

ARTICLE TYPE

High-resolution depth measurements in digital microscopic surgery

S. Babilon* | P. Myland | L. Schlestein | J. Klages | T. Q. Khanh

¹Laboratory of Lighting Technology,
Technical University of Darmstadt,
Germany

Correspondence

*Dr.-Ing. Sebastian Babilon, Email:
babilon@lichttechnik.tu-darmstadt.de

Present Address

Technical University of Darmstadt
Laboratory of Lighting Technology
Hochschulstr. 4a
64289 Darmstadt
Germany

Summary

Fully digital microscopes are becoming more and more common in surgical applications. In addition to high-resolution stereoscopic images of the operating field, which can be transmitted over long distances or stored directly, these systems offer further potentials by supporting the surgical workflow based on their fully digital image processing chain. For example, the image display can be adapted to the respective surgical scenario by adaptive color reproduction optimization or image overlays with additional information, such as the tissue topology. Knowledge of this topology can be used for computer-assisted or AR-guided microsurgical treatments and enables additional features such as spatially resolved spectral reconstruction of surface reflectance. In this work, a new method for high-resolution depth measurements in digital microsurgical applications is proposed, which is based on the principle of laser triangulation. Part of this method is a sensor data fusion procedure to properly match the laser scanner and camera data. In this context, a strategy based on RBF interpolation techniques is presented to handle missing or corrupt data, which, due to the measuring principle, can occur on steep edges and through occlusion. The proposed method is used for the acquisition of high-resolution depth profiles of various organic tissue samples, proving the feasibility of the proposed concept as a supporting technology in a digital microsurgical workflow.

KEYWORDS:

medical imaging, AR-guided microsurgical treatments, depth measurements, laser sensor, tissue topology, digital image processing

1 | INTRODUCTION

The use of surgical microscopes has become the gold standard for many ophthalmic, otorhinolaryngological and neuro-surgical procedures (see e.g.,^{1,2,3,4}). In addition to conventional optical surgical microscopes, fully digital solutions are more and more common in clinical practice. Besides offering high-resolution and even stereoscopic vision of the operative field, most of these systems are intended to provide additional benefit by supporting the surgical workflow based on their completely digital image processing chain. Potential advantages of digital over conventional optical systems comprise for example the *in-situ* visualization of preoperative image data (acquired e.g. from tomographic scanning procedures), the overlay of the surgeon's view with signals from external video sources (e.g., from an additional endoscope), and the possibility of collaborative viewing and improved surgical assistance by synchronizing the surgeon's exact view of the operative field to external monitors.

10 Various approaches to further explore the benefits of digital microscopes for surgical applications are reported in the literature,
11 most of which are related to the development of new and dedicated image processing algorithms. Gard *et al.*⁵ for example
12 proposed a new method of precise distance and contour measurements of patient specific anatomy without external hardware
13 by tracking a marked spherical instrument tip in the stereoscopic image pair provided by a digital surgical microscope. They
14 showed that by applying a self-updating template-matching algorithm sufficiently accurate distance measurements in the sub-
15 millimeter regime could be achieved depending on the zoom level and the microscope's depth-of-field. In combination with
16 stereo triangulation, their method enabled the reconstruction of instrument tip trajectories that could eventually be used as
17 precisely registered augmented-reality (AR) overlays projected into the surgeon's field of view.

18 Based on a similar approach, the MultiARC project⁶ aims to develop an interactive and multimodal AR system for computer-
19 assisted ear, nose and throat (ENT) microsurgery. This system is intended to allow for interactive remote surgical assistance
20 including live annotations, three-dimensional point-to-point distance measurements, and the overlaying visualization of pre-
21 operatively acquired patient's data (e.g., from magnetic resonance imaging). Potential benefits of providing additional AR
22 information as an integrative part of the digital microscope's image processing pipeline are discussed by Aschke *et al.*⁷ in the
23 context of neurosurgical treatments. A more general overview can be found in Khor *et al.*⁸, who besides emphasizing the huge
24 potential of such digital surgical environments also elaborate on their current practical limitations.

25 Another aspect recently discussed in the context of AR-supported microsurgery is the feasibility of automated tissue
26 recognition based on hyperspectral imaging (HSI, see Lu and Fei⁹ for a review on HSI in medical applications). As a first-proof-
27 of-concept, Wisotzky *et al.*^{10,11} developed an HSI setup to noninvasively analyze the optical characteristics of *in vivo* tissue
28 samples encountered during ENT microsurgery in order to explore the suitability of this approach. They equipped a digital sur-
29 gical microscope with an additional filter wheel which was placed in front of the white-light LED illumination unit holding 16
30 different narrow bandpass filters. By synchronizing the rotation of the filter wheel and, therefore, the spectral changes of the
31 illumination of the surgical field with the image capturing frame rate of the microscope, they were able to capture the spectral
32 reflection behavior of various tissue structures in the visible range from 400 to 700 nm. The evaluation of the collected data
33 showed that even for tissue samples which are not or hardly distinguishable for the human eye, the analysis in the 16-dimensional
34 wavelength domain lead to clearly different, individually unique trends in the samples' reflection characteristics allowing for a
35 robust tissue differentiation.

36 One of the downsides of this method as discussed by Wisotzky *et al.* is the existing time misalignment which is caused by the
37 rather slow rotation of the filter wheel making its current implementation inapplicable for clinical routine. In a further paper,
38 Wisotzky *et al.*¹² therefore proposed a novel HSI approach replacing the filter wheel by two hyperspectral snapshot cameras
39 covering the visible and near-infrared range from 460 to 980 nm. This new setup allowed for acquiring the complete hyperspectral
40 dataset in a single shot making it more suitable for clinical application. However, single snapshot HSI always comes with the
41 cost of significantly reduced spatial resolution due to the large filter arrays (here: 4×4 and 5×5 , respectively) applied to the
42 camera sensor's surface. In addition, both the filter wheel and the camera-based approach require very expensive and complex
43 components to be additionally mounted to the surgical microscope, considerably increasing engineering effort and production
44 costs.

45 For these reasons, we worked on a new HSI approach for estimating the spectral reflectance characteristics of organic tissue
46 samples without the need for expensive hardware supplements. The theoretical framework and first results of this approach
47 have recently been published elsewhere¹³. In principle, the idea is to combine the light emission of various narrow-banded
48 monochromatic and phosphor-converted white LEDs creating the multi-channel illumination unit of the digital microscope with
49 the imaging capabilities of its standard three-channel RGB camera system. During data acquisition, the tissue sample under
50 inspection is illuminated by flashing the individual LED channels one after the other. At the same time, the image capturing
51 has to be synchronized to this illumination sequence, which artificially increases the dimensionality of the output data without
52 the need of additional hardware or costly equipment. As a result, one obtains $n \times 3$ monochromatic images of the same tissue
53 sample, where n denotes the number of different LED channels, which can subsequently be used as input for applying a Wiener
54 filter estimation approach^{14,15,16,17,18} to reconstruct the tissue's spectral reflectance characteristics in the visible regime. Note
55 that a similar spectral reconstruction procedure is proposed as part of the MultiARC project discussed above.

56 Once the reflectance characteristics are known, it was found that significantly enhanced color correction can be achieved dur-
57 ing image processing¹³. Leading to improved color reproduction properties, such an enhancement is considered to be beneficial
58 for the success of video-assisted surgeries, where, in general, different kinds of tissues and critical structures must be distin-
59 guished by the surgeon not just through their textures but also through their colors. In addition, promising results were obtained
60 with regard to the premises of automated tissue recognition based on previous reflectance estimation.

61 However, as shown in our recent paper, for the required absolutely correct intensity scaling, the underlying Wiener-filter
 62 estimation technique demands the exact knowledge of the measurement geometry, which is determined by the distance and angle
 63 of the tissue surface with respect to the illumination and image capturing modules of the surgical microscope. A corresponding
 64 pixel-position dependent scaling factor $\delta^{(i,j)}$ was therefore introduced and determined for the reported measurements by previous
 65 calibration using a standard reflectance target at a fixed geometry. However, such a calibration procedure to be conducted in
 66 advance of a surgery is not really convenient in practice and must be re-performed every time the microscope head is moved to
 67 another position.

