

i. **CONFLICT OF INTEREST:** The authors have no conflict of interest to declare

ii. **FINANCIAL SUPPORT:** this work was supported from Children's Hospital Bambino
Gesù, and from Ministry of Health Ricerca Corrente 2021 to NC and PP.

iii. **ABSTRACT (244/250)**

BACKGROUND: Despite SARS-CoV-2 immunizations have started in most countries, children are not currently included in the vaccination programs, thus it remains crucial to define their anti-SARS-CoV-2 immune response in order to minimize the risk for other epidemic waves. This study seeks to provide a description of the virology and anti-SARS-CoV-2 immunity in children with distinct symptomatology.

METHODS: Between March and July 2020, we recruited 15 SARS-CoV-2 asymptomatic (AS) and 51 symptomatic children (SY), stratified according to WHO clinical classification. We measured SARS-CoV-2 viral load using ddPCR and qPCR in longitudinally collected nasopharyngeal swabs samples. To define anti-SARS-CoV-2 antibodies we measured neutralization activity and total IgG load (Diasorin). We also evaluated antigen-specific B and CD8+T-cells, using a labelled S1+S2 protein and ICAM expression, respectively. Plasma protein profiling was performed with Olink.

RESULTS: Virological profiling showed that AS had lower viral load at diagnosis ($p=0.004$) and faster virus clearance ($p=0.0002$) compared to SY. Anti-SARS CoV-2 humoral and cellular response did not appear to be associated with the presence of symptoms. AS and SY showed similar titers of SARS-CoV-2 IgG, levels of neutralizing activity, and frequency of Ag-specific B and CD8+T-cells. Whereas pro-inflammatory plasma protein profile was associated to symptomatology.

CONCLUSION: We demonstrated the development of anti-SARS-CoV-2 humoral and cellular response with any regards to symptomatology, suggesting the ability of both SY and AS to contribute towards herd immunity. The virological profiling of AS suggested that they have lower virus load associated with faster virus clearance.

KEY WORDS: SARS-CoV-2, Asymptomatic, Symptomatic, Ag-specific cellular response, neutralization humoral activity

iv. **MAIN TEXT**

INTRODUCTION

While SARS-CoV-2 immunization programs have started in most countries, the achievement of herd immunity still seems far ahead. Indeed, due to different restrictive measures amongst countries and the lack of vaccination programs for children, the risk for second epidemic waves and health systems overburden remains high. Children infected by SARS-CoV-2 usually present with a milder course of disease compared to COVID-19 adults, with a consistent proportion being fully asymptomatic (AS). Pathogenic reasons underlying such differences have been poorly defined¹. Children play a crucial impact on SARS-CoV-2 transmission and on epidemic waves, especially in school settings²⁻⁴. Test and trace interventions, implemented by government policies for epidemic control may fail in the pediatric population, where AS range between 5 and 16%⁵⁻⁸ and where a consistent proportion remain undiagnosed⁹. In this scenario, some scientific questions arise about AS: i) do they present the same virological features of symptomatic (SY)? ii) do they develop an adaptive and protective immune memory response against SARS-CoV-2? iii) are there inflammatory cytokines profiles associated with clinical manifestations?

49 Several hypotheses have been advanced in the attempt to explain the AS status of SARS-CoV-2
50 infected patients, but no specific investigations on pediatric population are available. The humoral
51 SARS-CoV-2 responses showed a lower level of anti-S IgG in children compared to adults¹⁰,
52 considering both pediatric patients developing multi-inflammatory syndrome associated to COVID-
53 19 (MIS-C) and in those with a milder clinical presentations. Our results in MIS-C ¹¹ showed the
54 absence of pre-existing humoral responses upon other “common cold coronaviruses” in comparison
55 to mild COVID19 children. However, the true influence of pre-existing humoral and T-cell
56 responses towards coronaviruses ¹² on mitigating symptoms in children still need to be defined. In
57 addition, the magnitude of the inflammatory phase associated with viral infection in severe cases ¹³
58 may represent a distinctive feature of AS children compared to SY.

59

60 In the present work, we attempt to define virological and immunological characteristics of 15 AS
61 patients, compared to 51 SY in order to define their ability to produce anti-SARS-CoV-2 immunity.
62 AS and SY patients were further compared to 11 SARS-CoV-2 negative (CoV-2-) patients that
63 were enrolled for suspicion of COVID-19, but that tested negative to both nasopharyngeal swab and
64 serology.

