

Title: The Molecular Epidemiology and Clinical Phylogenetics of Rhinoviruses among Paediatric Cases in Sydney, Australia.

Abstract

Background

Rhinoviruses (RV) represent the most common aetiological agent of all acute respiratory tract infections across all age groups and a significant burden of disease among children. Recent studies have shown that RV-A and RV-C species are associated with varying degrees of disease severity and clinical symptoms.

Methods

In this study, we uncovered potential associations between RV species and subtypes, and clinical disease severity using a matched dataset of 52 RV isolates sampled from children (<18 years) in Sydney, Australia between 2006 and 2009 using epidemiological and phylogenetic methods.

Results

We found that RV-C was significantly more likely to be isolated from paediatric cases under two years of age compared to RV-A, although no significant differences in recorded symptoms were observed. Significant phylogenetic-trait associations between age and the VP4/VP2 capsid protein phylogeny suggests age-specific variations in infectivity among subtypes might also be possible.

Conclusions

This study adds to the growing body of epidemiological evidence concerning RV. Improving surveillance and testing for RV, including routine whole genome sequencing may improve our understanding of the varied disease outcomes of RV species and subtypes. Future studies could aim to identify specific genetic markers associated with age-specific

25 infectivity of RV which could inform treatment practices and public health surveillance of
26 RV.

27 **Keywords:** rhinoviruses, epidemiology, phylogenetics, paediatric infections

28 **List of abbreviations**

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RV	Rhinoviruses
DALY	Disability adjusted life years
USA	United States of America
EV-D68	Enterovirus-D68
5'UTR	5' untranslatable regions
RSV	Respiratory syncytia virus
MCMC	Markov Chain Monte Carlo
PS	Parsimony scores
AI	Association index

31 Background

32 Rhinoviruses (RV) are a highly prevalent and diverse respiratory pathogen of the
33 *Enterovirus* genus and are a major cause of all acute respiratory illnesses throughout the year
34 in both adults and children (1, 2). The attributable burden of RV in both developed and
35 developing countries is significant, accounting for approximately 34.4 disability adjusted life
36 years (DALY) per 1000 population among children under five (3, 4). The direct and in-direct
37 socio-economic burden of RV has also been estimated to cost approximately \$60B/year in the
38 United States of America (USA) alone (5). There are three recognised species of RV denoted
39 RV-A, RV-B, and RV-C, among which there are currently 179 classified subtypes based on
40 sequence homology to former serology-based prototype strains. Since the shift from
41 serology-based to sequence-based typing of RV, our understanding of the RV genome and
42 diversity has increased. For example, RV-C was first recognised as a distinct RV species in
43 2006 (6) but had evaded serology-based detection until molecular methods became routine
44 (7). The transition to sequence-based RV typing has also led to numerous taxonomic changes
45 as old serotypes were regrouped (such as RV-A98 into RV-A54 (8)), and some species re-
46 classified entirely (such as RV-A87 to Enterovirus-D68 (EV-D68) (9, 10)). Despite the sheer
47 diversity of RV, they circulate freely around the world with no geographic structure of either
48 species or subtype (11).

49 Typical symptoms of upper respiratory infection with RV include rhinorrhoea, nasal
50 congestion, cough, headache, and malaise (12). However asymptomatic infection is also
51 prevalent, particularly within households (13). It is now well established that RV can also
52 compromise the lower respiratory tract, in contrast to previous evidence (14), and are
53 sometimes associated with severe disease such as pneumonia and bronchiolitis (15).
54 Exacerbations of asthma and other pre-existing diseases such as chronic obstructive
55 pulmonary disease and cystic fibrosis in both adults and children has furthermore been
56 associated with lower respiratory infection with RV (16). There is increasing (17), but yet
57 inconclusive (18, 19) evidence to suggest RV-A and RV-C species may be associated with
58 more severe disease compared to RV-B strains, and similar efforts have been made to

examine the varied clinical manifestations between RV subtypes, again with inconclusive or contrasting results (20, 21).

In order to better understand potential associations between RV species and clinical features among paediatric cases, we aimed to integrate genetic and epidemiological data using Bayesian phylogenetic methods.