68 Basically, this scaling factor can also be determined from the known pixel position (i, j) of the point-of-interest (POI) in the
 69 projected x - y -plane, for which the reflectance spectra should be estimated, and an additional depth measurement to determine
 70 the absolute distance value ρ between the POI and its corresponding location on the imaging sensor along the z -axis. From these
 71 geometric measurements, the known position of the multi-channel LED illuminant in relation to the image capturing module,
 72 and an initial one-time calibration during the manufacturing process, the absolute spectral power distribution (SPD) in terms
 73 of the spectral flux received by the tissue surface at the POI position can be derived for each given LED channel. Once the
 74 individual SPDs at the POI position are known, they can be used as input in the Wiener filter matrix (see previous work¹³ for
 75 corresponding equations) to properly estimate the spectral surface reflectance at the POI on an absolute scale.

76 Even though the actual calculations and the algorithmic implementation are straightforward, the challenging task is to accu-
 77 rately perform the depth measurements. It was found in a pre-test in preparation of the current work that uncertainties of up
 78 to 3 mm in the depth measurements result in average deviations of the reconstructed reflectance spectra from the ground truth
 79 of about 2.5%. Hence, with regard to the idea of automated tissue recognition based on reflectance estimation, a depth accu-
 80 racy of less than ± 0.3 mm would be favorable to keep the expected deviations from the ground truth due to systematic depth
 81 measurement inaccuracies below 1%.

Since modern digital surgical microscopes usually provide stereoscopic imaging, trying to extract depth information directly
 from the corresponding disparity mapping seems to be quite obvious. Based on stereo computer vision theory¹⁹, the accuracy
 in depth estimation can be approximated by

$$|\Delta\rho| = \frac{Z^2}{fB} \Delta d, \quad (1)$$

82 where f is the focal length, B is the baseline distance of the stereo camera configuration, and Δd is the error of the disparity
 83 calculation between the left and right image. This also called correspondence accuracy describes the expected error caused by
 84 an incorrect matching of both images at the POI location and depends on image noise, scene texture, and other scene properties
 85 such as occlusions and non-Lambertian surfaces²⁰, the used matching algorithm as well as on the resolution and pixel size of
 86 the camera sensors¹⁹.

87 In benchmark tests for the performance evaluation of different matching algorithms, such as for example the Middlebury
 88 Stereo Evaluation²¹, average disparity errors of 2 to 3.5 pixels can be found for the best performing algorithms under controlled
 89 experimental conditions. Assuming further a typical focal length of surgical microscopes ranging between 45 to 80 mm, a
 90 baseline distance of the (virtual) stereo camera configuration ranging from 16 to 22 mm, and a pixel size of the camera sensors
 91 of 6 to 8 μm^6 , the accuracy in depth estimation under optimal conditions obtain from disparity mapping is expected to be of the
 92 order of ± 0.6 to ± 1.6 mm for an assumed working distance of 250 mm.

93 With regard to the indicated depth resolution of ± 0.3 mm required for accurate *in-situ* reflectance reconstruction, a direct
 94 depth estimation from stereoscopic imaging of currently available surgical microscopes seems to be inexpedient. Hence, a new
 95 approach for high-resolution *in-situ* depth measurements based on laser triangulation was developed by the authors and will be
 96 presented in the following. Although laser triangulation is an established method in industrial production processesing and has
 97 already been tested in various other biomedical applications (see e.g.,^{22,23,24}), the presented work, to the authors' best knowledge,
 98 is the first to make use of this concept for achieving highly accurate depth measurements in microsurgical treatments.

99 The structure of this paper is organized as follows. Sec. 2 starts with a short discussion of the experimental setup and the
 100 measurement protocol used for data acquisition. In Sec. 3, the applied strategies for accuracy estimation, sensor data fusion, and
 101 data visualization will be presented. Based on these preconditions, first depth profiling results for various organic tissue samples
 102 will be reported in order to qualitatively evaluate the performance of the proposed method. Finally, a short summary of the key
 103 findings of this work as well as an outlook on future research intentions will be given in Sec. 4.

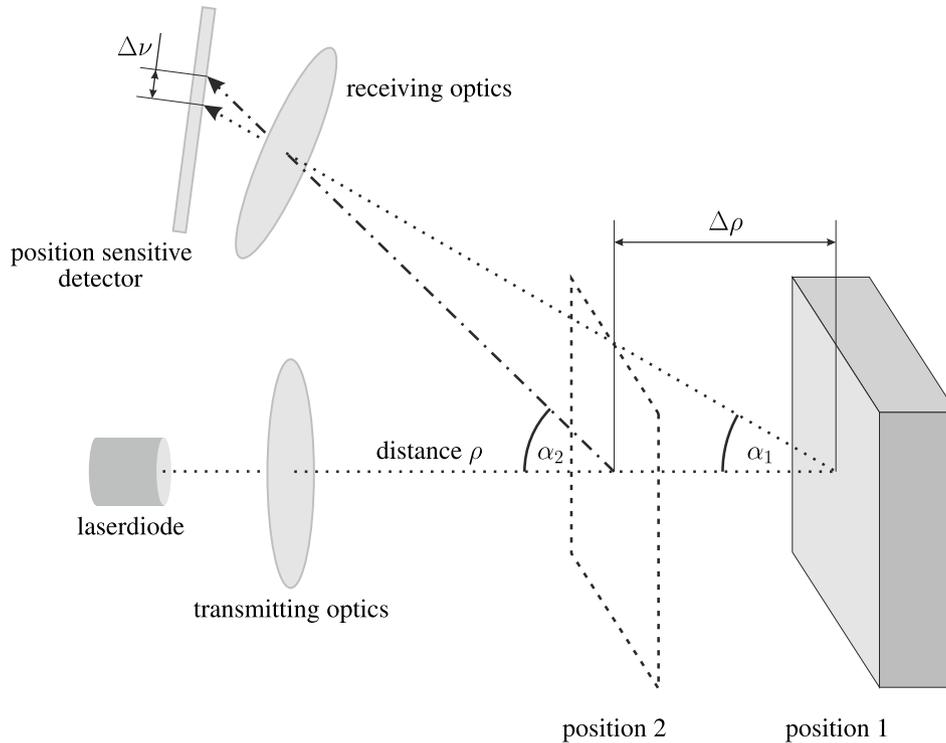


FIGURE 1 Illustration of the laser triangulation principle. Based on an initial distance calibration, it is possible to determine the corresponding distance ρ between the light emitting laser diode and the reflecting object for each position of the imaged laser spot on the detector's surface in the receiving unit.

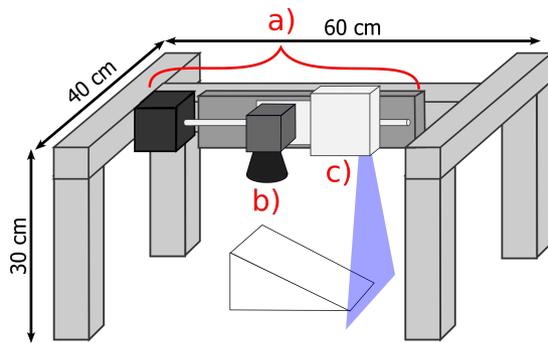
2 | EXPERIMENTAL SETUP AND MEASUREMENT PROTOCOL

High-precision laser-triangulation modules are common components of modern coordinate measuring machines in various industrial production processes. They are typically used for quality control, safety, level measurements, and positioning applications. Being solely based on trigonometry, they are relatively simple and cost-efficient measuring tools that allow for robust and accurate distance measurements well into the micrometer range. Due to this accuracy and outstanding depth resolution, laser triangulation is used in this work to determine the pixel-dependent scaling factor $\delta^{(i,j)}$ required for the microsurgical application sketched in the introduction.