65

66 **METHODS**

67 **Study participants**

68 Sixty-six SARS-CoV-2 infected children (CoV-2+) and 11 SARS-CoV-2 negative controls (CoV-2)
69 were enrolled from March to April 2020 at Bambino Gesù’ Children’s Hospital in Rome for the
70 CACTUS (Immunological studies in Children AffeCTed by COVID and acUte reSpiRatory
71 diseases). The study was approved by local ethical committee and written informed consent was
72 obtained from all participants or legal guardians. Age, gender, clinical and routine laboratory

characteristics are described in Table 1. Inclusion criteria for positive cases was detection of SARS-CoV-2 in nasopharyngeal (NP) swab using SARS-CoV-2 real-time reverse transcriptase-polymerase chain reaction (RT-PCR) tests (GeneXpert, Cepheid, Sunnyvale, CA; 250 copies/mL sensitivity, 100% specificity). Serology was performed as additional confirmatory test using LIAISON® SARS-CoV2 S1/S2 IgG test (DiaSorin, Stillwater, MN, USA). CoV-2+ were stratified according to WHO clinical classification (<https://www.who.int/publications/i/item/WHO-2019-nCoV-clinical-2021-1>) as follow: i) asymptomatic CoV-2+ (AS), patients without any symptoms despite confirmed SARS-CoV-2 infection, that were summoned to the hospital since they belonged to the same nuclear family of symptomatic patients; ii) symptomatic CoV-2+ (SY), patients. We also included 11 children (SARS-CoV-2 negative) with suspected SARS-CoV-2 infection, but tested negative for 2 consecutive NP swabs (performed 24 hours apart) and for SARS-CoV-2 S1/S2 IgG.

Sample collection and storage

Prior to therapy initiation, venous blood was collected in EDTA tubes and processed within 2 hrs. Plasma was isolated from blood and stored at -80°C. Peripheral blood mononuclear cells (PBMCs) were isolated by Ficoll and cryopreserved in liquid nitrogen. NP swab preserving media was stored at -80°C until use. Virological analysis was performed on NP collected at diagnosis and every 48 hours up to virus clearance. Serologic analysis were performed on vials collected at diagnosis and after 10-14 days (herein referred to as “late phase”).

SARS-CoV-2 viral load measurement in swabs by ddPCR

NP swabs were collected by using flocked swabs in liquid-based collection and transport systems. Total nucleic acids were purified from 200µl NP swab preserving media and eluted in a final volume of 100µl. Copies of SARS-CoV-2 were quantified by a home-made multiplex quantitative assay based on One-Step RT-ddPCR, as previously described^{14,15}. Each sample was run at least in duplicate. Results were expressed as SARS-CoV-2 copies/5µl.

98

99

100 **Allplex™ 2019-nCoV Assay**

101 NP swabs were longitudinally collected from CoV-2+ and analyzed using the multiplex Allplex™
102 2019-nCoV Assay (Seegene, Seoul, South Korea) following manufacturer's instructions. The
103 analyzed genes were RdRp and N gene of COVID-19 and E gene of *Sarbecovirus*.

104 **Virus titration by focus forming assay (FFA)**

105 FFA was performed as previously described (Cotugno N. *, Ruggiero A. *, Bonfante F. * [...] Palma P., in
106 press Cell reports). Focus forming units per ml (FFU/ml) were counted after acquisition of pictures at
107 a high resolution of 4800 x 9400dpi, on a flatbed scanner.

108 **Ab-mediated neutralizing activity measured with Plaque reduction neutralization test (PRNT)**

109 A high- throughput PRNT method was developed *in-house*, as described before (Cotugno N. *,
110 Ruggiero A. *, Bonfante F. * [...] Palma P., in press Cell reports). The serum neutralization titer was defined
111 as the reciprocal of the highest dilution resulting in a reduction of the control plaque count >50%
112 (PRNT50). We considered a titre of 1:10 as the seropositive threshold.

113 **Ag-specific B-cells by Flow Cytometry**

114 Ag-specific B-cells were analyzed used a S1+S2 Spike SARS-CoV-2 PE-labelled protein as
115 described before¹⁶⁻¹⁸(Cotugno N. *, Ruggiero A. *, Bonfante F. * [...] Palma P., in press Cell
116 reports). Gating strategy is shown in Fig. 2B. Data analyses were performed using Kaluza software
117 (Beckman Coulter).

118 **Labeling of human recombinant ICAM-1-Fc multimers and CD8 Ag-specific T-cells analysis**
119 **by FACS**

120 Following a previously validated method¹⁹ ICAM-1-Fc were labeled in house with polyclonal anti-
121 human Fc-PE F(ab')₂ fragments. Full labeling and staining protocol details can be found in the
122 Supplementary information file.

