitumor activity against NSCLC in vitro and in vivo. Thus, these antigens are potentially novel targets for NSCLC treatment ^[142].

4. Conclusions and future prospects

Immunotherapy for NSCLC based on DC and T cells is still in the development stage, and it has been clinically proven to be effective against certain cancers. Recent studies have shown that the interaction between the DC and T cells is an emerging strategy for treating NSCLC. Determining the role of DC-T cell interaction in NSCLC not only helps to reveal the mechanism of NSCLC but also provides important clues for treating this cancer. At the same time, based on understanding the interaction between DC and T cells, engineered DC and CAR-T are of great potential for treating solid tumors.

AUTHORS' CONTRIBUTIONS

All authors contributed equally. Shuangcui Wang designed the review, prepared the figure, and wrote the manuscript. Guan Zhang and Qian Cui were involved in the conception and design of the study. Yanjie Yang and Dong Wang searched the literature. Aqing Liu and Ying Xia were involved in the conception and design of the study and revised the manuscript. Wentao Li, Jing Zhang and Yunhe Liu provided helpful comments. Jianchun Yu revised the manuscript. All authors read and approved the final manuscript.

Disclosure statement

No potential conflict of interest was reported by the authors.

References

- [1]Remark R, Becker C, Gomez JE, Damotte D, Dieu-Nosjean MC, Sautès-Fridman C, Fridman WH, Powell CA, Altorki NK, Merad M, Gnjatic S. The non-small cell lung cancer immune contexture. A major determinant of tumor characteristics and patient outcome. *Am J Respir Crit Care Med*. 2015 Feb 15;191(4):377-90. doi: 10.1164/rccm.201409-1671PP. PMID: 25369536; PMCID: PMC5447326.
- [2]Liang G, Meng W, Huang X, et al. miR-196b-5p-mediated downregulation of TSPAN12 and GATA6 promotes tumor progression in non-small cell lung cancer. *Proc Natl Acad Sci U S A*. 2020;117(8):4347-4357. doi:10.1073/pnas.1917531117
- [3]Liu JC, Narva S, Zhou K, Zhang W. A Review on the Antitumor Activity of Various Nitrogenous-based Heterocyclic Compounds as NSCLC Inhibitors. *Mini Rev Med Chem*. 2019;19(18):1517-1530. doi:10.2174/1389557519666190312152358
- [4]Grootjans W, de Geus-Oei LF, Troost EG, Visser EP, Oyen WJ, Bussink J. PET in the management of locally advanced and metastatic NSCLC. *Nat Rev Clin Oncol*. 2015;12(7):395-407. doi:10.1038/nrclinonc.2015.75
- [5]Socinski MA, Obasaju C, Gandara D, et al. Clinicopathologic Features of Advanced Squamous NSCLC. *J Thorac Oncol*. 2016;11(9):1411-1422. doi:10.1016/j.jtho.2016.05.024
- [6]Chae YK, Chang S, Ko T, et al. Epithelial-mesenchymal transition (EMT) signature is inversely associated with T-cell infiltration in non-small cell lung cancer (NSCLC). *Sci Rep*. 2018;8(1):2918. Published 2018 Feb 13. doi:10.1038/s41598-018-21061-1
- [7]Ettinger DS, Wood DE, Aisner DL, et al. Non-Small Cell Lung Cancer, Version 5.2017, NCCN Clinical Practice Guidelines in Oncology. *J Natl Compr Canc Netw*. 2017;15(4):504-535. doi:10.6004/jnccn.2017.0050