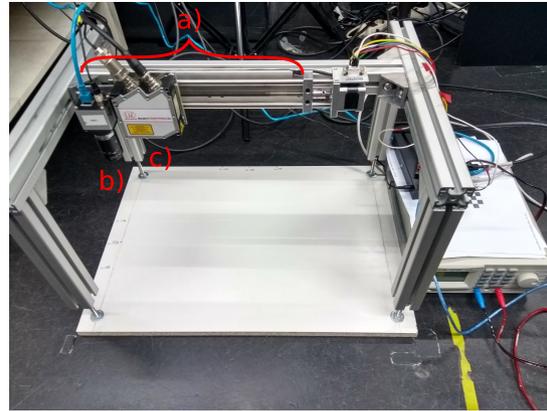
Its basic principle is shown in Fig. 1. A laser emitter, which consists of a laser diode of a certain wavelength supplemented with dedicated transmitting optics components, is used to project a light spot onto the surface of an object whose distance ρ should be measured. The scattered light from the object's surface is collected by the receiving optics and focused on a position sensitive detector (PSD; e.g., a CCD sensor). If a change $\Delta\rho$ in the distance between the laser diode and the object's surface occurs, the angle α under which the scattered light is received by the PSD changes accordingly, leading to a position shift $\Delta\nu$ of the focused image of the light spot on the detector's surface. As a result of this relationship, the PSD can be calibrated such that absolute measures of the distance ρ between the laser diode and the surface of the object become feasible.

In order to suppress disturbances caused by the ambient light, the receiving unit is usually equipped with an additional spectral band-pass filter, which is optimized to match the specific wavelength of the laser diode. Other factors that may reduce the accuracy of the distance measurements are for example unfavorable surface characteristics, distance-dependent laser spot enlargement, out-of-focus measurements and optical aberrations such as coma, astigmatism, or spherical errors of the lens systems. With regard to the intended microsurgical application, it is very important to be aware of these error sources and a careful characterization of the final experimental setup must therefore be performed on a dedicated reference target to obtain an estimate of the overall depth measurement accuracy.

For the experimental test setup used in this work, two different laser triangulation modules with similar specifications but different wavelengths were at our disposal: A red-emitting Micro-Epsilon scanCONTROL 2900-100 with 658 nm wavelength and



(a) Schematic illustration of the experimental setup including its dimensions



(b) Picture of the experimental setup as it can be found in the laboratory

FIGURE 2 Experimental setup as a first proof-of-concept for high-resolution depth profiling of organic tissue samples based on laser triangulation. Its main parts are a) the linear motion axis, b) a digital camera system to simulate the imaging workflow of a surgical microscope, and c) the laser-triangulation profile scanner.

127 a blue-emitting Micro-Epsilon scanCONTROL 2900-100/BL with 405 nm wavelength. Both devices are Class 2M (≤ 8 mW)
 128 laser profile scanners, i.e., instead of using a single light spot, as shown in Fig. 1, they project a complete line segment compris-
 129 ing 1,280 individual measuring points onto the object's surface, which will be denoted as the x -axis in the following. According
 130 to the corresponding data sheet²⁵, they both offer a reference depth resolution of $12\ \mu\text{m}$ within the identical measurement range
 131 (working distance) from 190 mm to 290 mm along the z -axis and the same maximum measurement frequency of 300 Hz. How-
 132 ever, pre-tests with both devices revealed that the blue-emitting laser module is better suited for profiling organic tissue than its
 133 red-emitting equivalent, which compared to the former suffered from too much failed measurements when being applied to scan
 134 various organic test samples. The remaining part of this paper should therefore consider the blue-emitting results only.

135 For a complete scan of an arbitrary object, the scanning device must be moved along the y -axis at a constant velocity v .
 136 Fig. 2 shows the corresponding experimental setup. As can be seen, the laser scanner and an additional industrial GigE camera
 137 are mounted to the carriage of a linear motion system driven by a stepper motor. The linear motion system is held by a stable
 138 aluminum profile fixture and oriented such that both scanner and the camera are facing perpendicularly downwards to the bottom
 139 plate, where the test objects are placed for scanning. Care must be taken to ensure an almost vibration-free attachment. In this
 140 setup, the camera, which is a monochrome IDS UI-5490SE-M-GL Rev.2²⁶, is intended to simulate the digital imaging of a
 141 surgical microscope in order to address the question of how the output of both systems can efficiently be combined in future
 142 applications.

143 The dimensions of the aluminum profile fixture (cf. Fig. 2) as well as the camera lens ($f = 12$ mm) were chosen to match the
 144 measurement range and scanning width of the laser triangulation module. The stepper motor is used in half step mode to achieve
 145 a smooth and even running behavior^{27,28}. A relatively low measurement frequency of 50 Hz is chosen as a trade-off between
 146 a moderate scanning velocity to reduce vibrations and the total time required for a single scan. The corresponding scanning
 147 velocity was adjusted such that an approximately equal resolution in x - and y -direction can be achieved. As a result, an average
 148 velocity of $(4.153 \pm 0.002)\ \text{mm s}^{-1}$ is obtained, which was determined empirically from repeated measurements ($n = 100$). This
 149 gives a resolution along the y -axis of $(83.1 \pm 4.2)\ \mu\text{m}$, while the resolution in x -direction depends on the distance between laser
 150 scanner and test object and ranges from approximately $81\ \mu\text{m}$ (for objects with a height of 4 cm) to $94\ \mu\text{m}$ (on the level of the
 151 bottom plate). Hence, it can be concluded that the proposed experimental setup allows for high-resolution measurements with
 152 sufficiently small uncertainties to generate, as a first proof-of-concept, diagnostically conclusive depth profiles of organic tissue
 153 samples with regard to the requirements defined in the introduction.

154 The corresponding measurement protocol is conceived as follows. After the test object/tissue sample has been centered on
 155 the bottom plate, the carriage of the linear motion axis, with both camera and laser scanner being attached to it, is moved
 156 from one end of the axis to the other at the given constant scanning velocity. The actual data acquisition starts 1 s after the
 157 movement has been initiated and terminates 1 s before the end position is reached in order to exclude the phases of nonlinear

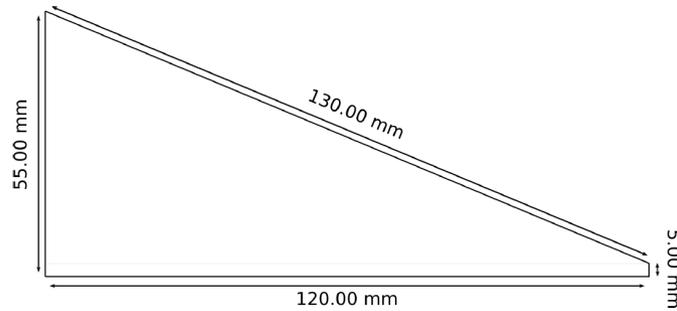


FIGURE 3 Reference target used to determine the overall depth measurement accuracy of the proposed laser-triangulation method. The wedge-shape was chosen to imitate a well-defined surface gradient enabling a more realistic estimation of the application-related measurement errors.

158 motor acceleration and deceleration. During the phase of constant movement, measurements from the laser profile scanner are
 159 read every 20 ms, which complies with the desired 50 Hz measurement frequency, so that a total number of approximately
 160 1,500×1,280 measurements are performed on the tissue sample resulting in a high-resolution depth profile of this test object.
 161 After the depth scanning is finished, the carriage is moved backwards until the camera and the center of the bottom plate are
 162 aligned to ensure the largest possible overlap between the measurement range of the laser scanner and the camera's field of view
 163 for a subsequent sensor data fusion (see Sec. 3.2). Once this specific position is reached, an image of the test object is captured
 164 at a resolution of 3,840×2,748 pixels. Afterwards, the carriage is eventually moved back to its initial position and the whole
 165 setup is ready for the next acquisition run. In total, it takes approximately 46.5 s to complete a single scan.