123 **Olink assay**

124 Proteins in plasma were analyzed through a multiplexed proximity ligation assay previously
125 described²⁰.

126 **Statistical analysis**

127 Statistical analyses were performed using R software (version 3.6.2) and GraphPad Prism 6
128 (GraphPad Software, Inc., San Diego, CA). Mann-Whitney test and Chi-square test were used to
129 continuous and discrete variables, respectively. D'Agostino-Pearson test was used to assess
130 normality distribution and use the appropriate test. The Area Under the curve (AUC) was calculated
131 with the MESS R library. Plasma proteins were statistically processed as previously described¹¹.
132 Principal Component Analysis (PCA) was performed on proteomics data with the prcomp R
133 function, meanwhile, the PC contribution plot was done using the library factoextra. Statistical
134 significance was set at $p < 0.05$ and the test were two-tailed. Full details for statistical analysis can be
135 found in the supplementary information file.

136

137 **RESULTS**

138 **Study participants**

139 Overall, we analyzed 66 CoV-2+ and 11 CoV-2- children. Our results show that 15/66 (23%) were
140 asymptomatic (AS). No significant differences in terms of age, gender, platelets, white blood cell
141 count (WBC), neutrophils, lymphocytes and hemoglobin (Hb) emerged between symptomatic (SY)
142 and AS (Table 1). WBC and platelets resulted significantly lower in SY compared to CoV-2- while
143 C reactive protein (CRP) resulted significantly higher in CoV-2- compared to the CoV-2+ groups

144 (Table 1). Full spectrum of symptoms and therapy administered to symptomatic patients is shown in
145 Supplementary Table 1.

146

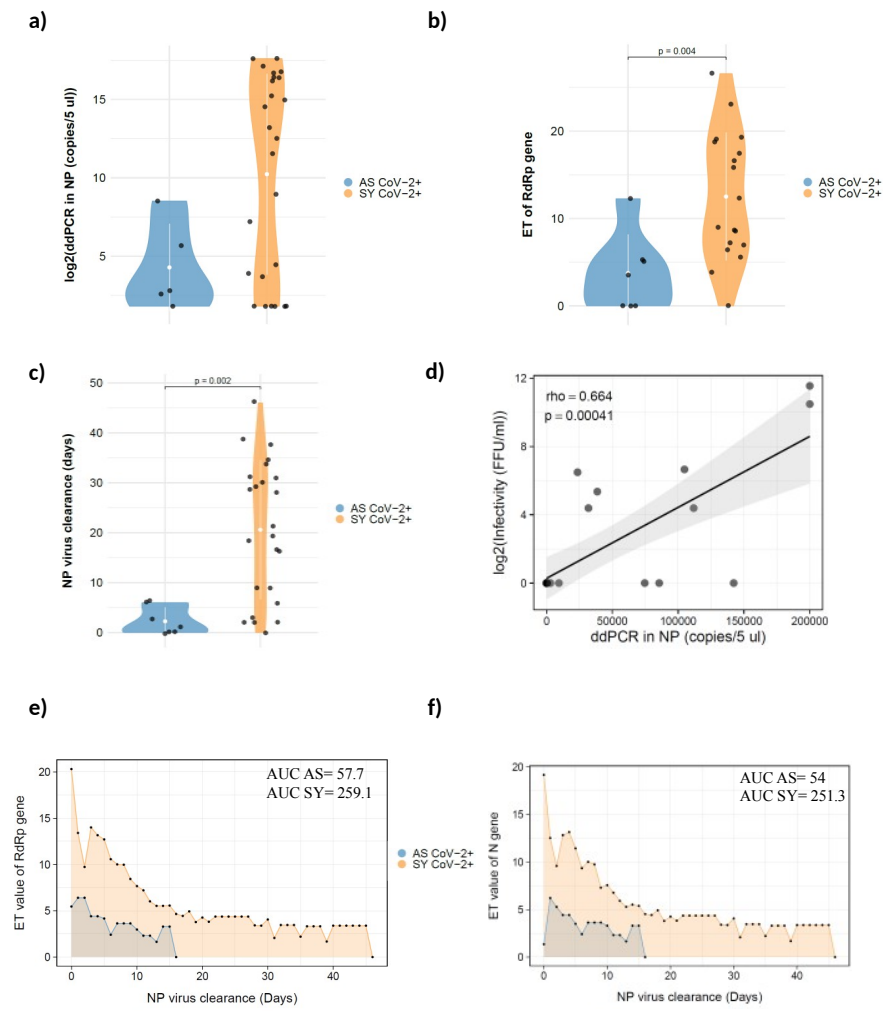
147 **Virologic profiling of AS and SY SARS-CoV-2+**

