

**Supporting Information**

**Impacts of hydrodynamic conditions and surface roughness on the critical condition to develop and thickness of *Pseudomonas putida* biofilms**

*Guanju Wei*<sup>1,2</sup>, *Judy Q. Yang*<sup>1,2\*</sup>

<sup>1</sup>Saint Anthony Falls Laboratory, University of Minnesota, Minneapolis MN, USA; <sup>2</sup>Department of Civil, Environmental, and Geo-Engineering

Corresponding Author:

\*Judy Q. Yang (judyyang@umn.edu)

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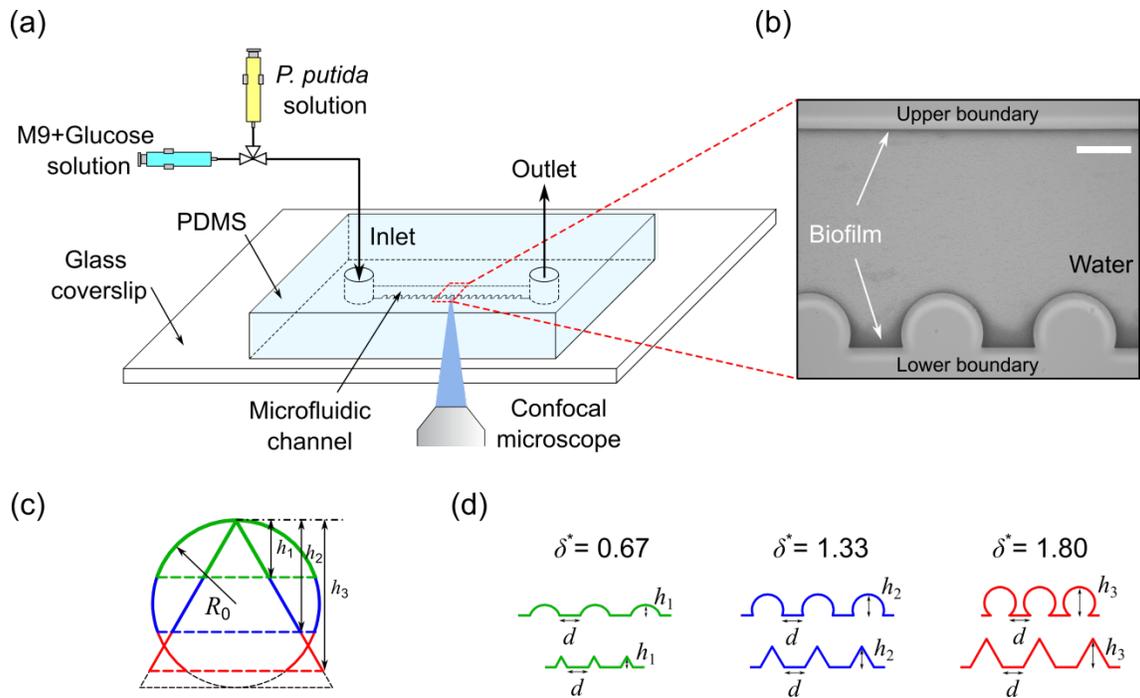
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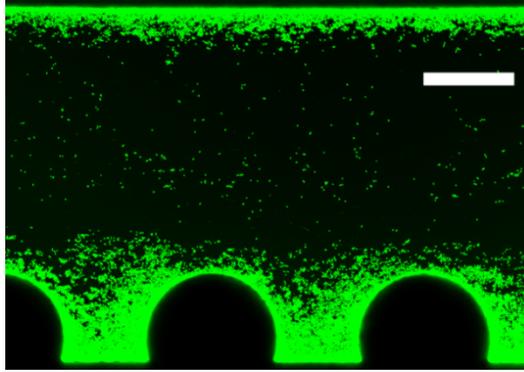
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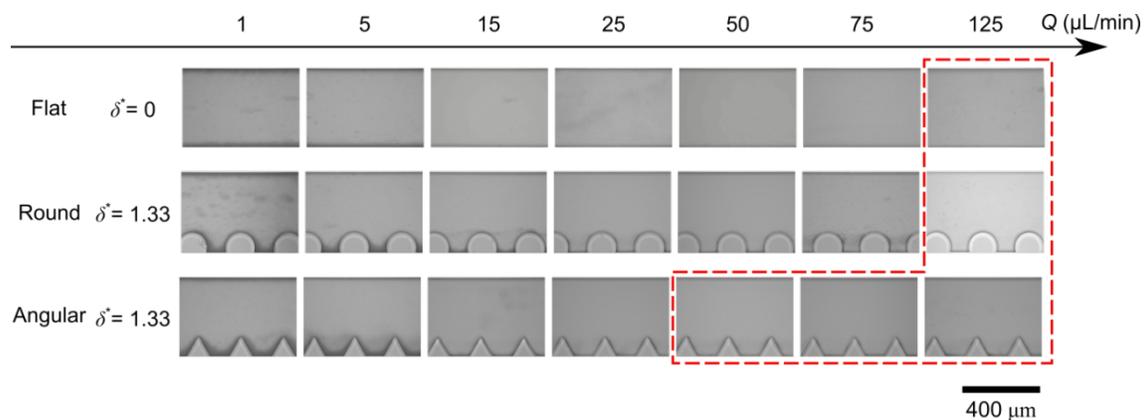
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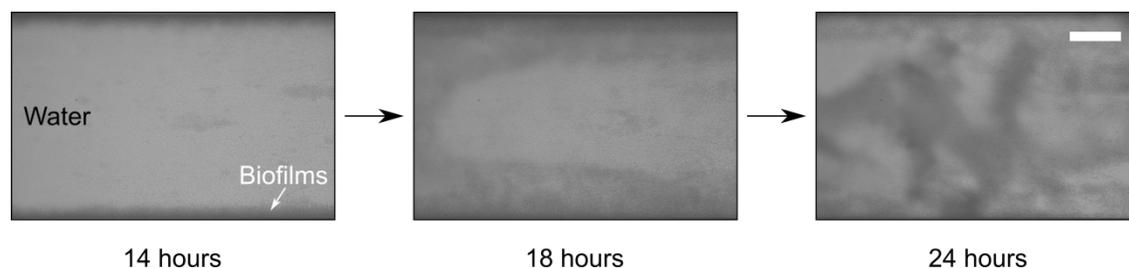
**Figure S1.** (a) Schematic diagram of the experimental set up. (b) Cross-sectional image of the microfluidic channel. Biofilms (gray color) accumulate at the upper and lower boundaries of the channel. The scale bar is 100  $\mu\text{m}$ . (c) The relative roughness height  $\delta^*$  is defined as  $h/R_0$ .  $R_0 = 75$   $\mu\text{m}$  denotes the radius of the circle and half of the triangle height.  $h$  denotes the height of each roughness on the surface:  $h_1 = 50$   $\mu\text{m}$ ,  $h_2 = 100$   $\mu\text{m}$ ,  $h_3 = 135$   $\mu\text{m}$ . (d) Six types of rough surfaces were used in this study with three relative heights ( $\delta^* = 0.67, 1.33, 1.80$ ) and two types of roughness shape (round and angular) at the lower boundary. The distance between the neighboring roughness elements for all cases is the same, i.e.,  $d = 100$   $\mu\text{m}$ .