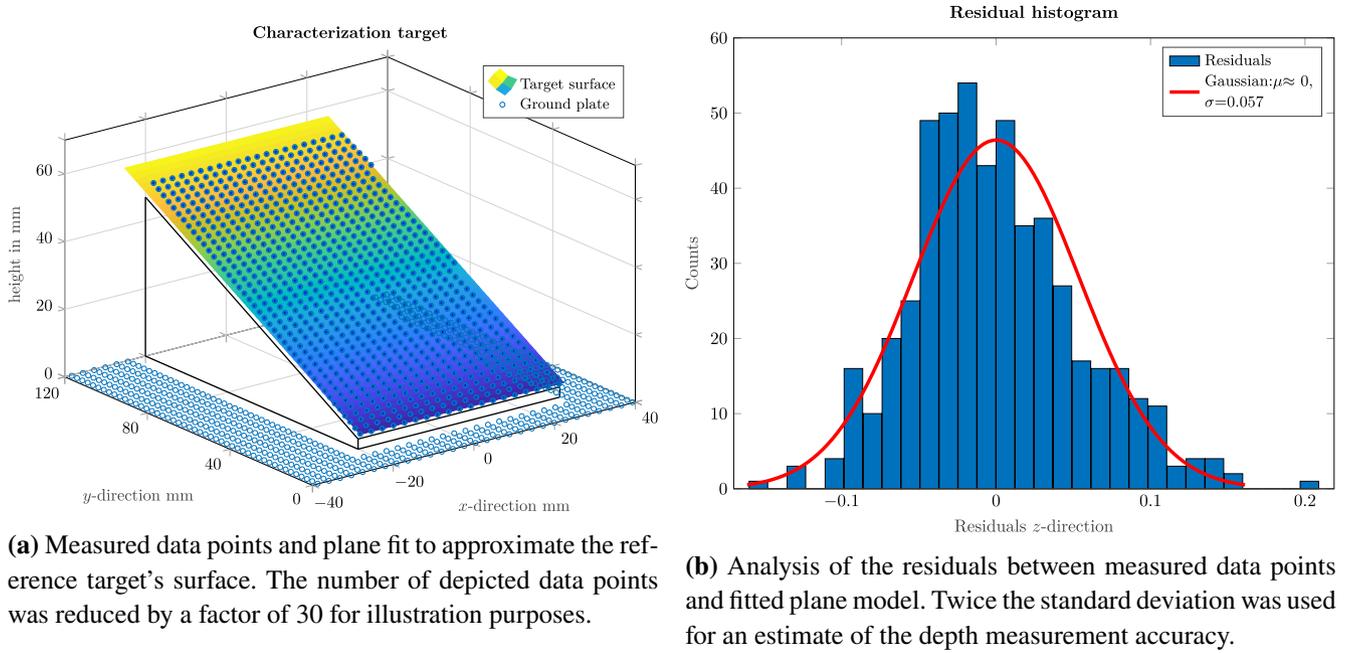
166 3 | SCANNING CHARACTERIZATION AND DEPTH MEASUREMENT RESULTS

167 Based on the description of the experimental setup and the measurement protocol given in the previous section, the following
 168 parts are intended to address the issues of data acquisition, error estimation, and data processing. In Sec. 3.1, the overall depth
 169 measurement accuracy along the z-direction should be evaluated. For this purpose, a dedicated reference target was used as a
 170 known test sample from which an error estimate is obtained by comparing nominal and measured values by means of residual
 171 analysis. This serves as an indicator whether the proposed triangulation method could meet the required depth resolution to be
 172 applicable for accurate *in-situ* reflectance estimation as discussed in the introduction of this paper. In Sec. 3.2, an efficient method
 173 of sensor data fusion is presented for matching both laser scanner and camera data as well as for visualizing the generated depth
 174 profiles in a genuine workflow. Finally, in Sec. 3.3, measurement results and acquired depth profiles of two different organic
 175 tissue samples are reported as a first proof-of-concept of the proposed method.

176 3.1 | Depth Measurements on Reference Target

177 In order to determine the overall error in the depth measurements of the current experimental setup, a dedicated reference target
 178 was designed and manufactured accordingly for system characterization. Its form and dimensions are shown in Fig. 3. As can
 179 be seen, a wedge-shaped reference target was chosen. It is made of purely white polyethylene material with excellently diffuse
 180 reflection properties and a nominal production tolerance of $\pm 50 \mu\text{m}$. The shape of the reference target is intended to simulate
 181 a well-defined surface gradient giving a more realistic estimate of the application-related measurement errors. In general, the
 182 measurement uncertainty in z-direction also depends on the uncertainties observed for the other directions. In the current setup,
 183 mainly the deviations along the y-axis are crucial for the overall depth measurement accuracy because of the respective carriage
 184 movement and must therefore be implicated by using a non-flat, gradient-showing reference target, like the one presented here,
 185 for the characterization process.

186 Before the actual characterization started, a baseline measurement had been performed first in order to account for measure-
 187 ment noise and flatness imperfections of the bottom plate and/or the underground. This baseline was then used to correct the
 188 actual measurement of the reference target, for which the protocol described in Sec. 2 was used.



(a) Measured data points and plane fit to approximate the reference target's surface. The number of depicted data points was reduced by a factor of 30 for illustration purposes.

(b) Analysis of the residuals between measured data points and fitted plane model. Twice the standard deviation was used for an estimate of the depth measurement accuracy.

FIGURE 4 Results of the laser-triangulation depth measurements performed on the wedge-shaped reference target. For convenience, a re-scaling was applied to transform the z -axis from a distance measure to a height scale (left). Based on the residual analysis (right), an estimate of $\pm 104 \mu\text{m}$ is found for the overall depth measurement accuracy.

189 Fig. 4(a) depicts the results of the laser-triangulation depth measurements performed on the wedge-shaped reference target.
 190 Re-scaling was applied to transform the z -axis from a distance measure to height scale. As can be seen, a plane was fitted to the
 191 data points to approximate the reference target's surface. The error estimation was then performed based on the corresponding
 192 residuals shown in Fig. 4(b). Here, twice the standard deviation was chosen giving an estimate of the overall depth measurement
 193 accuracy of $\pm 104 \mu\text{m}$.

194 Compared to the initially formulated requirements, the laser triangulation method consequently provides a sufficiently accurate
 195 depth profiling and distance measurement, which particularly outperforms depth estimation based on stereo vision. Even when
 196 considering the full range of the residuals plotted in Fig. 4(b), which goes up to $205 \mu\text{m}$, the required depth resolution of $\pm 0.3 \text{ mm}$
 197 is still met.

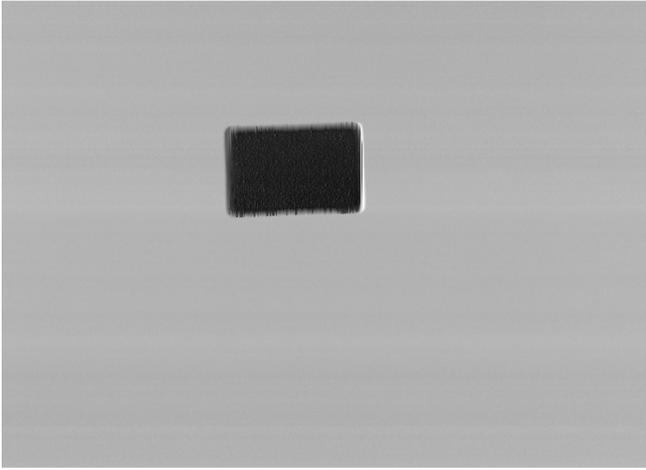
198 3.2 | Sensor Data Fusion and Visualization

199 Given that the proposed laser triangulation method clearly complies with the requirements for distance measurement and depth
 200 resolution, the next step is to perform the sensor data fusion in order to match the laser scanner and camera data. Since the
 201 measurement geometry is always fixed and given by the experimental setup shown in Fig. 2, an initial one-time calibration
 202 is sufficient. For the sake of efficiency, a standard object recognition approach can be applied to determine the corresponding
 203 transformation from the coordinate system of the laser scanner to the coordinate system of the camera.

Assuming affinity between both coordinate systems²⁹, this transformation basically consists of a translation, dilation, and rotation, which can be summarized by a single matrix $\mathbf{T}_{\text{affine}}$ projecting data points from one coordinate system into the other, i.e.,

$$\mathbf{x}_{\text{camera}} = \mathbf{T}_{\text{affine}} \mathbf{x}_{\text{laser}} \quad (2)$$

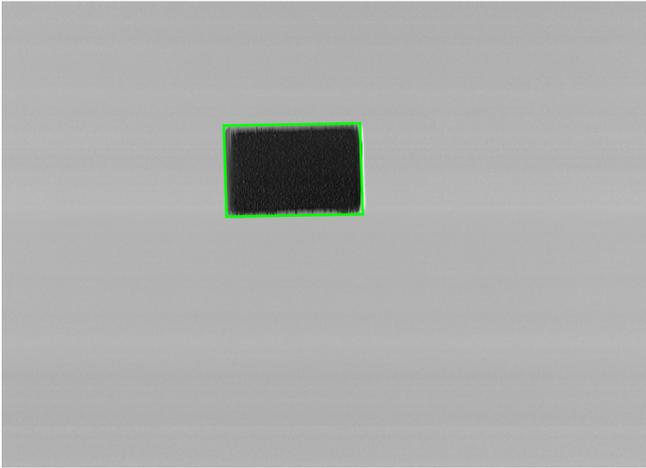
204 In order to determine this projection matrix, a simple cuboid calibration target was used. It is made of black polyethylene
 205 and its dimensions read $27.4 \text{ mm} \times 19.6 \text{ mm} \times 2.5 \text{ mm}$. This calibration target was again centered on the bottom plate and the
 206 scanning and measurement protocol as described in Sec. 2 was initiated. The collected profile data of the laser scanner were
 207 then transformed to an 8 bit gray-scale image so that an object recognition algorithm can be applied. In Figs. 5(a) and (b), the
 208 resulting "images" of the calibration target as obtained from the laser scanner and camera output are shown and compared to
 209 each other.