148 In order to define virological differences according to the symptoms, we measured SARS-CoV-2
149 viral load by digital droplet PCR (ddPCR) and qRT-PCR on NP swabs collected at diagnosis and
150 longitudinally up to first negative test. The analysis revealed a lower viral load at diagnosis in AS vs
151 SY, found statistically significant for RdRp qRT-PCR ($p=0.004$) (Fig.1A and B). The virus
152 clearance expressed in days resulted significantly lower in AS vs SY ($p=0.002$) (Fig. 1C). To define
153 the virus potential of infectivity and its relation to viral load we used a focus forming assay (FFA)
154 and found a positive association with viral load suggesting that higher SARS-CoV-2 loads correlate
155 with higher infectivity potential ($\rho=0.66$; $p=0.0004$) (Fig.1D). The longitudinal analysis
156 performed on NP collected every 48 hours after diagnosis revealed a lower viral area under the
157 curve for both RdRp gene (AUC AS= 57.7; AUC SY= 259.1; $p = 0.08$) and N gene (AUC AS= 54;
158 AUC SY= 251.3; $p = 0.08$) (Fig.1E and F).

159 **Both AS and SY were able to develop specific anti-SARS-CoV-2 humoral and cellular**
160 **responses**

161 To assess whether asymptomatic children presented similar ability to induce protective and
 162 neutralizing humoral response, we quantified serum levels of SARS-CoV-2 specific Ab and Ab-
 163 neutralizing activity at diagnosis and at a “late phase” (10-14 days after diagnosis). No differences
 164 emerged in terms of both SARS-CoV-2 IgG and Ab-mediated neutralization activity (PRNT)
 165 (Figure 2A) between the groups at both time-points. At diagnosis, 5/15 (33%) AS and 17/51 (33%)
 166 SY had developed similar anti-SARS-CoV-2 antibody levels (Fig.2A upper panels). At “late
 167 phase”, the majority of AS and SY had developed neutralizing antibodies (Fig.2A bottom panel,
 168 83% for AS and 84% for SY).

169 We also investigated SARS-CoV-2 specific cellular immunity in peripheral blood prior to any
 170 therapy initiation. We studied Ag-specific B-cells gated on switched memory B-cells (CD10-
 171 CD19+CD27+IgD-), using an in-house fluorescently labeled probe expressing S1+S2 SARS-CoV-
 172 2 proteins (gating strategy in Fig.2B). Ag-specific B-cells were detectable in both AS and SY in



173 similar levels (Figure 2C). Also, the analysis of maturational subsets of CD19+ B-cells through the
174 surface expression of CD10, CD27, IgD and CD21, showed no differences between SY and AS
175 (data not shown).

176 We further assessed the frequency of Ag-specific CD8+T-cells at diagnosis in both groups as
177 previously described¹⁹ (gating strategy in Figure 2D). The frequency of CD8+Ag-specific T-cells
178 was similar between AS and SY children both in terms of frequency (Fig.2E) and absolute counts
179 (not shown). The cytotoxic potential of ICAM+CD8+T-cells, was measured after SARS-CoV-2
180 peptide *in-vitro* stimulation by intracellular production of IL2, TNF-alpha and IFN-gamma.
181 Boolean analysis showed that TNF-alpha positive and TNF-alpha and IL-2 bi-functional
182 ICAM+CD8+T-cells were significantly higher in AS vs SY (p=0.003 and p=0.004 respectively)
183 (Fig.2F) suggesting how these cells in AS maintain an effective antiviral cytotoxic response. No
184 differences in terms of frequency of total CD8+ (Supplementary Figure 1) nor maturational subsets
185 were found among the groups (not shown).

186 **Plasma protein profile of SARS-CoV-2 children**

187 We deepened the characterization of AS and SY by investigating their immunological profiles at
188 admission, using two Olink panels focused on both immune response and inflammation. PCA
189 suggested that proteomic data could only partially define differences between AS and SY (Figure
190 3A), with the top 15 contributing factors including pro-inflammatory cytokines and chemokines
191 (CXCL10, LAMP3) (Fig.3B). We then further explored the levels of expression of each of the 121
192 plasma proteins analyzed, in AS vs SY. Several pro-inflammatory cytokines and chemokines such
193 as CXCL10 and CCL19 results higher in SY compared to AS, conversely others (MCP-4 and IL8),
194 were found lower in SY. Whilst this result confirms the role of inflammatory pathways in inducing
195 severe symptomatology manifestations, it also suggests that the proteomic profiling can only be
196 partially used for the identification of AS patients and that symptomatology severity may be driven
197 by mechanisms not related to inflammation.