- [8]Stewart DJ. Wnt signaling pathway in non-small cell lung cancer. *J Natl Cancer Inst.* 2014;106(1):djt356. doi:10.1093/jnci/djt356
- [9]Chen Z, Fillmore CM, Hammerman PS, Kim CF, Wong KK. Non-small-cell lung cancers: a heterogeneous set of diseases [published correction appears in *Nat Rev Cancer.* 2015 Apr;15(4):247]. *Nat Rev Cancer.* 2014;14(8):535-546. doi:10.1038/nrc3775
- [10]Naylor EC, Desani JK, Chung PK. Targeted Therapy and Immunotherapy for Lung Cancer. *Surg Oncol Clin N Am.* 2016;25(3):601-609. doi:10.1016/j.soc.2016.02.011
- [11]Zhao Y, Qiao G, Wang X, Song Y, Zhou X, Jiang N, Zhou L, Huang H, Zhao J, Morse MA, Hobeika A, Ren J, Lyerly HK. Combination of DC/CIK adoptive T cell immunotherapy with chemotherapy in advanced non-small-cell lung cancer (NSCLC) patients: a prospective patients' preference-based study (PPPS). *Clin Transl Oncol.* 2019 Jun;21(6):721-728. doi: 10.1007/s12094-018-1968-3. Epub 2018 Oct 29. PMID: 30374838.
- [12]Lurje I, Hammerich L, Tacke F. Dendritic Cell and T Cell Crosstalk in Liver Fibrogenesis and Hepatocarcinogenesis: Implications for Prevention and Therapy of Liver Cancer. *Int J Mol Sci.* 2020 Oct 6;21(19):7378. doi: 10.3390/ijms21197378. PMID: 33036244; PMCID: PMC7583774.
- [13]Singh SK, Larsson M, Schön T, Stendahl O, Blomgran R. HIV Interferes with the Dendritic Cell-T Cell Axis of Macrophage Activation by Shifting Mycobacterium tuberculosis-Specific CD4 T Cells into a Dysfunctional Phenotype. *J Immunol.* 2019;202(3):816-826. doi:10.4049/jimmunol.1800523
- [14]Beyersdorf N, Kerkau T, Hünig T. CD28 co-stimulation in T-cell homeostasis: a recent perspective. *Immunotargets Ther.* 2015 May 28;4:111-22. doi: 10.2147/ITT.S61647. PMID: 27471717; PMCID: PMC4918251.
- [15]Kumar BV, Connors TJ, Farber DL. Human T Cell Development, Localization, and Function throughout Life. *Immunity.* 2018;48(2):202-213. doi:10.1016/j.immuni.2018.01.007
- [16]Kondo K, Ohigashi I, Takahama Y. Thymus machinery for T-cell selection. *Int Immunol.* 2019;31(3):119-125. doi:10.1093/intimm/dxy081
- [17]Germain RN. T-cell development and the CD4-CD8 lineage decision. *Nat Rev Immunol.* 2002;2(5):309-322. doi:10.1038/nri798
- [18]Bellón T. Mechanisms of Severe Cutaneous Adverse Reactions: Recent Advances. *Drug Saf.* 2019;42(8):973-992. doi:10.1007/s40264-019-00825-2
- [19]Fang D, Zhu J. Dynamic balance between master transcription factors determines the fates and functions of CD4 T cell and innate lymphoid cell subsets. *J Exp Med.* 2017 Jul 3;214(7):1861-1876. doi: 10.1084/jem.20170494. Epub 2017 Jun 19. PMID: 28630089; PMCID: PMC5502437.
- [20]Zhu X, Zhu J. CD4 T Helper Cell Subsets and Related Human Immunological Disorders. *Int J Mol Sci.* 2020;21(21):8011. Published 2020 Oct 28. doi:10.3390/ijms21218011
- [21]Ikeogu NM, Edechi CA, Akaluka GN, et al. Semaphorin 3E Promotes Susceptibility to Leishmania major Infection in Mice by Suppressing CD4+ Th1 Cell Response. *J Immunol.* 2021;206(3):588-598. doi:10.4049/jimmunol.2000516
- [22]Rana AK, Li Y, Dang Q, Yang F. Monocytes in rheumatoid arthritis: Circulating precursors of macrophages and osteoclasts and, their heterogeneity and plasticity role in RA pathogenesis. *Int Immunopharmacol.* 2018;65:348-359. doi:10.1016/j.intimp.2018.10.016
- [23]Shen L, Zhang H, Caimol M, et al. Invariant natural killer T cells in lupus patients promote IgG and IgG autoantibody production. *Eur J Immunol.* 2015;45(2):612-623. doi:10.1002/eji.201444760
- [24]Jorgovanovic D, Song M, Wang L, Zhang Y. Roles of IFN- γ in tumor progression and regression: a review. *Biomark Res.* 2020;8:49. Published 2020 Sep 29. doi:10.1186/s40364-020-00228-x