Methods

Compilation of Dataset & Sequencing

We extracted the sequence data of 91 RV isolates collected between 2006 and 2009 in Sydney, Australia from GenBank. Thirty-eight sequences (42%) were originally collected as part of a household transmission study (22) and the other 53 (58%) isolated from a hospital department (23). Among these, 52 RV isolates (28 & 24 from each study respectively) were sampled from paediatric cases defined here as under the age of 18 years. Where available, we matched these sequences to corresponding clinical and demographic data collected during the same period: data on age was available for all 52 paediatric isolates, while clinical data (including symptoms) was only available for 28 sequences. Sequences were first generated according to (23) with relevant detail included here. RNA was extracted directly using LabTurbo Viral DNA/RNA Extraction kits (TaiGen Biotechnology Inc., Taiwan) according to the manufacturer's specifications, and cDNAs synthesized using SuperScript III reverse-transcriptase (Invitrogen, USA). Both 5' untranslatable regions (5'UTR) and VP4/VP2 capsid protein segments were sequenced in both directions using respective forward and reverse primers listed in Supplementary Table S1. Amplicons were purified from agarose gel using QIA PCR purification kits (Qiagen GmbH, Hilden, Germany) and sequenced bi-directionally to confirm specificity in an ABI-3730 XL DNA Analyzer (Applied Biosystem Inc., Foster City, CA).

Characterisation and Epidemiology

Previous phylogenetic analyses found serotype disagreements between the phylogenetic results of RV-A and RV-C species using the 5'UTR region (23). This incongruence can be explained by inter and intra-species recombination, particularly between the 5'UTR segment of RV-A and RV-C (24). We therefore recharacterized isolates by RV species and genotype if possible, primarily using assembled VP4/VP2 sequences (or 5'UTR sequences where VP4/VP2 amplification or assembly failed) with NCBI BLAST. We constructed contingency tables by RV species and symptoms for all 28 paediatric cases with matching clinical data. We considered all 52 paediatric isolates for analysis by age groups here separated as over and under two years of age. We performed Fisher Exact tests to measure statistical associations between age groups and clinical symptoms by RV species using R v3.4.4.

Phylogeny-Trait Association

We confirmed the suitability of both 5'UTR and VP4/VP2 sequence regions for temporal Bayesian phylogenetic analysis by producing preliminary Maximum Likelihood trees using RAxML v8.2 (25) via Geneious v11.1.5 (26) and Tempest v1.5.1 (27) (Supplementary Figure S1). We excluded 17 sequences from the demographic phylogenetic analysis (N = 35) and seven from the clinical phylogenetic analysis (N = 21) since either the 5'UTR and VP4/VP2 sequences were not available. We aligned sequences using MUSCLE v3.8 (28) and used BEAST v1.10.4 (29) to produce posterior phylogeny distributions of both 5'UTR and VP4/VP2 independently and as a multi-locus partition model. We specified a relaxed molecular clock (uncorrelated lognormal prior), constant demographic tree prior, and GTR+I+ Γ_4 nucleotide substitution model as determined per previous studies (24). We ran each model individually for 50 million Markov Chain Monte Carlo (MCMC) generations sampling every 5,000 steps. We checked for parameter convergence and sufficient mixing of the posterior (effective sample size > 200) using Tracer 1.7 (30). For each model, we calculated phylogenetic parsimony scores (PS) (31) association index (AI) (32) statistics as a measure of phylogeny-trait correlation for the matched demographic and clinical symptoms. We used BaTS v0.9.0 (33) to account for statistical uncertainty in each phylogeny across a

posterior distribution of 9,000 trees (after removing 10% burn-in) for each model. As a positive control, we also calculated the PS and AI statistic between serotypes. This study was approved by the UNSW Human Research Ethics Committee (HC17284).