198 DISCUSSION

199 This study provides a description of the virological and immunological profiles of 66 SARS-CoV-2
200 infected children with distinct symptomatology. In particular, this work attempted to contribute to
201 the current need for a precise identification of asymptomatic pediatric patients in order to define
202 public restrictive measures. We here investigated viral dynamics, SARS-CoV-2 humoral response
203 and Ag-specific B and CD8 T-cells SARS-CoV-2+. Quantification of SARS-CoV-2 using ddPCR
204 in NP swabs revealed a lower virus load associated with reduced infectivity in AS patients
205 compared to SY. Besides, these data suggested that virus clearance in the upper respiratory system
206 (NP) was faster in AS patients than SY.

207 As previously reported, pediatric population experience milder clinical manifestation resulting in a
208 higher rate of asymptomatic and undiagnosed patients compared to adults^{1,9}. The mechanisms
209 behind such differences are still poorly defined, which renders very difficult the timely
210 identification of AS to prevent virus spread that could fuel further epidemic waves as consequence
211 of school re-openings. Our data suggested that overall AS had lower viral load and associated *in*
212 *vitro* infectivity, alongside capacity to clear the virus faster compared to SY. This difference could
213 suffer of an inescapable bias given by the fact the time of infection cannot be determined, as
214 discussed in other studies^{21,22}. On the other hand, several studies in adults have shown that SARS-
215 CoV-2 load is typically lower after seroconversion^{23,24} underpinning the close relationship between
216 development of humoral response and viral load reduction. In our population, seroconversion rate
217 in AS vs SY at diagnosis was comparable. Whilst this finding partially compensate for the bias
218 discussed above and strengthen the data about AS being less infectious, we acknowledge that
219 further study in bigger population are needed to define the virological characteristics of AS.

220 We then investigated the immunological profiling in relation to symptomatology, showing that both
221 AS and SY were capable to produce Ag-specific B and CD8 T-cells. In a viral respiratory infection,
222 virological control is also maintained thanks to cytotoxic activity of effector CD8+T lymphocytes at

223 the site of infection²⁵. Our results did not identify any difference in terms of maturational subset
224 distribution and frequency between AS and SY SARS-CoV-2 infected children, as previously
225 shown in adults²⁶. On the other hand, our data showed that AS had higher levels of polyfunctional
226 Ag-specific CD8+T-cells compared to SY, demonstrated by the higher frequency of CD8+T-cell
227 producing both TNF-alpha and IL-2 upon *in vitro* stimulation with SARS-CoV-2 peptides.
228 Although this result cannot discriminate whether this is the cause or the effect of symptomatology,
229 it clearly suggests the presence of an effective adaptive immunity in AS. We further evaluated
230 seroconversion rate at a later time point and found no difference between AS and SY. Further, our
231 data confirmed what found in adults ²⁴ showing that approximately 15% were still seronegative at
232 10-14 days after diagnosis, regardless the symptomatology. Our data interestingly suggest that AS
233 have an intact ability to seroconvert, hence contributing to Ab-mediated herd immunity at similar
234 levels compared to SY. In a recent study, similar levels of anti-S IgG were found in
235 paucisymptomatic SARS-COV-2 infected children and COVID19 adults not requiring
236 hospitalization²⁷. However, few uncertainties remain on how the presence of neutralizing antibodies
237 translates in a protected status, since higher antibody concentrations have been observed in adult
238 patients with severe disease than in those with mild disease^{28,29}. In line with this, it must be noted as
239 a suboptimal antibody response could fuel the production of inflammatory cytokines through
240 antibody-dependent enhancement³⁰.

241

242 To define the cytokines profiles of these patients and its association with clinical course of SARS-
243 CoV-2 infection, we analyzed plasma proteome. In a recent work¹¹, we demonstrated that severe
244 COVID-19 manifestations such as the multisystem inflammatory syndrome (MIS-C) were
245 characterized by a specific cytokine storm with unique features as compared to mild COVID-19 and
246 Kawasaki disease. In this patients' cohort, we find that proteomics could only partially discriminate
247 between AS and SY, which may be a reflection of the characteristics of our cohort which lacked of
248 severe cases such as MIS-C. On the other hand, in line with another study in adults³¹, SY appeared

249 to have higher levels of pro-inflammatory cytokines, confirming the role of the cytokines storms in
250 driving severe clinical outcome.

251

252 This study presents some limitations. First, we could only include a small group of the AS, albeit
253 this is a reflection of the clinical reality, which could not be resolved considering that the
254 recruitment included only hospital admissions and not home-assisting surveillance. As previously
255 stated, the infection onset cannot be clearly defined especially in AS children and this could affect
256 the viral load at diagnosis. Further longitudinal studies on larger cohorts with quantitative correlates
257 of viral dynamics and an adult COVID19 cohort for comparison, would be crucial to confirm our
258 observations.