- [25]Leong JW, Chase JM, Romee R, et al. Preactivation with IL-12, IL-15, and IL-18 induces CD25 and a functional high-affinity IL-2 receptor on human cytokine-induced memory-like natural killer cells. *Biol Blood Marrow Transplant.* 2014;20(4):463-473. doi:10.1016/j.bbmt.2014.01.006
- [26]Tiegs G, Horst AK. TNF in the liver: targeting a central player in inflammation. *Semin Immunopathol.* 2022;44(4):445-459. doi:10.1007/s00281-022-00910-2
- [27]Guo Z, Gao WS, Wang YF, Gao F, Wang W, Ding WY. MiR-502 Suppresses TNF- α -Induced Nucleus Pulposus Cell Apoptosis by Targeting TARF2. *Biomed Res Int.* 2021;2021:5558369. Published 2021 Apr 1. doi:10.1155/2021/5558369
- [28]Kumar S, Jeong Y, Ashraf MU, Bae YS. Dendritic Cell-Mediated Th2 Immunity and Immune Disorders. *Int J Mol Sci.* 2019;20(9):2159. Published 2019 May 1. doi:10.3390/ijms20092159
- [29]Stark JM, Tibbitt CA, Coquet JM. The Metabolic Requirements of Th2 Cell Differentiation. *Front Immunol.* 2019;10:2318. Published 2019 Sep 27. doi:10.3389/fimmu.2019.02318
- [30]Nakayama T, Hirahara K, Onodera A, et al. Th2 Cells in Health and Disease. *Annu Rev Immunol.* 2017;35:53-84. doi:10.1146/annurev-immunol-051116-052350
- [31]Nicola S, Ridolfi I, Rolla G, et al. IL-17 Promotes Nitric Oxide Production in Non-Small-Cell Lung Cancer. *J Clin Med.* 2021;10(19):4572. Published 2021 Oct 1. doi:10.3390/jcm10194572
- [32]Li B, Huang L, Lv P, et al. The role of Th17 cells in psoriasis. *Immunol Res.* 2020;68(5):296-309. doi:10.1007/s12026-020-09149-1
- [33]Karczewski J, Mazur M, Rychlewska-Hańczewska A, Adamski Z. Rola limfocytów Th17 w patogenezie raka jelita grubego [Role of Th17 lymphocytes in pathogenesis of colorectal cancer]. *Postepy Hig Med Dosw (Online).* 2014;68:42-47. Published 2014 Jan 22. doi:10.5604/17322693.1086074
- [34]Dario A A Vignali, Lauren W Collison, Creg J Workman. How regulatory T cells work. *Nat Rev Immunol.* 2008 Jul;8(7):523-32. doi: 10.1038/nri2343.
- [35]Durgeau A, Virk Y, Cognac S, Mami-Chouaib F. Recent Advances in Targeting CD8 T-Cell Immunity for More Effective Cancer Immunotherapy. *Front Immunol.* 2018;9:14. Published 2018 Jan 22. doi:10.3389/fimmu.2018.00014
- [36]Zhang N, Bevan MJ. CD8(+) T cells: foot soldiers of the immune system. *Immunity.* 2011;35(2):161-168. doi:10.1016/j.immuni.2011.07.010
- [37]Sabat R, Wolk K, Loyal L, Döcke WD, Ghoreschi K. T cell pathology in skin inflammation. *Semin Immunopathol.* 2019;41(3):359-377. doi:10.1007/s00281-019-00742-7
- [38]Coe GL, Redd PS, Paschall AV, et al. Ceramide mediates FasL-induced caspase 8 activation in colon carcinoma cells to enhance FasL-induced cytotoxicity by tumor-specific cytotoxic T lymphocytes. *Sci Rep.* 2016;6:30816. Published 2016 Aug 4. doi:10.1038/srep30816
- [39]Jimbo H, Nagai H, Fujiwara S, Shimoura N, Nishigori C. Fas-FasL interaction in cytotoxic T cell-mediated vitiligo: The role of lesional expression of tumor necrosis factor- α and interferon- γ in Fas-mediated melanocyte apoptosis. *Exp Dermatol.* 2020;29(1):61-70. doi:10.1111/exd.14053
- [40]Lurje I, Hammerich L, Tacke F. Dendritic Cell and T Cell Crosstalk in Liver Fibrogenesis and Hepatocarcinogenesis: Implications for Prevention and Therapy of Liver Cancer. *Int J Mol Sci.* 2020;21(19):7378. Published 2020 Oct 6. doi:10.3390/ijms21197378
- [41]Liu J, Zhang X, Cheng Y, Cao X. Dendritic cell migration in inflammation and immunity. *Cell Mol Immunol.* 2021;18(11):2461-2471. doi:10.1038/s41423-021-00726-4
- [42]Murphy TL, Murphy KM. Dendritic cells in cancer immunology. *Cell Mol Immunol.* 2022;19(1):3-13. doi:10.1038/s41423-021-00741-5