Results

Sequence Characterisation

In Table 1, we show the complete distribution of RV species and subtype characterisation with additional metadata in Supplementary Table S2. Both 5'UTR and VP4/VP2 sequences were available for 35 (67%; N=35/52) isolates while 5'UTR was only available for the remaining 17 (33%; N=17/52) isolates. The majority (92%; N=48/52) of sequences isolated were characterised as RV-A (48%; N=25/52) and RV-C (44%; N=23/52) species. We determined the complete subtype recharacterization for 43 isolates (83%; N=43/52). The majority (78%; N=7/9) of untyped isolates was among the 17 only sequenced for 5'UTR. Overall, RV-C11 was the most frequently detected subtype among characterised isolates (9%; N=4/43) followed by RV-A01 (7%; N=3/43). Just over half of all characterised isolates (51%; N=22/43) were unique i.e. had no common subtype among all isolates characterised.

Clinical and Demographic Epidemiology

Among the 28 RV isolates with matching clinical data, the majority were characterised as RV-A and RV-C. Only a single RV-B isolate had matching clinical data. Cough (96%; N=27/28), nasal congestion (96%; N=27/28) and sneezes (79%; N=22/28) were reported in the majority of cases with matching clinical data (Table 2). All other reported symptoms were less frequent (46% and below).

Cough was absent in a single case of subtype RV-C11 while nasal congestion was absent in a case of RV-C02, however, both subtypes were detected among other cases reporting these symptoms. Diarrhoea was the least reported symptom (14%; N=4/28). The

limited number of RV-B isolates in the dataset precluded inclusion in subsequent statistical comparisons. Differences therefore between RV-A and RV-C species were considered only, however there was no statistically significant difference between clinical symptoms by RV species (Table 2). Among the demographic dataset of 52 RV isolates, 34 cases (65%; $N=34/52$) were under the age of two years. Within species, RV-A was isolated from 13 cases under two years (52%; $N=13/25$), one from RV-B (25%; $N=1/4$), and 20 from RV-C (89%; $N=20/23$). Again, only RV-A and RV-C were tested for significant differences by demographic age group due to low numbers of RV-B isolated from cases under two years, and for consistency with the previous clinical analysis. In this case RV-C was significantly more likely to be isolated from cases under two compared to RV-A ($p = 0.01$).

Phylogenetic Analysis & Trait Association

In each case, our positive control performed as expected, demonstrating significant phylogeny-trait associations by serotype for both PS and AI statistics (Tables 3 and 4). Overall, no clinical symptom demonstrated significant phylogeny-trait associations between or within species. There was a significant association between the VP4/VP2 capsid protein and age group for the PS statistic ($p < 0.01$) but not AI statistic ($p = 0.12$). Figure 1 shows an unrooted maximum clade credibility tree generated from the Bayesian posterior tree distribution coloured by age group.

Discussion

In this study, we report and compare the molecular epidemiology and clinical features of a small sample of paediatric RV cases in Sydney, Australia. Almost all RV species isolated were characterised as RV-A (48%; $N=25/52$) or RV-C (44%; $N=23/52$) with very few RV-B (8%; $N=4/52$) cases observed. This is consistent with many molecular epidemiology studies on RV that demonstrate the predominant circulation of RV-A and RV-C species compared to RV-B (34, 35). We found that RV-C was significantly more likely to be isolated from paediatric cases under two years of age compared to RV-A ($p = 0.01$). This significant difference was also apparent ($p < 0.01$) in one of two Bayesian phylogeny-trait association test statistics (PS not AI) of the VP4/VP2 capsid gene (Tables 3 and 4). More recent studies

have also shown differences in the rates of RV infection by species among early and late childhood (36, 37). Together with our results, this reinforces the potential for age-specific infectivity by RV species, particularly RV-C at younger age groups. The clustering of clinical features within RV subtypes in our dataset may also indicate associations with age of infection beyond the current species classification system, e.g. RV-C02, RV-A01 and RV-A20 in children under two years of age (Figure 1). Future studies could look to identify genetic markers exclusively observed among these subtypes which could be associated with susceptibility at younger ages. For this purpose, larger phylogenetic-trait association studies should be implemented.