(a) Gray-scale image of the laser-scanner profile data obtained for the rectangular calibration target



(b) Monochrome camera image of the rectangular calibration target



(c) Contour detection of the calibration target from the laser-scanner profile data. The green rectangle represents the minimum enclosing area.



(d) Contour detection of the imaged calibration target. The green rectangle represents the minimum enclosing area.

FIGURE 5 Illustration of the sensor data fusion process to match laser scanner and camera data using a rectangular calibration target. Assuming affinity, the transformation from one coordinate system into the other, and therefore the matching procedure, can be described by a translation, dilation, and rotation, which can be summarized in a simple projection matrix. To determine this matrix, corresponding data points in the image representations (upper row) of both data acquisition systems are identified by means of object recognition (lower row).

Next, an adaptive thresholding algorithm^{30,31} is applied to transform both images of the calibration target into binary representations that subsequently allow for a straightforward, accurate edge and corner detection of its contours. This is performed by using the OpenCV implementation of the border following algorithm of Suzuki and Abe³² with a subsequent search for a minimum rectangular enclosing area³³ as shown in Figs. 5(c) and (d). By identifying three corresponding corners in the two different images, the projection matrix to transform from the laser scanner coordinate system to the camera coordinate system (see Eq. (2)) can eventually be determined and is given by

$$\mathbf{T}_{\text{affine}} = \begin{pmatrix} 2.5583 & -0.0321 & 253.27 \\ 0.0424 & 2.7456 & -523.44 \end{pmatrix} \quad (3)$$

for the current experimental setup. As long as the geometry between the laser scanner and the camera is not changed in the experimental setup, $\mathbf{T}_{\text{affine}}$ can automatically be applied right after each data acquisition run. The resulting depth profile for an arbitrary test object is then stored together with the captured image data as a new structure which, in the discussed microsurgical application, enables the direct access of depth and distance information at each given pixel location.

This, however, is only feasible as long as there are no perturbations in the acquired data of the laser scanner so that a pixel-wise mapping can be performed. In general, laser triangulation works best on diffusely reflecting, non-specular surfaces, where the incident laser beam is scattered such that a significant amount of scattered light is received by the PSD. In real-world measurements certain functional impairments, like for example specular reflections and occlusions, cannot be precluded entirely and, as a result, lead to missing data points in the laser scan matrix vitiating not just the mapping process but also reducing the visualization quality of the captured depth profiles.

To deal with these perturbations and to minimize the impact of missing data points, interpolation based on a radial basis function (RBF) approach should be applied. In general, RBF methods are used to approximate multivariate functions that are only known at a finite number of points by linear combinations of terms based on a single univariate, radially symmetric function. Here, we use them to interpolate missing data points in the laser scan matrix from their known surrounding measurement values and to approximate the test object's surface for visualization purposes.

Assume that the surface can be described by a smooth function $f : \mathbb{R}^2 \rightarrow \mathbb{R}$, where n function values

$$f(\mathbf{x}_i) = y_i, \quad \text{for } i = 1, \dots, n, \quad (4)$$

are known by measurement. This, in general, unknown function can then be approximated by

$$s(\mathbf{x}) = \sum_{i=1}^n \lambda_i \phi(\|\mathbf{x} - \mathbf{x}_i\|), \quad (5)$$

which eventually allows for a depth measure evaluation at any given point \mathbf{x} . Here, $\phi(r)$ is the adopted RBF, which is a radially symmetric, real-valued function defined on $[0, \infty)$, \mathbf{x}_i are the data points for which the function y_i is known, λ_i are scalar parameters, and $\|\cdot\|$ denotes a suitably chosen norm, usually (and also in the present case) the Euclidean norm $|\cdot|$.

As discussed by Carr *et al.*³⁴, a simple linear RBF $\phi(r) = r$ is a good choice for reconstructing surface data and should therefore be used in the following. Hence, Eq. (5) simplifies to

$$s(\mathbf{x}) = \sum_{i=1}^n \lambda_i |\mathbf{x} - \mathbf{x}_i|. \quad (6)$$

Given that

$$s(\mathbf{x}_i) = f(\mathbf{x}_i) = y_i, \quad \text{for } i = 1, \dots, n, \quad (7)$$

a $n \times n$ linear system of equations (LSE) can eventually be derived to solve for the coefficients λ_i that specify the RBF in Eq. (6). This LSE is given by

$$\mathbf{A} \boldsymbol{\lambda} = \mathbf{y}, \quad (8)$$

where $\boldsymbol{\lambda} = (\lambda_1, \lambda_2, \dots, \lambda_n)^T$ summarizes the RBF coefficients and $\mathbf{y} = (y_1, y_2, \dots, y_n)^T$ the known (measured) surface values. The entries of the square-symmetric interpolation matrix \mathbf{A} are then given by

$$A_{ij} = |\mathbf{x}_i - \mathbf{x}_j|, \quad \text{with } i, j = 1, \dots, n, \quad (9)$$

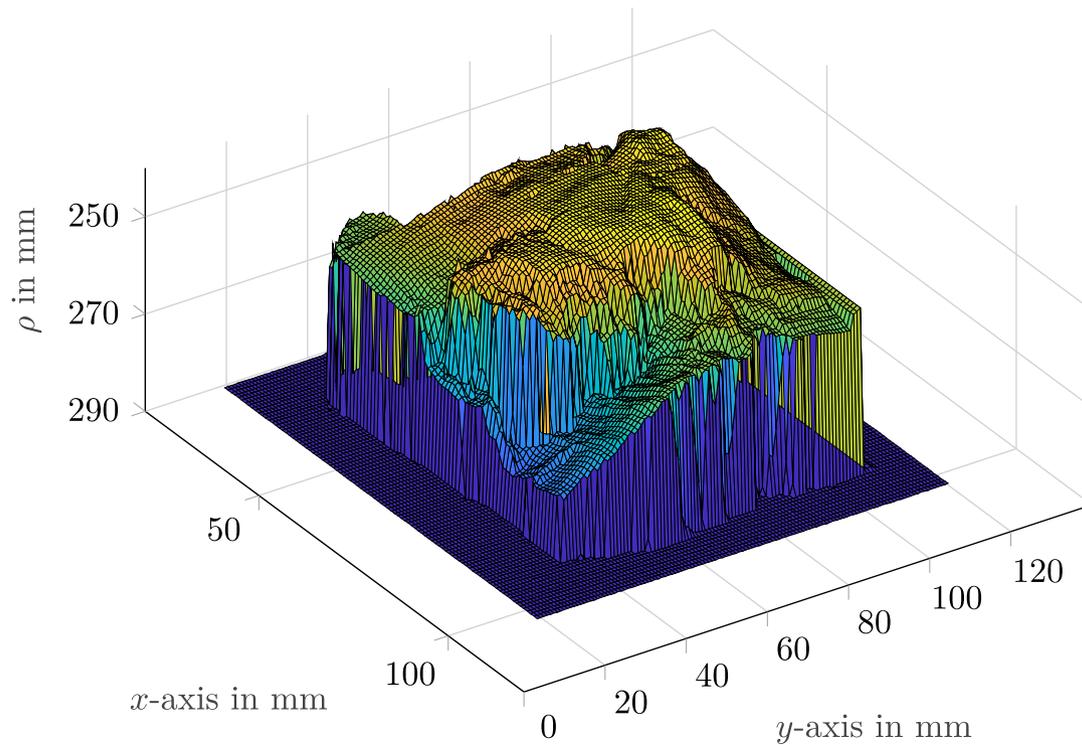
which is non-singular and, consequently, guarantees the unique existence of the coefficients λ_i as long as the chosen RBF $\phi(r)$ is strictly positive definite³⁵, which, using a simple linear RBF in conjunction with the Euclidean norm, is the case here.