259

260 In conclusion, this study demonstrated that AS has lower viral load and associated *in vitro*
261 infectivity in upper respiratory tract, compared to SY children. Development of both humoral and
262 cell-mediated immunity is not associated with symptomatology, suggesting that importantly AS
263 contribute to achieve herd immunity at similar levels compared to SY. During later time points, the
264 rate of failure in achieving seroconversion is similar in AS and SY: this data may inform alternative
265 diagnostic algorithm to establish mitigated restrictive measures. Additional studies investigating the
266 long-term maintenance of humoral and cell-mediate immunity in these populations are warranted.

267

268 **ACKNOWLEDGMENTS**

269 We would like to thank all patients and guardians who participated to the study and all the
270 CACTUS study nurses team of the COVID-19 Center of “Bambino Gesù Children’s Hospital”. We
271 also thank Jennifer Faudella for her administrative assistance.

272

273 **IMPACT STATEMENT**

274 SARS-CoV-2-infected children present with a milder course of disease compared to COVID-19
275 adults, with a consistent proportion of fully asymptomatic (AS). Such difference in clinical
276 presentations are still poorly defined, even though this population plays a crucial role on SARS-
277 CoV-2 transmission. In this work, we attempted to fill this gap in knowledge by providing a
278 virological and immunological characterization of SARS-CoV-2 infected children with distinct
279 symptomatology. Comparison of SARS-CoV-2 viral load between AS and symptomatic (SY)
280 showed that AS patients had lower viral load at admission and faster virus clearance.
281 Immunological analysis revealed that AS are able to develop SARS-CoV-2 specific adaptive
282 immunity at similar level compared to SY in terms of total SARS-CoV-2 Ab, Ab-mediated
283 neutralization, and Ag-specific B and CD8 T-cells. Conversely, the analysis of plasma profiling
284 showed differences between AS and SY, supporting that pro-inflammatory mechanisms may drive
285 worse clinical outcome in SY. Overall our results show that AS have lower viral load in upper
286 respiratory tract at diagnosis suggesting a lower infectivity potential compared to SY. Furthermore,
287 development of both humoral and cell-mediated immunity is not associated with symptomatology,
288 demonstrating that AS importantly contribute to achieve herd immunity at similar levels compared
289 to SY. This data may inform alternative diagnostic algorithm to establish mitigated restrictive
290 measures for AS.

	<i>SARS-CoV-2 +</i> (N= 66)		<i>SARS-CoV-2 neg</i> (N= 11)	p value
	Asymptomatic (AS, N= 15)	Symptomatic (SY, N= 51)		
Age mean years (SD)	4.6 (3.4)	7.4 (5.6)	6.2 (5.5)	n.s.
Male N (%)	6/15 (40%)	32/51 (63%)	5/11 (45%)	n.s.
Platelets mean 10 ³ /uL (SD)	309.1 (97.3)	293.5 (134.4), n=50	390.9 (201.5)	n.s.
WBC count mean 10 ³ /uL (SD)	7.4 (2.7)	6.4 (2.6)	9.4 (3.8)	SY vs SARS-CoV-2neg p=0.006
Neutrophils (mean 10 ³ /uL (SD)	2.9 (1.4)	2.5 (1.8), n=50	4.8 (3.1)	SY vs SARS-CoV-2neg p=0.003
Lymphocytes mean 10 ³ /uL (SD)	3.9 (1.9)	3.5 (2.2), n=48	3.4 (1.3)	n.s.
Hb mean g/dL (SD)	12.0 (2)	12.8 (1.7), n=50	12.1 (1.4)	n.s.
CRP mean mg/dL (SD)	0.46 (1.3), n=10	1.6 (3.8), n=41	6.5 (6.8), n=11	AS vs SY p=0.01 AS vs SARS-CoV-2neg p=0.0002 SY vs SARS-CoV-2neg p=0.0001

291

292 **Table 1.** Continuous data were presented as mean (SD) calculated on the total number of patients, unless otherwise stated. Mann Whitney test was used for
293 comparison. SD=standard deviation. n.s = not significant. N=number of patients included in this study. n=number of patients available for the analysis.