- [43]Garri CS, Luke JJ. Dendritic Cells, the T-cell-inflamed Tumor Microenvironment, and Immunotherapy Treatment Response. *Clin Cancer Res.* 2020;26(15):3901-3907. doi:10.1158/1078-0432.CCR-19-1321
- [44]Gajewski TF, Cron KR. cDC1 dysregulation in cancer: An opportunity for intervention. *J Exp Med.* 2020;217(8):e20200816. doi:10.1084/jem.20200816
- [45]Noubade R, Majri-Morrison S, Tarbell KV. Beyond cDC1: Emerging Roles of DC Crosstalk in Cancer Immunity. *Front Immunol.* 2019;10:1014. Published 2019 May 9. doi:10.3389/fimmu.2019.01014
- [46] Shen Chunyi, Zhang Zhen, Tian Yonggui, Zhang Yi. Tws119 combined with cytokines promotes the differentiation and function of CD8⁺ + memory T cells [J]. *Chinese Journal of immunology*, 2019,35 (04): 435-439 + 445
- [47]Borzova NY, Ivanenkova NI, Sotnikova NY, Malyshkina AI. *Klin Lab Diagn.* 2020;65(5):294-298. doi:10.18821/0869-2084-2020-65-5-294-298
- [48]Zhang M, Yang W, Wang P, et al. CCL7 recruits cDC1 to promote antitumor immunity and facilitate checkpoint immunotherapy to non-small cell lung cancer. *Nat Commun.* 2020;11(1):6119. Published 2020 Nov 30. doi:10.1038/s41467-020-19973-6
- [49]Balan S, Saxena M, Bhardwaj N. Dendritic cell subsets and locations. *Int Rev Cell Mol Biol.* 2019;348:1-68. doi:10.1016/bs.ircmb.2019.07.004
- [50]Zhu S, Yang N, Wu J, et al. Tumor microenvironment-related dendritic cell deficiency: a target to enhance tumor immunotherapy. *Pharmacol Res.* 2020;159:104980. doi:10.1016/j.phrs.2020.104980
- [51]Rönnblom L. The importance of the type I interferon system in autoimmunity. *Clin Exp Rheumatol.* 2016;34(4 Suppl 98):21-24.
- [52]Chen K, Liu J, Cao X. Regulation of type I interferon signaling in immunity and inflammation: A comprehensive review. *J Autoimmun.* 2017;83:1-11. doi:10.1016/j.jaut.2017.03.008
- [53]Finetti F, Baldari CT. The immunological synapse as a pharmacological target. *Pharmacol Res.* 2018;134:118-133. doi:10.1016/j.phrs.2018.06.009
- [54]Dustin ML. The immunological synapse. *Cancer Immunol Res.* 2014;2(11):1023-1033. doi:10.1158/2326-6066.CIR-14-0161
- [55]van Panhuys N. TCR Signal Strength Alters T-DC Activation and Interaction Times and Directs the Outcome of Differentiation. *Front Immunol.* 2016;7:6. Published 2016 Jan 25. doi:10.3389/fimmu.2016.00006
- [56]Pang YG, Chang CC. Artificial Antigen Presentosomes for T Cell Activation. *Methods Mol Biol.* 2020;2111:141-151. doi:10.1007/978-1-0716-0266-9_12
- [57]Kishton RJ, Sukumar M, Restifo NP. Metabolic Regulation of T Cell Longevity and Function in Tumor Immunotherapy. *Cell Metab.* 2017;26(1):94-109. doi:10.1016/j.cmet.2017.06.016
- [58]Chapman NM, Boothby MR, Chi H. Metabolic coordination of T cell quiescence and activation. *Nat Rev Immunol.* 2020;20(1):55-70. doi:10.1038/s41577-019-0203-y
- [59]Shimauchi T, Piguet V. DC-T cell virological synapses and the skin: novel perspectives in dermatology. *Exp Dermatol.* 2015;24(1):1-4. doi:10.1111/exd.12511
- [60]Letourneur F, Klausner RD. Activation of T cells by a tyrosine kinase activation domain in the cytoplasmic tail of CD3 epsilon. *Science.* 1992;255(5040):79-82. doi:10.1126/science.1532456
- [61]Okoye IS, Houghton M, Tyrrell L, Barakat K, Elahi S. Coinhibitory Receptor Expression and Immune Checkpoint Blockade: Maintaining a Balance in CD8⁺ T Cell Responses to Chronic Viral Infections and Cancer. *Front Immunol.* 2017;8:1215. Published 2017 Sep 29. doi:10.3389/fimmu.2017.01215

- [62]Thangavelu G, Smolarchuk C, Anderson CC. Co-inhibitory molecules: Controlling the effectors or controlling the controllers? *Self Nonself*. 2010;1(2):77-88. doi:10.4161/self.1.2.11548
- [63]Sanchez-Lockhart M, Rojas AV, Fettis MM, Bauserman R, Higa TR, Miao H, Waugh RE, Miller J. T cell receptor signaling can directly enhance the avidity of CD28 ligand binding. *PLoS One*. 2014 Feb 24;9(2):e89263. doi: 10.1371/journal.pone.0089263. PMID: 24586641; PMCID: PMC3933428.
- [64]Crespo J, Sun H, Welling TH, Tian Z, Zou W. T cell anergy, exhaustion, senescence, and stemness in the tumor microenvironment. *Curr Opin Immunol*. 2013;25(2):214-221. doi:10.1016/j.coi.2012.12.003
- [65] Qin Qiuhong, Zhang Yaoyao, pan Jian, Huang Tianming, Chen Chengxiao, Luo Guorong. Effect of anti-CD3 / CD28 monoclonal antibody combined with PHA on T lymphocyte activation and proliferation [J]. *Progress in Microbiology and immunology*, 2020,48 (02): 16-21. Doi: 10.13309/j.cnki.pmi.2020.02.003
- [66] Xu Xuemei, Tang Zongsheng, Li Zhihong. The role of T cell CD28 family receptors in the pathogenesis of asthma [J]. *Journal of Southeast University (Medical Edition)*, 2016,35 (06): 1009-1013
- [67]Porciello N, Tuosto L. CD28 costimulatory signals in T lymphocyte activation: Emerging functions beyond a qualitative and quantitative support to TCR signalling. *Cytokine Growth Factor Rev*. 2016;28:11-19. doi:10.1016/j.cytogfr.2016.02.004
- [68]Beyersdorf N, Kerkau T, Hünig T. CD28 co-stimulation in T-cell homeostasis: a recent perspective. *Immunotargets Ther*. 2015;4:111-122. Published 2015 May 28. doi:10.2147/ITT.S61647
- [69]Minato N, Hattori M, Hamazaki Y. Physiology and pathology of T-cell aging. *Int Immunol*. 2020;32(4):223-231. doi:10.1093/intimm/dxaa006
- [70]Beckermann KE, Dudzinski SO, Rathmell JC. Dysfunctional T cell metabolism in the tumor microenvironment. *Cytokine Growth Factor Rev*. 2017;35:7-14. doi:10.1016/j.cytogfr.2017.04.003
- [71]Chen L, Flies DB. Molecular mechanisms of T cell co-stimulation and co-inhibition [published correction appears in *Nat Rev Immunol*. 2013 Jul;13(7):542]. *Nat Rev Immunol*. 2013;13(4):227-242. doi:10.1038/nri3405
- [72]Osii RS, Otto TD, Garside P, Ndungu FM, Brewer JM. The Impact of Malaria Parasites on Dendritic Cell-T Cell Interaction. *Front Immunol*. 2020;11:1597. Published 2020 Jul 24. doi:10.3389/fimmu.2020.01597
- [73]Chalupova AM, Vosahlikova S, Rozkova D, et al. Methods to assess DC-dependent priming of T cell responses by dying cells. *Methods Enzymol*. 2020;632:55-65. doi:10.1016/bs.mie.2019.05.045
- [74]Wonderlich ER, Wu WC, Normolle DP, Barratt-Boyes SM. Macrophages and Myeloid Dendritic Cells Lose T Cell-Stimulating Function in Simian Immunodeficiency Virus Infection Associated with Diminished IL-12 and IFN- α Production. *J Immunol*. 2015;195(7):3284-3292. doi:10.4049/jimmunol.1500683
- [75]Jung HJ, Park SH, Cho KM, Jung KI, Cho D, Kim TS. Threonyl-tRNA Synthetase Promotes T Helper Type 1 Cell Responses by Inducing Dendritic Cell Maturation and IL-12 Production via an NF- κ B Pathway. *Front Immunol*. 2020;11:571959. Published 2020 Oct 14. doi:10.3389/fimmu.2020.571959
- [76]Garris CS, Arlauckas SP, Kohler RH, et al. Successful Anti-PD-1 Cancer Immunotherapy Requires T Cell-Dendritic Cell Crosstalk Involving the Cytokines IFN- γ and IL-12. *Immunity*. 2018;49(6):1148-1161.e7. doi:10.1016/j.immuni.2018.09.024
- [77] Chen Gang, Wang Jingke. Correlation between relative lymphocyte count and acute myocardial infarction [J]. *Xinjiang Medical Journal*, 2007 (02): 95
- [78]Petersen RP, Campa MJ, Sperlazza J, et al. Tumor infiltrating Foxp3+ regulatory T-cells are associated with recurrence in pathologic stage I NSCLC patients. *Cancer*. 2006;107(12):2866-2872. doi:10.1002/cncr.22282