By solving the LSE of Eq. (8), one obtains the coefficient vector $\boldsymbol{\lambda}$ and, thus, the surface approximation function $s(\mathbf{x})$ of Eq. (6), which can subsequently be used for visualization and sensor data fusion, as will be shown in the following section. In each case, a Python implementation of the LAPACK driver routine `sSYSV`³⁶ was used. It is based on the diagonal pivoting method with a partial pivoting strategy introduced by Bunch and Kaufmann³⁷ and allows for finding the corresponding solution of the symmetric LSE in a very efficient manner.

3.3 | Results for Organic Tissue Samples

As a first proof-of-concept of the proposed method for high-resolution depth measurements in microsurgery based on laser triangulation, results obtained for two different organic tissue samples, i.e., a piece of fractured bone from a pig's femur and muscle tissue from the upper part of a pig's shoulder, are reported in this section. Figs. 6 and 7 visualize the respective measurement

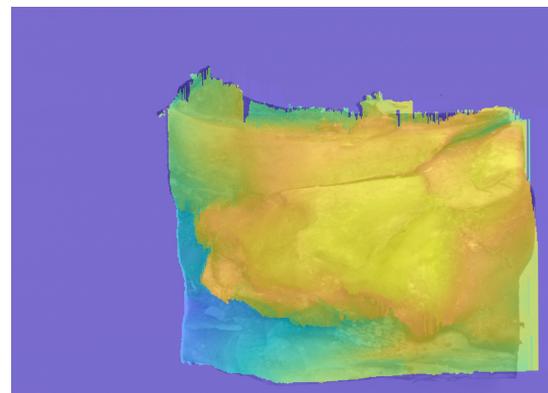
fractured bone



(a) Three-dimensional depth reconstruction of the fractured bone sample. The RBF method was applied for the interpolation of missing data points.



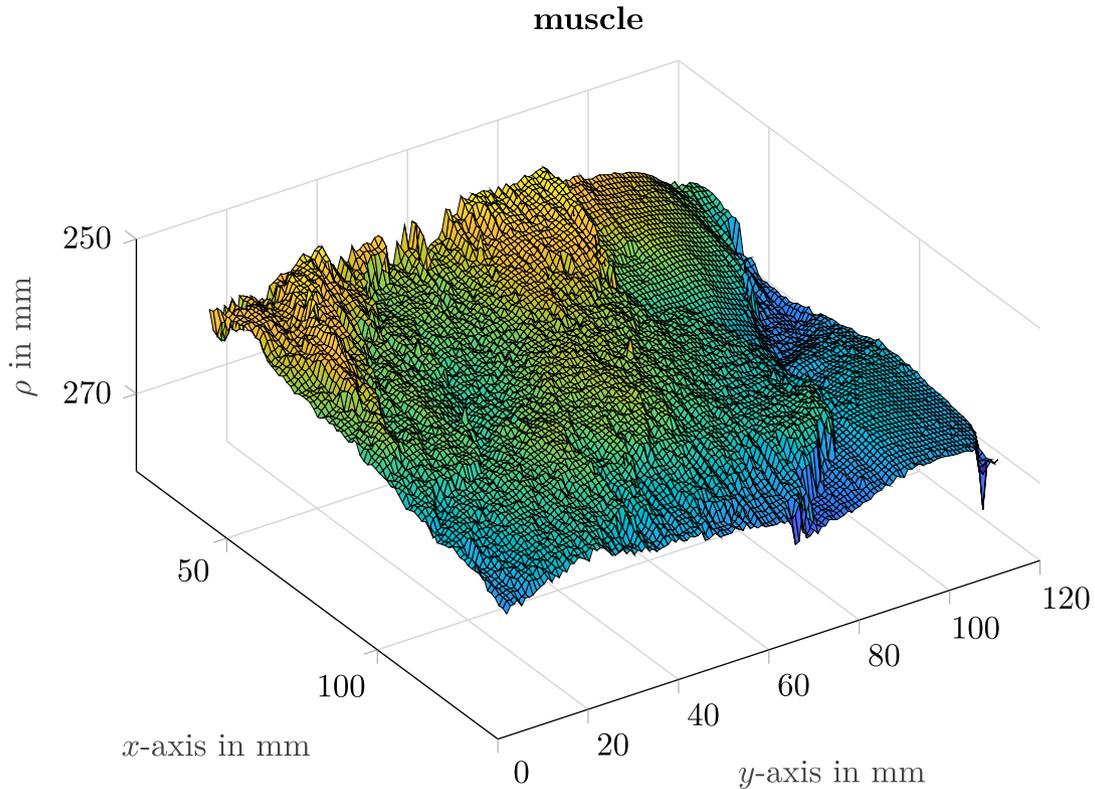
(b) Gray-scale image of the fractured bone sample as acquired by the monochrome image capturing system



(c) Color map overlay to visualize the sensor data fusion process for the fractured bone sample

FIGURE 6 Visual representation of the laser-triangulation depth measurement results for the fractured bone sample. The fusion of laser scanner and camera data eventually yields a data matrix containing both image and depth information at each given pixel location.

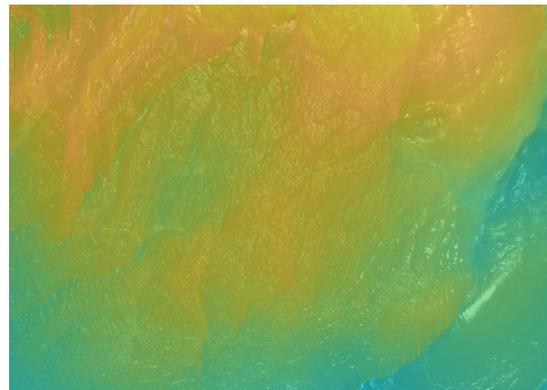
239 and sensor data fusion outcomes. The corresponding raw and RBF data can be downloaded via our institutional webpage³⁸.
 240 This source also provides further access to the results of some additional tissue samples (webpage-only) which, for the sake of
 241 brevity and due to similar conclusions to be drawn, are not explicitly discussed here.



(a) Three-dimensional depth reconstruction of the muscle tissue sample. The RBF method was applied for the interpolation of missing data points.



(b) Gray-scale image of the muscle tissue sample as acquired by the monochrome image capturing system



(c) Color map overlay to visualize the sensor data fusion process for the muscle tissue sample

FIGURE 7 Visual representation of the laser-triangulation depth measurement results for the muscle tissue sample. The fusion of laser scanner and camera data eventually yields a data matrix containing both image and depth information at each given pixel location.

242 In all cases – including the webpage-only samples – the proposed method works quite well. It does not only allow for a high-
 243 resolution depth map visualization of the respective tissue structures illustrated as three-dimensional surface plots but, based on
 244 the applied sensor data fusion, also enables the accurate, pixelwise determination of distance values in order to determine the
 245 pixel-dependent scaling factor $\delta^{(i,j)}$ that is required for reflectance estimation and subsequent tissue recognition as outlined in
 246 the introduction section of this paper. Problems of incorrect data representations because of failed RBF interpolation are only
 247 observed at the edges of the tissue samples, where the corresponding surface gradient becomes overly steep so that the scattered

light is not reflected back onto the PSD of the laser scanner. As a result, this yields too large areas of missing data points along the edges. Another source of such functional impairments compromising the RBF processing are occlusions of lower lying tissue layers at the edge structures due to the finite opening angle of the line segment emitted by the laser scanner.

Besides these small, edge-related disturbances, no further severe measurement errors are observed. Comparing for example the results obtained for bone and muscle tissue shown in Figs. 6 and 7, respectively, an equally good performance of the proposed method can be concluded when assessing the inner structure of the tissue surfaces, i.e., all parts that are sufficiently far from the samples' edges. As can be seen, the tissue fine structures are resolved properly in both cases without showing any non-smooth perturbations neither at the transitions from osseous to marrow structures in case of the fractured bone sample nor at the transition from fiber to adipose or conjunctive components in case of the muscle tissue. This basically indicates that neither residual surface moisture nor a specific tissue color prevent the proposed method from providing accurate depth measurements when being applied to different kinds of organic tissue samples.