In our study we examined a wide range of clinical RV features. The most frequent symptoms reported among the 28 cases with matching clinical data were cough, nasal congestion and sneezes (Table 2). A variety of other symptoms such as loss of appetite, abdominal pain and vomiting were also reported in the minority. Previous studies have reported inconsistent results concerning the clinical presentation and severity between RV species (17-21, 38). For example, in a sample of hospitalised patients in Taiwan, upper respiratory tract infection and nasal congestion was significantly higher in paediatric cases compared to adults (39). In other studies, exacerbations of asthma were often observed among cases of RV-A and RV-C, but not RV-B (40). Furthermore, between species, lower respiratory compromise and pneumonia has been commonly reported among cases of RV-C but less so for RV-A (41). In other molecular epidemiology and phylogenetic studies like ours (42), no significant difference in clinical features were observed between species (Table 2). These contrasting results mean it is difficult to determine conclusively the presence of epidemiological differences in the clinical presentation of RV species. Our approach to explore phylogeny-trait associations aimed to identify potential clustering by clinical traits within RV subtypes, however these results were also not significant (Tables 3 and 4). This suggests that potential variations in symptom presentation might be mostly dependant of individual host-pathogen responses rather than exclusively viral (43). We had a small sample size, and further research with larger samples is required to understand the role these factors play on clinical RV presentation.

Overall this study has a few limitations. Firstly, the small sample size and limited availability of matched clinical data means any significant associations between RV species and clinical features are potentially concealed. We could only find 91 RV isolates, including 52 isolates from paediatric cases in Sydney collected between 2006 and 2009 available in GenBank, however no additional RV sequences were available in Sydney from more recent years. While some differences were observed, e.g. cough was reported among 64% of RV-A cases compared to 31% in RV-C (Table 2) they did not reach significance ($p = 0.13$). Our approach, particularly the unique application of clinical RV phylogeny-trait association testing within a Bayesian phylogenetic framework could be replicated across larger datasets containing matched clinical data to overcome this in the future. Secondly, the clinical features recorded in our study could be attributable to coinfection with other viruses. Although RV continue to represent the most common aetiological agent of all acute respiratory tract infections across all age groups (1, 2), coinfection with other viruses such as respiratory syncytia virus (RSV) is common particularly among children (44). While our study did not determine any significant associations between clinical presentation and RV species or subtype, any future study must consider the potential for any confounding due to viral coinfection. Finally, this study only incorporated the commonly sequenced 5'UTR and VP4/VP2 gene regions into our analysis. Translation of the approximately 7kb RV genome produces a single polyprotein which is cleaved to form mature capsid (VP1-4) and replication (2A-C, 3A-D) peptides (45). Studies have shown that the 5'UTR common among species within the *Enterovirus* genus have significant impacts on viral pathogenesis (46, 47), however differences in clinical manifestation are also attributed to other translatable peptides and genome regions including the 3'UTR (48). Additionally, mutations within the VP1 gene of EV-A71 are important determinants of pathogenesis in mice (49), while VP1 and VP3 are critical for viral attachment of RV to nasal and bronchial epithelia (50). Studies incorporating whole genome sequencing could be used to uncover significant clinical associations between RV strains, with the aim of identifying genetic markers of severity in the future.

Conclusions

223 Our results show that RV-C is significantly more likely to be isolated from paediatric
224 cases under two years of age compared to RV-A despite near equal predominance in
225 circulation. These results were supported by Bayesian phylogenetic-trait association testing
226 of the VP4/VP2 capsid protein suggesting genetic factors may influence the age-specific
227 infectivity of RV species. No statistically significant difference in clinical symptom
228 manifestation was detected between species or subtypes using phylogenetic methods,
229 however the small sample size likely limited statistical power. These results add to the
230 growing body of epidemiological evidence concerning RV. Improving surveillance and
231 testing for RV, including routine whole genome sequencing may improve our understanding
232 of the varied disease outcomes of RV species and subtypes.