- [79]Liu H, Zhang T, Ye J, et al. Tumor-infiltrating lymphocytes predict response to chemotherapy in patients with advance non-small cell lung cancer. *Cancer Immunol Immunother.* 2012;61(10):1849-1856. doi:10.1007/s00262-012-1231-7
- [80]Kalafati L, Kourtzelis I, Schulte-Schrepping J, et al. Innate Immune Training of Granulopoiesis Promotes Anti-tumor Activity. *Cell.* 2020;183(3):771-785.e12. doi:10.1016/j.cell.2020.09.058
- [81]Bonanni V, Sciumè G, Santoni A, Bernardini G. Bone Marrow NK Cells: Origin, Distinctive Features, and Requirements for Tissue Localization. *Front Immunol.* 2019;10:1569. Published 2019 Jul 10. doi:10.3389/fimmu.2019.01569
- [82]Han X, Yang Q, Zhang J, Cao J. Correlation between changes in the number of peripheral blood lymphocytes and survival rate in patients with cervical cancer after radio-chemotherapy. *Cancer Radiother.* 2021;25(1):72-76. doi:10.1016/j.canrad.2020.08.045
- [83]Jia W, Fu ZL, Wang X, et al. Decreased Absolute Number of Circulating Regulatory T Cells in Patients With Takayasu's Arteritis. *Front Immunol.* 2021;12:768244. Published 2021 Dec 23. doi:10.3389/fimmu.2021.768244
- [84]Saito H, Shimizu S, Kono Y, et al. Score of the preoperative absolute number of lymphocytes, monocytes, and neutrophils as a prognostic indicator for patients with gastric cancer. *Surg Today.* 2019;49(10):850-858. doi:10.1007/s00595-019-01817-6
- [85]Pike LRG, Bang A, Mahal BA, et al. The Impact of Radiation Therapy on Lymphocyte Count and Survival in Metastatic Cancer Patients Receiving PD-1 Immune Checkpoint Inhibitors. *Int J Radiat Oncol Biol Phys.* 2019;103(1):142-151. doi:10.1016/j.ijrobp.2018.09.010
- [86]Oh SY, Heo J, Noh OK, Chun M, Cho O, Oh YT. Absolute Lymphocyte Count in Preoperative Chemo-radiotherapy for Rectal Cancer: Changes Over Time and Prognostic Significance. *Technol Cancer Res Treat.* 2018;17:1533033818780065. doi:10.1177/1533033818780065
- [87]Xia Y, Li W, Li Y, Liu Y, Ye S, Liu A, Yu J, Jia Y, Liu X, Chen H, Guo Y. The clinical value of the changes of peripheral lymphocyte subsets absolute counts in patients with non-small cell lung cancer. *Transl Oncol.* 2020 Dec;13(12):100849. doi: 10.1016/j.tranon.2020.100849. Epub 2020 Aug 28.
- [88]Kiritsy MC, McCann K, Mott D, et al. Mitochondrial respiration contributes to the interferon gamma response in antigen-presenting cells. *Elife.* 2021;10:e65109. Published 2021 Nov 2. doi:10.7554/eLife.65109
- [89]Wu Shanshan, Wang Baishan, Yan Feng, Zhang Ning, Zhang Cheng, Li Zhijing Analysis on the correlation between the expression of NKG2D on CD8⁺ T cells and their cytotoxic activity in patients with non-small cell lung cancer before and after taking Fuzheng anticancer formula [C] // Proceedings of the 16th Shenyang Science and Technology Annual Conference (Science, engineering, agriculture and medicine). [publisher unknown], 2019:180-188
- [90]Rotte A. Combination of CTLA-4 and PD-1 blockers for treatment of cancer. *J Exp Clin Cancer Res.* 2019;38(1):255. Published 2019 Jun 13. doi:10.1186/s13046-019-1259-z
- [91]Anderson AC, Joller N, Kuchroo VK. Lag-3, Tim-3, and TIGIT: Co-inhibitory Receptors with Specialized Functions in Immune Regulation. *Immunity.* 2016;44(5):989-1004. doi:10.1016/j.immuni.2016.05.001
- [92]Hosseini A, Gharibi T, Marofi F, Babaloo Z, Baradaran B. CTLA-4: From mechanism to autoimmune therapy. *Int Immunopharmacol.* 2020;80:106221. doi:10.1016/j.intimp.2020.106221
- [93]Dyck L, Mills KHG. Immune checkpoints and their inhibition in cancer and infectious diseases. *Eur J Immunol.* 2017;47(5):765-779. doi:10.1002/eji.201646875
- [94]Chen R, Tao Y, Xu X, et al. The efficacy and safety of nivolumab, pembrolizumab, and atezolizumab in treatment of advanced non-small cell lung cancer. *Discov Med.* 2018;26(143):155-166.