Failures in the data acquisition and/or RBF interpolation of the samples' inner parts are therefore only expected if the tissue itself absorbs too much of the incident light from the laser scanner or prevents the light from being scattered diffusely back into the PSD unit because of the occurrence of too much specular reflections in conjunction with an unfavorable incident angle. In either case, missing data points in the laser scanner data matrix representing these surface areas would be the consequence. As long as these areas are relatively small, the missing data can still be compensated appropriately by the implemented RBF approach. Thus, larger errors or pronounced perturbations were not observed for any of the various organic tissue samples considered in this work. It can therefore be concluded that both absorption and specular reflection seem to be minor issues and little to worry about in practice – at least with regard to the current experimental setup.

4 | CONCLUSION AND OUTLOOK

The exact knowledge of the tissue topology including the option of performing precise three-dimensional distance and contour measurements can facilitate and support the surgeon's workflow within the framework of computer-assisted or AR-guided microsurgical treatments. For example, it may help to choose the appropriate size of patient-tailored implants and prostheses and allow for accurate real-time tracking of instrument tips in order to reduce the risk of complications caused by harmful tissue contact^{5,39}.

In the current paper, however, a slightly different use case was sketched for research motivation. As part of a previous exploratory study on automated tissue recognition in digital stereoscopic microsurgery based on Wiener filter reflectance estimation from HSI input, it has been shown that the distance and angle of the captured tissue surface with respect to the illumination and image capturing modules of the microscope must be determined with high accuracy in order to achieve a sufficiently good tissue recognition performance. In this context, a depth accuracy of less than ± 0.3 mm has been concluded to be favorable for application. Based on the assumption of typical microscope specifications and the benchmark results of high-performance stereo-matching algorithms usually applied for such tasks, it has been demonstrated that the naïve approach of using the microscope's stereoscopic image data for directly extracting depth information seems to be inappropriate to comply with these accuracy requirements.

In this work, a new method for high-resolution depth measurements in digital microsurgical applications was therefore proposed adopting the principle of laser triangulation. With the experimental setup discussed in Sec. 2, the expected error in depth measurements performed on a dedicated reference target was shown to be of the order of ± 104 μm , which conforms to the required depth resolution of at least ± 0.3 mm or better. Given that the proposed laser triangulation method clearly complies with these requirements, a method of sensor data fusion was subsequently discussed in order to properly match the laser scanner and camera data in a straightforward and intuitive manner. In this context, a strategy based on RBF interpolation techniques was introduced to handle missing or corrupt data as an integral part of this automated mapping process. The proposed method was eventually used to acquire – as a first proof-of-concept – high-resolution depth profiles of various organic tissue samples. It could be shown that, besides some small disturbances at the samples' edges, excellent measurement performance was achieved for all considered tissue structures without observing any larger errors or pronounced perturbations. Thus, it was concluded that, with regard to the current experimental setup, failures in the data acquisition and RBF interpolation are only expected if the tissue itself absorbs too much of the incident light or prevents the light from being scattered diffusely back into the PSD unit because of

an unfavorable combination of specular reflections and a too large incident angle. In all other cases, the system performs sufficiently well to provide high-resolution depth measurements that comply with the requirements to be used for subsequent Wiener filter reflectance estimation or any other microsurgical treatment that requires accurate depth and distance measurements.

Based on these encouraging results, the next logical step would be to proceed with the system integration into a real digital stereoscopic microsurgical device. The aim of this step is to combine the sensor data of the laser scanner with the stereoscopic image data of the microscope in a similar manner performed here on two-dimensional image data. The expected benefit of such a system integration is that the additional availability of native stereoscopic information from the microscope may help to improve the RBF performance and to deal with missing data of the laser scanner at edge-like structures in the surgical field. In these critical regions, depth information reconstructed from the microscopes stereoscopic image data, even though less accurate, can be used to fill the voids in the corresponding data matrix for achieving a more accurate RBF interpolation showing less pronounced perturbations at the tissue edges. In addition to the RBF method adopted for the current work due to its simplicity, more sophisticated interpolation algorithms should be tested, not only for overall interpolation performance but also for computational speed with regard to an intended real-time implementation. For a quantitative performance evaluation, it is further necessary to know the ground truth of the tissues' surface structures used for testing. However, organic tissue seems inappropriate for this purpose. With recent breakthroughs in 3D-printing techniques^{40,41,42}, models of known geometry and similar optical properties as their organic counterparts might be an adequate solution to become an integral part of these future research intentions.

DISCLOSURES

The authors declare that there were no conflicts of interest related to this article.

ACKNOWLEDGMENTS

The authors would like to thank the Micro-Epsilon Messtechnik GmbH & Co. KG for providing the two laser triangulation devices and their very appreciated support regarding all issues of data acquisition and device control. Special thanks go to Dr.-Ing. Armin Schneider from the ARRI Medical GmbH for fruitful discussions on digital microsurgery. The authors received no specific funding for this work.

SUPPLEMENTARY MATERIAL

Supplementary material associated with this article can be found online at url: https://www.lichttechnik.tu-darmstadt.de/forschung_lt/daten_zu_wissenschaftl___veroeffentlichungen/inhalt_mit_marginalienspalte_71.de.jsp.

References

1. Franklin AJ, Sarangapani R, Yin L, Tripathi B, Riemann C. Digital vs analog surgical visualization for vitreoretinal surgery. *Retinal Physician* 2017; 14(May): 34–36 & 38–40.
2. Samy RN. Adoption of new technology. *Current Opinion in Otolaryngology & Head and Neck Surgery* 2016; 24(5): 381.
3. Mayer HM., ed. *Minimally Invasive Spine Surgery: A Surgical Manual*. New York: Springer-Verlag . 2006.
4. Herlan S, Marquardt JS, Hirt B, Tatagiba M, Ebner FH. 3D exoscope system in neurosurgery—Comparison of a standard operating microscope with a new 3D exoscope in the Cadaver Lab. *Operative Neurosurgery* 2019; 17(5): 518–524.
5. Gard N, Rosenthal JC, Jurk S, Schneider A, Eisert P. Image-based measurement by instrument tip tracking for tympanoplasty using digital surgical microscopy. In: Proceedings of SPIE 10951, Medical Imaging 2019: Image-Guided Procedures, Robotic Interventions, and Modeling. The International Society for Optics and Photonics (SPIE); 2019; San Diego, CA, USA: 1095119.