233 **References**

- 234 1. Mäkelä MJ, Puhakka T, Ruuskanen O, Leinonen M, Saikku P, Kimpimäki M, et al.
235 Viruses and bacteria in the etiology of the common cold. *Journal of clinical microbiology*.
236 1998;36(2):539-42.
- 237 2. Monto AS. Epidemiology of viral respiratory infections. *The American Journal of*
238 *Medicine*. 2002;112(6, Supplement 1):4-12.
- 239 3. Gaunt ER, Harvala H, McIntyre C, Templeton KE, Simmonds P. Disease burden of
240 the most commonly detected respiratory viruses in hospitalized patients calculated using the
241 disability adjusted life year (DALY) model. *Journal of Clinical Virology*. 2011;52(3):215-21.
- 242 4. Weiss KB, Sullivan SD. The health economics of asthma and rhinitis. I. Assessing the
243 economic impact. *Journal of Allergy and Clinical Immunology*. 2001;107(1):3-8.
- 244 5. Fendrick AM, Monto AS, Nightengale B, Sarnes M. The economic burden of non-
245 influenza-related viral respiratory tract infection in the United States. *Archives of internal*
246 *medicine*. 2003;163(4):487-94.
- 247 6. Lamson D, Renwick N, Kapoor V, Liu Z, Palacios G, Ju J, et al. MassTag
248 polymerase-chain-reaction detection of respiratory pathogens, including a new rhinovirus
249 genotype, that caused influenza-like illness in New York State during 2004–2005. *The*
250 *Journal of infectious diseases*. 2006;194(10):1398-402.
- 251 7. Kistler A, Avila PC, Rouskin S, Wang D, Ward T, Yagi S, et al. Pan-viral screening
252 of respiratory tract infections in adults with and without asthma reveals unexpected human
253 coronavirus and human rhinovirus diversity. *Journal of Infectious Diseases*. 2007;196(6):817-
254 25.
- 255 8. Rathe JA, Liu X, Tallon LJ, Gern JE, Liggett SB. Full-genome sequence and analysis
256 of a novel human rhinovirus strain within a divergent HRV-A clade. *Archives of virology*.
257 2010;155(1):83-7.

258 9. Savolainen C, Blomqvist S, Mulders MN, Hovi T. Genetic clustering of all 102
259 human rhinovirus prototype strains: serotype 87 is close to human enterovirus 70. *Journal of*
260 *General Virology*. 2002;83(2):333-40.

261 10. Blomqvist S, Savolainen C, Råman L, Roivainen M, Hovi T. Human rhinovirus 87
262 and enterovirus 68 represent a unique serotype with rhinovirus and enterovirus features.
263 *Journal of clinical microbiology*. 2002;40(11):4218-23.

264 11. McIntyre CL, Knowles NJ, Simmonds P. Proposals for the classification of human
265 rhinovirus species A, B and C into genotypically assigned types. *Journal of General Virology*.
266 2013;94(8):1791-806.

267 12. Fields BN, Knipe DM, Howley PM. *Fields virology* 6th Edition. Volume 1, Chapter
268 18 - Rhinovirus. Philadelphia: Wolters Kluwer Health/Lippincott Williams & Wilkins; 2013.
269 p. 532 - 50.

270 13. Peltola V, Waris M, Österback R, Susi P, Ruuskanen O, Hyypiä T. Rhinovirus
271 transmission within families with children: incidence of symptomatic and asymptomatic
272 infections. *The Journal of infectious diseases*. 2008;197(3):382-9.

273 14. Papadopoulos NG, Bates PJ, Bardin PG, Papi A, Leir SH, Fraenkel DJ, et al.
274 Rhinoviruses infect the lower airways. *Journal of Infectious Diseases*. 2000;181(6):1875-84.

275 15. Hayden FG. Rhinovirus and the lower respiratory tract. *Reviews in medical virology*.
276 2004;14(1):17-31.

277 16. Varkey JB, Varkey B. Viral infections in patients with chronic obstructive pulmonary
278 disease. *Current opinion in pulmonary medicine*. 2008;14(2):89-94.

279 17. Lee W-M, Lemanske Jr RF, Evans MD, Vang F, Pappas T, Gangnon R, et al. Human
280 rhinovirus species and season of infection determine illness severity. *American journal of*
281 *respiratory and critical care medicine*. 2012;186(9):886-91.