294 REFERENCES

- 295 1 Brodin, P. Why is COVID-19 so mild in children? *Acta paediatrica* **109**, 1082-1083,
296 doi:10.1111/apa.15271 (2020).
- 297 2 Panovska-Griffiths, J. *et al.* Determining the optimal strategy for reopening schools, the impact of
298 test and trace interventions, and the risk of occurrence of a second COVID-19 epidemic wave in the
299 UK: a modelling study. *The Lancet. Child & adolescent health* **4**, 817-827, doi:10.1016/S2352-
300 4642(20)30250-9 (2020).
- 301 3 Head, J. R. *et al.* The effect of school closures and reopening strategies on COVID-19 infection
302 dynamics in the San Francisco Bay Area: a cross-sectional survey and modeling analysis. *medRxiv :
303 the preprint server for health sciences*, doi:10.1101/2020.08.06.20169797 (2020).
- 304 4 Esposito, S., Cotugno, N. & Principi, N. Comprehensive and safe school strategy during COVID-19
305 pandemic. *Italian journal of pediatrics* **47**, 6, doi:10.1186/s13052-021-00960-6 (2021).
- 306 5 Parri, N., Lenge, M., Buonsenso, D. & Coronavirus Infection in Pediatric Emergency Departments
307 Research, G. Children with Covid-19 in Pediatric Emergency Departments in Italy. *The New England
308 journal of medicine* **383**, 187-190, doi:10.1056/NEJMc2007617 (2020).
- 309 6 Lu, X. *et al.* SARS-CoV-2 Infection in Children. *The New England journal of medicine* **382**, 1663-1665,
310 doi:10.1056/NEJMc2005073 (2020).
- 311 7 Lu, X., Xiang, Y., Du, H. & Wing-Kin Wong, G. SARS-CoV-2 infection in children - Understanding the
312 immune responses and controlling the pandemic. *Pediatric allergy and immunology : official
313 publication of the European Society of Pediatric Allergy and Immunology* **31**, 449-453, doi:10.1111/
314 pai.13267 (2020).
- 315 8 Dong, Y. *et al.* Epidemiology of COVID-19 Among Children in China. *Pediatrics* **145**,
316 doi:10.1542/peds.2020-0702 (2020).
- 317 9 Li, R. *et al.* Substantial undocumented infection facilitates the rapid dissemination of novel
318 coronavirus (SARS-CoV-2). *Science* **368**, 489-493, doi:10.1126/science.abb3221 (2020).
- 319 10 Weisberg, S. P. *et al.* Distinct antibody responses to SARS-CoV-2 in children and adults across the
320 COVID-19 clinical spectrum. *Nature immunology* **22**, 25-31, doi:10.1038/s41590-020-00826-9
321 (2021).
- 322 11 Consiglio, C. R. *et al.* The Immunology of Multisystem Inflammatory Syndrome in Children with
323 COVID-19. *Cell* **183**, 968-981 e967, doi:10.1016/j.cell.2020.09.016 (2020).
- 324 12 Grifoni, A. *et al.* Targets of T Cell Responses to SARS-CoV-2 Coronavirus in Humans with COVID-19
325 Disease and Unexposed Individuals. *Cell* **181**, 1489-1501 e1415, doi:10.1016/j.cell.2020.05.015
326 (2020).
- 327 13 Peng, Y. *et al.* Broad and strong memory CD4(+) and CD8(+) T cells induced by SARS-CoV-2 in UK
328 convalescent individuals following COVID-19. *Nature immunology* **21**, 1336-1345,
329 doi:10.1038/s41590-020-0782-6 (2020).
- 330 14 Corman, V. M. *et al.* Detection of 2019 novel coronavirus (2019-nCoV) by real-time RT-PCR.
331 *Eurosurveillance* **25**, doi:10.2807/1560-7917.ES.2020.25.3.2000045 (2020).
- 332 15 Rampazzo, E. *et al.* Telomere length and telomerase levels delineate subgroups of B-cell chronic
333 lymphocytic leukemia with different biological characteristics and clinical outcomes. *Haematologica*
334 **97**, 56-63, doi:10.3324/haematol.2011.049874 (2012).
- 335 16 Cotugno, N. *et al.* Early antiretroviral therapy-treated perinatally HIV-infected seronegative children
336 demonstrate distinct long-term persistence of HIV-specific T-cell and B-cell memory. *Aids* **34**, 669-
337 680, doi:10.1097/QAD.0000000000002485 (2020).
- 338 17 Cotugno, N. *et al.* Artificial Intelligence Applied to in vitro Gene Expression Testing (IVIGET) to
339 Predict Trivalent Inactivated Influenza Vaccine Immunogenicity in HIV Infected Children. *Frontiers in
340 immunology* **11**, 559590, doi:10.3389/fimmu.2020.559590 (2020).
- 341 18 Cotugno, N. *et al.* Higher PIK3C2B gene expression of H1N1+ specific B-cells is associated with lower
342 H1N1 immunogenicity after trivalent influenza vaccination in HIV infected children. *Clinical
343 immunology* **215**, 108440, doi:10.1016/j.clim.2020.108440 (2020).