- [95]Halmos B, Burke T, Kalyvas C, et al. A Matching-Adjusted Indirect Comparison of Pembrolizumab + Chemotherapy vs. Nivolumab + Ipilimumab as First-Line Therapies in Patients with PD-L1 TPS [?]1% Metastatic NSCLC. *Cancers (Basel)*. 2020;12(12):3648. Published 2020 Dec 4. doi:10.3390/cancers12123648
- [96]Zahran AM, Hetta HF, Mansour S, Saad ES, Rayan A. Reviving up dendritic cells can run cancer immune wheel in non-small cell lung cancer: a prospective two-arm study. *Cancer Immunol Immunother*. 2021;70(3):733-742. doi:10.1007/s00262-020-02704-7
- [97]Ahluwalia P, Ahluwalia M, Mondal AK, Sahajpal NS, Kota V, Rojiani MV, Kolhe R. Natural Killer Cells and Dendritic Cells: Expanding Clinical Relevance in the Non-Small Cell Lung Cancer (NSCLC) Tumor Microenvironment. *Cancers (Basel)*. 2021 Aug 11;13(16):4037. doi: 10.3390/cancers13164037. PMID: 34439191; PMCID: PMC8394984.
- [98]Wang Y, Xiang Y, Xin VW, Wang XW, Peng XC, Liu XQ, Wang D, Li N, Cheng JT, Lyv YN, Cui SZ, Ma Z, Zhang Q, Xin HW. Dendritic cell biology and its role in tumor immunotherapy. *J Hematol Oncol*. 2020 Aug 3;13(1):107. doi: 10.1186/s13045-020-00939-6. PMID: 32746880; PMCID: PMC7397618.
- [99]Zahran AM, Hetta HF, Mansour S, Saad ES, Rayan A. Reviving up dendritic cells can run cancer immune wheel in non-small cell lung cancer: a prospective two-arm study. *Cancer Immunol Immunother*. 2021 Mar;70(3):733-742. doi: 10.1007/s00262-020-02704-7. Epub 2020 Sep 12. PMID: 32918587.
- [100]Wang Y, Zhao N, Wu Z, Pan N, Shen X, Liu T, Wei F, You J, Xu W, Ren X. New insight on the correlation of metabolic status on 18F-FDG PET/CT with immune marker expression in patients with non-small cell lung cancer. *Eur J Nucl Med Mol Imaging*. 2020 May;47(5):1127-1136. doi: 10.1007/s00259-019-04500-7. Epub 2019 Sep 9. PMID: 31502013.
- [101]Bianchi F, Alexiadis S, Camisaschi C, Truini M, Centonze G, Milione M, Balsari A, Tagliabue E, Sfondrini L. TLR3 Expression Induces Apoptosis in Human Non-Small-Cell Lung Cancer. *Int J Mol Sci*. 2020 Feb 20;21(4):1440. doi: 10.3390/ijms21041440. PMID: 32093313; PMCID: PMC7073031.
- [102] Wang Jingbo, Huang Xue, Li Furong. Research progress and Prospect of impaired dendritic cell function in lung cancer [J]. *Cancer*, 2020,39 (06): 248-260
- [103]Martinez M, Moon EK. CAR T Cells for Solid Tumors: New Strategies for Finding, Infiltrating, and Surviving in the Tumor Microenvironment. *Front Immunol*. 2019;10:128. Published 2019 Feb 5. doi:10.3389/fimmu.2019.00128
- [104]Sternern RC, Sternern RM. CAR-T cell therapy: current limitations and potential strategies. *Blood Cancer J*. 2021;11(4):69. Published 2021 Apr 6. doi:10.1038/s41408-021-00459-7
- [105]Turtle, C.J., Riddell, S.R., 2010. Artificial antigen-presenting cells for use in adoptive immunotherapy. *Cancer J*. 16, 374–381.
- [106]Kim, J.V., Latouche, J.B., Riviere, I., Sadelain, M., 2004. The ABCs of artificial antigen presentation. *Nat. Biotechnol*. 22, 403–410.
- [107]Zeng W, Su M, Anderson KS, Sasada T. Artificial antigen-presenting cells expressing CD80, CD70, and 4-1BB ligand efficiently expand functional T cells specific to tumor-associated antigens. *Immunobiology*. 2014;219(8):583-592. doi:10.1016/j.imbio.2014.03.003
- [108]Sun, S., Cai, Z., Langlade-Demoyen, P., Kosaka, H., Brunmark, A., Jackson, M.R., Peterson, P.A., Sprent, J., 1996. Dual function of Drosophila cells as APCs for naive CD8+ T cells: implications for tumor immunotherapy. *Immunity* 4, 555–564.
- [109]Latouche, J.B., Sadelain, M., 2000. Induction of human cytotoxic T lymphocytes by artificial antigen-presenting cells. *Nat. Biotechnol*. 18, 405–409.
- [110]Butler, M.O., Lee, J.S., Ansen, S., Neuberger, D., Hodi, F.S., Murray, A.P., Drury, L., Bere