282 18. Huang T, Wang W, Bessaud M, Ren P, Sheng J, Yan H, et al. Evidence of
283 recombination and genetic diversity in human rhinoviruses in children with acute respiratory
284 infection. *PLoS One*. 2009;4(7):e6355.

285 19. Moreira LP, Kamikawa J, Watanabe ASA, Carraro E, Leal É, Arruda E, et al.
286 Frequency of human rhinovirus species in outpatient children with acute respiratory
287 infections at primary care level in Brazil. *The Pediatric infectious disease journal*.
288 2011;30(7):612-4.

289 20. Martin EK, Kuypers J, Chu HY, Lacombe K, Qin X, Strelitz B, et al. Molecular
290 epidemiology of human rhinovirus infections in the pediatric emergency department. *Journal*
291 *of Clinical Virology*. 2015;62:25-31.

292 21. Bruning AH, Thomas XV, van der Linden L, Wildenbeest JG, Minnaar RP, Jansen
293 RR, et al. Clinical, virological and epidemiological characteristics of rhinovirus infections in
294 early childhood: a comparison between non-hospitalised and hospitalised children. *Journal of*
295 *Clinical Virology*. 2015;73:120-6.

296 22. MacIntyre CR, Cauchemez S, Dwyer DE, Seale H, Cheung P, Browne G, et al. Face
297 mask use and control of respiratory virus transmission in households. *Emerg Infect Dis*.
298 2009;15(2):233-41.

299 23. Ratnamohan VM, Zeng F, Donovan L, MacIntyre CR, Kok J, Dwyer DE.
300 Phylogenetic analysis of human rhinoviruses collected over four successive years in Sydney,
301 Australia. *Influenza and Other Respiratory Viruses*. 2016;10(6):493-503.

302 24. Waman VP, Kolekar PS, Kale MM, Kulkarni-Kale UJ. Population structure and
303 evolution of Rhinoviruses. 2014;9(2).

304 25. Stamatakis A. RAxML version 8: a tool for phylogenetic analysis and post-analysis of
305 large phylogenies. *Bioinformatics*. 2014;30(9):1312-3.

306 26. Kearse M, Moir R, Wilson A, Stones-Havas S, Cheung M, Sturrock S, et al. Geneious
307 Basic: an integrated and extendable desktop software platform for the organization and
308 analysis of sequence data. *Bioinformatics*. 2012;28(12):1647-9.

309 27. Rambaut A, Lam TT, Max Carvalho L, Pybus OG. Exploring the temporal structure
310 of heterochronous sequences using TempEst (formerly Path-O-Gen). *Virus Evolution*.
311 2016;2(1):vew007.

312 28. Edgar RCJBb. MUSCLE: a multiple sequence alignment method with reduced time
313 and space complexity. 2004;5(1):113.

314 29. Suchard MA, Lemey P, Baele G, Ayres DL, Drummond AJ, Rambaut A. Bayesian
315 phylogenetic and phylodynamic data integration using BEAST 1.10. *Virus Evolution*.
316 2018;4(1):vey016-vey.

317 30. Rambaut A, Suchard M, Xie D, Drummond A. Tracer v1.6. 2014.

318 31. Fitch WMJSB. Toward defining the course of evolution: minimum change for a
319 specific tree topology. 1971;20(4):406-16.

320 32. Wang T, Donaldson Y, Brettle R, Bell J, Simmonds PJJov. Identification of shared
321 populations of human immunodeficiency virus type 1 infecting microglia and tissue
322 macrophages outside the central nervous system. 2001;75(23):11686-99.

323 33. Parker J, Rambaut A, Pybus OG. Correlating viral phenotypes with phylogeny:
324 accounting for phylogenetic uncertainty. *Infection, Genetics and Evolution*. 2008;8(3):239-
325 46.

326 34. Saraya T, Kimura H, Kurai D, Ishii H, Takizawa HJM. The molecular epidemiology
327 of respiratory viruses associated with asthma attacks: a single-center observational study in
328 Japan. 2017;96(42).

329 35. Arden KE, Greer RM, Wang CYT, Mackay IM. Genotypic diversity, circulation
330 patterns and co-detections among rhinoviruses in Queensland, 2001. 2020;2(1).