- 332 6. Wisotzky EL, Rosenthal JC, Eisert P, et al. Interactive and multimodel-based augmented reality for remote assistance using
333 a digital surgical microscope. In: Proceedings of the 26th IEEE Conference on Virtual Reality and 3D User Interfaces.
334 IEEE; 2019; Osaka, Japan: 1477–1484.
- 335 7. Aschke M, Wirtz CR, Raczkowsky J, Wörn H, Kunze S. Augmented reality in operating microscopes for neurosurgical
336 interventions. In: Proceedings of the 1st International IEEE EMBS Conference on Neural Engineering. IEEE; 2003; Capri
337 Island, Italy: 652–655.
- 338 8. Khor WS, Baker B, Amin K, Chan A, Patel K, Wong J. Augmented and virtual reality in surgery—the digital surgical
339 environment: applications, limitations and legal pitfalls. *Annals of Translational Medicine* 2016; 4(23): 454.
- 340 9. Lu G, Fei B. Medical hyperspectral imaging: a review. *Journal of Biomedical Optics* 2014; 19(1): 10901.
- 341 10. Wisotzky EL, Arens P, Uecker FC, Hilsmann A, Eisert P. A hyperspectral method to analyze optical tissue characteristics
342 *in vivo*. *International Journal of Computer Assisted Radiology and Surgery* 2018; 13(Suppl 1): 46–47.
- 343 11. Wisotzky EL, Uecker FC, Arens P, Dommerich S, Hilsmann A, Eisert P. Intraoperative hyperspectral determination of
344 human tissue properties. *Journal of Biomedical Optics* 2018; 23(9): 091409.
- 345 12. Wisotzky EL, Kossack B, Uecker FC, et al. Validation of two techniques for intraoperative hyperspectral human tissue
346 determination. In: Proceedings of SPIE 10951, Medical Imaging 2019: Image-Guided Procedures, Robotic Interventions,
347 and Modeling. The International Society for Optics and Photonics (SPIE); 2019; San Diego, CA, USA: 109511Z.
- 348 13. Babilon S, Myland P, Klages J, Simon J, Khanh TQ. Spectral reflectance estimation of organic tissue for improved color
349 correction of video-assisted surgery. *Journal of Electronic Imaging* 2018; 27(5): 053012.
- 350 14. Stigell P, Miyata K, Hauta-Kasari M. Wiener estimation method in estimating of spectral reflectance from RGB images.
351 *Pattern Recognition and Image Analysis* 2007; 17(2): 233–242.
- 352 15. Haneishi H, Hasegawa T, Hosoi A, Yokoyama Y, Tsumura N, Miyake Y. System design for accurately estimating the spectral
353 reflectance of art paintings. *Applied Optics* 2000; 39(35): 6621–6632.
- 354 16. Shimano N, Terai K, Hironaga M. Recovery of spectral reflectances of objects being imaged by multispectral cameras.
355 *Journal of the Optical Society of America A* 2007; 24(10): 3211–3219.
- 356 17. Urban P, Rosen MR, Berns RS. A spatially adaptive Wiener Filter for reflectance estimation. In: Proceedings of the 16th
357 Color and Imaging Conference. Society for Imaging Science and Technology (IS&T); 2008; Portland, OR, USA: 279–284.
- 358 18. Urban P, Rosen MR, Berns RS. Spectral image reconstruction using an edge preserving spatio-spectral Wiener estimation.
359 *Journal of the Optical Society of America A* 2009; 26(8): 1865–1875.
- 360 19. Mansour M, Davidson P, Stepanov O, Piché R. Relative importance of binocular disparity and motion parallax for depth
361 estimation: A computer vision approach. *Remote Sensing* 2019; 11(17): 1990.
- 362 20. Gallup D, Frahm J, Mordohai P, Pollefeys M. Variable baseline/resolution stereo. In: 2008 IEEE Conference on Computer
363 Vision and Pattern Recognition. IEEE; 2008; Anchorage, AK, USA: 1–8.
- 364 21. Scharstein D, Szeliski R, Hirschmüller H. Middlebury Stereo Evaluation – Version 3. [http://vision.middlebury.edu/stereo/
365 eval3/](http://vision.middlebury.edu/stereo/eval3/); . [Online; accessed 13-March-2020].
- 366 22. Niazi ZBM, Essex TJH, Papini R, Scott D, McLean NR, Black MJM. New Laser Doppler Scanner, a Valuable Adjunct in
367 Burn Depth Assessment. *Burns* 1993; 19(6): 485–489.
- 368 23. Ireland AJ, McNamara C, Clover MJ, et al. 3D Surface Imaging in Dentistry – What We are Looking at. *British Dental
369 Journal* 2008; 205(7): 387–392.
- 370 24. Chromy A, Zalud L. Robotic 3D Scanner as an Alternative to Standard Modalities of Medical Imaging. *SpringerPlus* 2014;
371 3: 13.

- 372 25. Micro-Epsilon Messtechnik GmbH & Co. KG . scanCONTROL // 2D/3D laser scanners (laser profile sensors). [https://](https://www.micro-epsilon.com/download/products/cat-scancontrol/dax--scanCONTROL-29x0--en.html#page=2&zoom=Fit)
373 www.micro-epsilon.com/download/products/cat-scancontrol/dax--scanCONTROL-29x0--en.html#page=2&zoom=Fit; .
374 [Online; accessed 17-March-2020].
- 375 26. IDS Imaging Development Systems GmbH . Datenblatt UI-5490SE-M-GL Rev.2. [https://de.ids-imaging.com/IDS/](https://de.ids-imaging.com/IDS/datasheet_pdf.php?sku=AB.0010.1.52802.23)
376 [datasheet_pdf.php?sku=AB.0010.1.52802.23](https://de.ids-imaging.com/IDS/datasheet_pdf.php?sku=AB.0010.1.52802.23); . [Online; accessed 17-March-2020].
- 377 27. Xu D, Jiang Y. A method and implementation of fully digitized continuous microstep for step motor. In: 1997 IEEE
378 International Electric Machines and Drives Conference. IEEE; 1997; Milwaukee, WI, USA: TC2/9.1–TC2/9.3.
- 379 28. Watanabe K, Yokote H. A microstep controller of a DC servomotor. *IEEE Transactions on Instrumentation and Measure-*
380 *ment* 1990; 39(6): 867–869.
- 381 29. Yang Z, Cohen FS. Image registration and object recognition using affine invariants and convex hulls. *IEEE Transactions*
382 *on Image Processing* 1999; 8(7): 934–946.
- 383 30. Chan FY, Lam FK, Zhu H. Adaptive thresholding by variational method. *IEEE Transactions on Image Processing* 1998;
384 7(3): 468–473.
- 385 31. Kamel M, Zhao A. Extraction of binary character/graphics images from grayscale document images. *CVGIP: Graphical*
386 *Models and Image Processing* 1993; 55(3): 203–217.
- 387 32. Suzuki S, Abe K. Topological structural analysis of digitized binary images by border following. *Computer Vision, Graphics,*
388 *and Image Processing* 1985; 30(1): 32–46.
- 389 33. Toussaint GT. Solving geometric problems with the rotating calipers. In: Proceedings of the IEEE Mediterranean
390 Electrotechnical Conference 1983. IEEE; 1983; Athens, Greece: A10.02/1–4.
- 391 34. Carr JC, Beatson RK, Cherrie JB, et al. Reconstruction and representation of 3D objects with radial basis functions. In:
392 Proceedings of the 28th Annual Conference on Computer Graphics and Interactive Techniques. Association for Computing
393 Machinery; 2001; New York, NY, USA: 67–76.
- 394 35. Micchelli CA. Interpolation of scattered data: Distance matrices and conditionally positive definite functions. *Constructive*
395 *Approximation* 1986; 2: 11–22.
- 396 36. Higham NJ. Stability of the diagonal pivoting method with partial pivoting. *SIAM Journal on Matrix Analysis and*
397 *Applications* 1997; 18(1): 52–65.
- 398 37. Bunch JR, Kaufman L. Some stable methods for calculating inertia and solving symmetric linear systems. *Mathematics of*
399 *computation* 1977; 31(137): 163–179.
- 400 38. Babilon S. Raw data of the depth measurements performed on real organic tissue samples. [https://www.lichttechnik.](https://www.lichttechnik.tu-darmstadt.de/forschung_lt/daten_zu_wissenschaftl___veroeffentlichungen/inhalt_mit_marginalienspalte_71.de.jsp)
401 [tu-darmstadt.de/forschung_lt/daten_zu_wissenschaftl___veroeffentlichungen/inhalt_mit_marginalienspalte_71.de.jsp](https://www.lichttechnik.tu-darmstadt.de/forschung_lt/daten_zu_wissenschaftl___veroeffentlichungen/inhalt_mit_marginalienspalte_71.de.jsp);
402 2020. [Online; accessed 16-June-2020].
- 403 39. Wisotzky EL, Rosenthal JC, Eisert P, et al. Interactive and multimodal-based augmented reality for remote assistance using
404 digital surgical microscope. In: 26th IEEE Conference on Virtual Reality and 3D User Interfaces. IEEE; 2019; Osaka, Japan:
405 1477–1484.
- 406 40. Tack P, Victor J, Gemmel P, Annemans L. 3D-Printing Techniques in a Medical Setting: A Systematic Literature Review.
407 *BioMedical Engineering OnLine* 2016; 15: 115.
- 408 41. Lenzen-Schulte M. Anatomische Rekonstruktion – 3-D-Drucker erobern den OP-Saal. *Deutsches Ärzteblatt* 2017; 114(38):
409 1704–1707.
- 410 42. Brunton A, Arikan CA, Tanksale TM, Urban P. 3D Printing Spatially Varying Color and Translucency. *ACM Transactions*
411 *on Graphics* 2018; 37(4): 157.