- zovskaya, A., Mulligan, R.C., Nadler, L.M., Hirano, N., 2007. Long-lived antitumor CD8+ lymphocytes for adoptive therapy generated using an artificial antigen presenting cell. *Clin. Cancer Res.* 13, 1857–1867.
- [111]Sun, W., et al., Connecting the dots: artificial antigen presenting cell-mediated modulation of natural killer T cells. *J Interferon Cytokine Res*, 2012. 32(11): p. 505-16.
- [112]Eggermont LJ, Paulis LE, Tel J, Figdor CG. Towards efficient cancer immunotherapy: advances in developing artificial antigen-presenting cells. *Trends Biotechnol.* 2014;32(9):456-465. doi:10.1016/j.tibtech.2014.06.007
- [113]Shao J, Xu Q, Su S, et al. Artificial antigen-presenting cells are superior to dendritic cells at inducing antigen-specific cytotoxic T lymphocytes. *Cell Immunol.* 2018;334:78-86. doi:10.1016/j.cellimm.2018.10.002
- [114]East JE, Sun W, Webb TJ. Artificial antigen presenting cell (aAPC) mediated activation and expansion of natural killer T cells. *J Vis Exp.* 2012;(70):4333. Published 2012 Dec 29. doi:10.3791/4333
- [115]Schappert A, Schneck JP, Suarez L, Oelke M, Schutz C. Soluble MHC class I complexes for targeted immunotherapy. *Life Sci.* 2018;209:255-258. doi:10.1016/j.lfs.2018.08.023
- [116]Bottcher JP, Bonavita E, Chakravarty P, et al. NK Cells Stimulate Recruitment of cDC1 into the Tumor Microenvironment Promoting Cancer Immune Control. *Cell.* 2018;172(5):1022-1037.e14. doi:10.1016/j.cell.2018.01.004
- [117]Sun Z, Deng G, Peng X, et al. Intelligent photothermal dendritic cells restart the cancer immunity cycle through enhanced immunogenic cell death. *Biomaterials.* 2021;279:121228. doi:10.1016/j.biomaterials.2021.121228
- [118]Suarez L, Wang R, Carmer S, et al. AIM Platform: A Novel Nano Artificial Antigen-Presenting Cell-Based Clinical System Designed to Consistently Produce Multi-Antigen-Specific T-Cell Products with Potent and Durable Anti-Tumor Properties. *Transfus Med Hemother.* 2020;47(6):464-471. doi:10.1159/000512788
- [119]Cintolo JA, Datta J, Mathew SJ, Czerniecki BJ. Dendritic cell-based vaccines: barriers and opportunities. *Future Oncol.* 2012;8(10):1273-1299. doi:10.2217/fon.12.125
- [120]Park J, Andrade B, Seo Y, Kim MJ, Zimmerman SC, Kong H. Engineering the Surface of Therapeutic "Living" Cells. *Chem Rev.* 2018;118(4):1664-1690. doi:10.1021/acs.chemrev.7b00157
- [121]Abbina S, Siren EMJ, Moon H, Kizhakkedathu JN. Surface Engineering for Cell-Based Therapies: Techniques for Manipulating Mammalian Cell Surfaces. *ACS Biomater Sci Eng.* 2018;4(11):3658-3677. doi:10.1021/acsbomaterials.7b00514
- [122]Manning JC, Romero A, Habermann FA, Garcia Caballero G, Kaltner H, Gabius HJ. Lectins: a primer for histochemists and cell biologists. *Histochem Cell Biol.* 2017;147(2):199-222. doi:10.1007/s00418-016-1524-6
- [123]Glaffig M., Stergiou N., Hartmann S., Schmitt E., Kunz H., A synthetic MUC1 anticancer vaccine containing mannose ligands for targeting macrophages and dendritic cells. *ChemMedChem* 13, 25–29 (2018).
- [124]Yu L, Feng R, Zhu L, et al. Promoting the activation of T cells with glycopolymer-modified dendritic cells by enhancing cell interactions. *Sci Adv.* 2020;6(47):eabb6595. Published 2020 Nov 20. doi:10.1126/sciadv.abb6595
- [125]Qu J, Mei Q, Chen L, Zhou J. Chimeric antigen receptor (CAR)-T-cell therapy in non-small-cell lung cancer (NSCLC): current status and future perspectives. *Cancer Immunol Immunother.* 2021;70(3):619-631. doi:10.1007/s00262-020-02735-0
- [126] Zheng nairong, Xu Jianqing. Research progress of CAR-T cell immunotherapy [J]. *Journal of Fudan University: Medical Edition*, 2022,49 (2): 295-299