331 36. Bochkov Y, Evans M, Grindle K, Pappas T, Lemanske R, Jackson D, et al.
332 Differential Roles of Rhinovirus (RV) A and RV-C Species in Respiratory Illnesses in Early
333 Vs. Late Childhood. Chapter 26: Pediatric Asthma: Epidemiology and Epigenetics 2018. p.
334 A4597-A.

335 37. Luka MM, Kamau E, Adema I, Munywoki PK, Otieno GP, Gicheru E, et al.
336 Molecular epidemiology of human rhinovirus from one-year surveillance within a school
337 setting in rural coastal Kenya. medRxiv. 2020.

338 38. Chen W-J, Arnold JC, Fairchok MP, Danaher PJ, McDonough EA, Blair PJ, et al.
339 Epidemiologic, clinical, and virologic characteristics of human rhinovirus infection among
340 otherwise healthy children and adults: rhinovirus among adults and children. Journal of
341 Clinical Virology. 2015;64:74-82.

342 39. Hung H-M, Yang S-L, Chen C-J, Chiu C-H, Kuo C-Y, Huang K-YA, et al. Molecular
343 epidemiology and clinical features of rhinovirus infections among hospitalized patients in a
344 medical center in Taiwan. Journal of Microbiology, Immunology and Infection.
345 2019;52(2):233-41.

346 40. Zhao M, Zhu W-J, Qian Y, Sun Y, Zhu R-N, Deng J, et al. Association of different
347 human rhinovirus species with asthma in children: a preliminary study. 2016;129(13):1513.

348 41. Linder JE, Kraft DC, Mohamed Y, Lu Z, Heil L, Tollefson S, et al. Human rhinovirus
349 C: age, season, and lower respiratory illness over the past 3 decades. 2013;131(1):69-77. e6.

350 42. Tapparel C, Cordey S, Junier T, Farinelli L, Van Belle S, Soccal PM, et al. Rhinovirus
351 genome variation during chronic upper and lower respiratory tract infections. 2011;6(6).

352 43. Makris S, Johnston S. Recent advances in understanding rhinovirus immunity.
353 F1000Res. 2018;7:F1000 Faculty Rev-537.

- 354 44. Franz A, Adams O, Willems R, Bonzel L, Neuhausen N, Schweizer-Krantz S, et al.
355 Correlation of viral load of respiratory pathogens and co-infections with disease severity in
356 children hospitalized for lower respiratory tract infection. 2010;48(4):239-45.
- 357 45. Palmenberg AC, Rathe JA, Liggett SB. Analysis of Complete Genome Sequences of
358 Human Rhinovirus. The Journal of allergy and clinical immunology. 2010;125(6):1190-201.
- 359 46. Guest S, Pilipenko E, Sharma K, Chumakov K, Roos RP. Molecular mechanisms of
360 attenuation of the Sabin strain of poliovirus type 3. Journal of virology. 2004;78(20):11097-
361 107.
- 362 47. Kawamura N, Kohara M, Abe S, Komatsu T, Tago K, Arita M, et al. Determinants in
363 the 5' noncoding region of poliovirus Sabin 1 RNA that influence the attenuation phenotype.
364 Journal of virology. 1989;63(3):1302-9.
- 365 48. Merkle I, Van Ooij MJ, van Kuppeveld FJ, Glaudemans DH, Galama JM, Henke A, et
366 al. Biological significance of a human enterovirus B-specific RNA element in the 3'
367 nontranslated region. 2002;76(19):9900-9.
- 368 49. Zaini Z, Phuektes P, McMinn P. Mouse adaptation of a sub-genogroup B5 strain of
369 human enterovirus 71 is associated with a novel lysine to glutamic acid substitution at
370 position 244 in protein VP1. Virus research. 2012;167(1):86-96.
- 371 50. Blaas D, Fuchs RJM, pediatrics c. Mechanism of human rhinovirus infections.
372 2016;3(1):1-4.

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375 **Figure Legend**

376 **Figure 1.** Unrooted radial phylogenetic tree of RV-A and RV-C species and subtype coloured by age
377 group with branch lengths equally constrained