- Dimitrov, S. *et al.* Activated integrins identify functional antigen-specific CD8(+) T cells within minutes after antigen stimulation. *Proceedings of the National Academy of Sciences of the United States of America* **115**, E5536-E5545, doi:10.1073/pnas.1720714115 (2018).
- Lundberg, M. *et al.* Multiplexed homogeneous proximity ligation assays for high-throughput protein biomarker research in serological material. *Molecular and Cellular Proteomics* **10**, doi:10.1074/mcp.M110.004978 (2011).
- Meyerowitz, E. A., Richterman, A., Bogoch, II, Low, N. & Cevik, M. Towards an accurate and systematic characterisation of persistently asymptomatic infection with SARS-CoV-2. *The Lancet. Infectious diseases*, doi:10.1016/S1473-3099(20)30837-9 (2020).
- Ooi, E. E. & Low, J. G. Asymptomatic SARS-CoV-2 infection. *The Lancet. Infectious diseases* **20**, 996-998, doi:10.1016/S1473-3099(20)30460-6 (2020).
- Sayampanathan, A. A. *et al.* Infectivity of asymptomatic versus symptomatic COVID-19. *Lancet* **397**, 93-94, doi:10.1016/S0140-6736(20)32651-9 (2021).
- Peeling, R. W. *et al.* Serology testing in the COVID-19 pandemic response. *The Lancet. Infectious diseases* **20**, e245-e249, doi:10.1016/S1473-3099(20)30517-X (2020).
- Zheng, H. Y. *et al.* Vol. 17 541-543 (Springer Nature, 2020).
- Qin, C. *et al.* Dysregulation of immune response in patients with coronavirus 2019 (COVID-19) in Wuhan, China. *Clinical Infectious Diseases* **71**, 762-768, doi:10.1093/cid/ciaa248 (2020).
- Weisberg, S. P. *et al.* Distinct antibody responses to SARS-CoV-2 in children and adults across the COVID-19 clinical spectrum. *Nature immunology* **22**, doi:10.1038/s41590-020-00826-9 (2020).
- Zhao, J. *et al.* Antibody Responses to SARS-CoV-2 in Patients with Novel Coronavirus Disease 2019. *Clinical Infectious Diseases* **71**, 2027-2034, doi:10.1093/cid/ciaa344 (2020).
- Peeling, R. W. *et al.* Vol. 20 e245-e249 (Lancet Publishing Group, 2020).
- Iwasaki, A. & Yang, Y. The potential danger of suboptimal antibody responses in COVID-19. *Nature reviews. Immunology* **20**, 339-341, doi:10.1038/s41577-020-0321-6 (2020).
- Arunachalam, P. S. *et al.* Systems biological assessment of immunity to mild versus severe COVID-19 infection in humans. *Science* **369**, 1210-1220, doi:10.1126/science.abc6261 (2020).

386 **FIGURE LEGEND**

387 **Figure 1. Virological analysis of SARS-CoV-2 infected children.** (A) Log_2 (ddPCR in NP swabs
388 (copies/5 μl) in AS vs SY. (B) ET of RdRp gene (C) NP virus clearance (days) in AS vs SY (D)
389 Association between infectivity (measured as FFU/ml) and ddPCR in NP swabs (E) AUC for RdRp
390 gene in AS vs SY (F) AUC for N gene in AS vs SY. Mann Whitney test were used to define
391 differences in (A)-(B)-(C) were considered significant; Spearman's test was used in (D), AUC was
392 calculated as described in methods. p values lower than 0.05 were considered significant.

393 **Figure 2. SARS-CoV2 seroconversion and Ag-specific B and CD8 T-cells in in children with**
394 **distinct symptomatology.** (A) Left-hand side represents SARS-CoV-2 Ab titers at admission
395 (upper dot plots) and at "late phase" (lower dot plots). Right-hand side represents SARS-CoV-2
396 PRNT at admission (upper dot plots) and at "late phase" (lower dot plots). Neutralization values
397 were identified by dilution factor. Contingency plots show frequency of seronegative, seropositive
398 and equivocal results in both phases, following the same pattern described for the dot plots. Gating
399 strategy (B) and frequencies of Ag-specific CD19+IgD-CD27+switched B-cells (C) are shown in
400 AS and SY. Gating strategy (D) and frequencies of Ag-specific ICAM+ CD8+T-cells (E) are shown
401 in AS and SY. Box plot in (F) shows results from Boolean gating of intracellular staining analysis
402 from ICAM+ CD8 T cells. Mann Whitney test were used to define differences, p values lower than
403 0.05 were considered significant.

404 **Figure 3. Proteomic profile in AS vs SY.** PCA in (A) shows distribution according to protein
405 profile between AS and SY. (B) PC loading for PC4 is showed in the box plot. The red line
406 indicates the expected values. Violin plots in (C) shows differentially expressed proteins among the
407 groups.

408

409