- [127] Cheng Hao, ihobali Chi, Zhou Quan, Ying Jianming, Shi Susheng. Research progress of CAR-T in immunotherapy of gastric cancer [J]. Chinese cancer clinic, 2022,49 (9): 480-486
- [128] Xu Guangxian. Research progress of CAR-T cell immunotherapy in tumor treatment [J]. Journal of Guangdong Medical University, 2022,40 (2): 121-131
- [129]NARAYAN V, BARBER-ROTENBERG J, JUNG I, et al. PSMA-targeting TGF β -insensitive armored CAR T cells in metastatic castration-resistant prostate cancer: a phase 1 trial [J/OL]. [2022-03-25].<https://doi.org/10.1038/s41591-022-01726-1>.
- [130]HEITZENEDER S, BOSSE K, ZHU Z,et al. GPC2-CAR T cells tuned for low antigen density mediate potent activity against neuroblastoma without toxicity[J]. Cancer Cell, 2022, 40(1):53-69.
- [131]Mao X, Xu J, Wang W, et al. Crosstalk between cancer-associated fibroblasts and immune cells in the tumor microenvironment: new findings and future perspectives. Mol Cancer. 2021;20(1):131. Published 2021 Oct 11. doi:10.1186/s12943-021-01428-1
- [132]HOU A, CHEN L, CHEN Y. Navigating CAR-T cells through the solid-tumour microenvironment[J]. Nat Rev Drug Discov, 2021, 20(7):531-550.
- [133]HONG M, CLUBB J, CHEN Y.Engineering CAR-T cells for next-generation cancer therapy[J]. Cancer Cell, 2020, 38 (4):473-488.
- [134]THE LANCET ONCOLOGY. CAR T-cell therapy for solid tumours[J]. Lancet Oncol, 2021, 22:893.
- [135]Min J, Long C, Zhang L, et al. c-Met specific CAR-T cells as a targeted therapy for non-small cell lung cancer cell A549. Bioengineered. 2022;13(4):9216-9232. doi:10.1080/21655979.2022.2058149
- [136]Li H, Harrison EB, Li H, et al. Targeting brain lesions of non-small cell lung cancer by enhancing CCL2-mediated CAR-T cell migration. Nat Commun. 2022;13(1):2154. Published 2022 Apr 20. doi:10.1038/s41467-022-29647-0
- [137]Hung LVM, Ngo HT, Van Pham P. Clinical Trials with Cytokine-Induced Killer Cells and CAR-T Cell Transplantation for Non-small Cell Lung Cancer Treatment. Adv Exp Med Biol. 2020;1292:113-130. doi:10.1007/5584_2020_522
- [138]Liu H, Ma Y, Yang C, et al. Severe delayed pulmonary toxicity following PD-L1-specific CAR-T cell therapy for non-small cell lung cancer. Clin Transl Immunology. 2020;9(10):e1154. Published 2020 Oct 9. doi:10.1002/cti2.1154
- [139]Jiang ZB, Huang JM, Xie YJ, et al. Evodiamine suppresses non-small cell lung cancer by elevating CD8+ T cells and downregulating the MUC1-C/PD-L1 axis. J Exp Clin Cancer Res. 2020;39(1):249. Published 2020 Nov 19. doi:10.1186/s13046-020-01741-5
- [140]Wang H, Meng AM, Li SH, Zhou XL. A nanobody targeting carcinoembryonic antigen as a promising molecular probe for non-small cell lung cancer. Mol Med Rep. 2017;16(1):625-630. doi:10.3892/mmr.2017.6677
- [141]Li BT, Smit EF, Goto Y, et al. Trastuzumab Deruxtecan in HER2-Mutant Non-Small-Cell Lung Cancer. N Engl J Med. 2022;386(3):241-251. doi:10.1056/NEJMoa2112431
- [142]Li N, Liu S, Sun M, et al. Chimeric Antigen Receptor-Modified T Cells Redirected to EphA2 for the Immunotherapy of Non-Small Cell Lung Cancer. Transl Oncol. 2018;11(1):11-17. doi:10.1016/j.tranon.2017.10.009