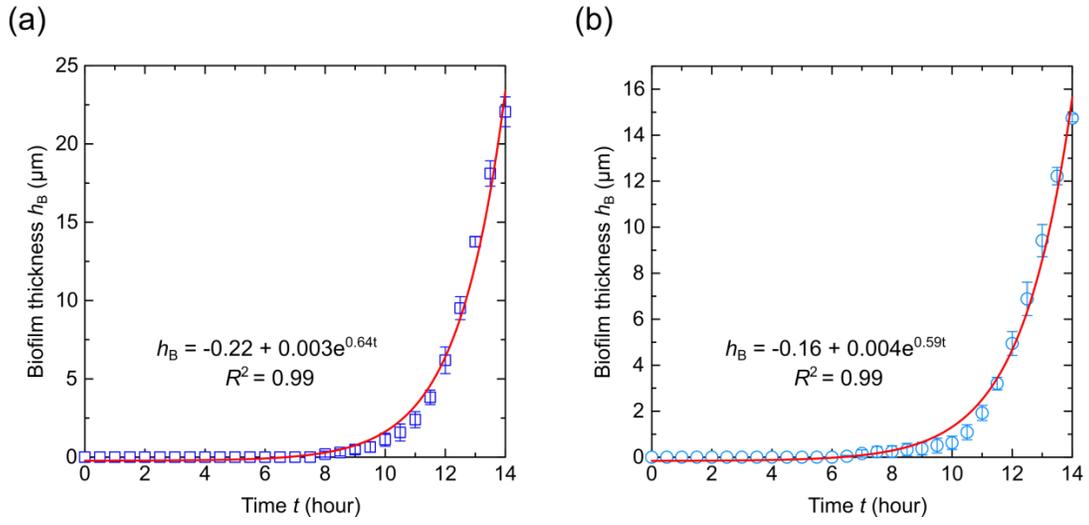
**Figure S2.** Confocal image of the biofilm stained with EPS dyes after the 14-hour experiments at flow rate of 5  $\mu\text{L}/\text{min}$ ,  $\delta^* = 1.33$ . EPS is visualized with green fluorescent dyes<sup>1</sup>. The EPS stains included 5  $\mu\text{M}$  of SYTO-9 green fluorescent nucleic acid stain (Thermo Fisher Scientific, USA), 20  $\mu\text{g}/\text{mL}$  fluorescein isothiocyanate (FITC) conjugated Concanavalin A from *Canavalia ensiformis* (Sigma), and 20  $\mu\text{g}/\text{mL}$  FITC conjugated lectin from *Triticum vulgaris* (Sigma). The scale bar is 100  $\mu\text{m}$ .



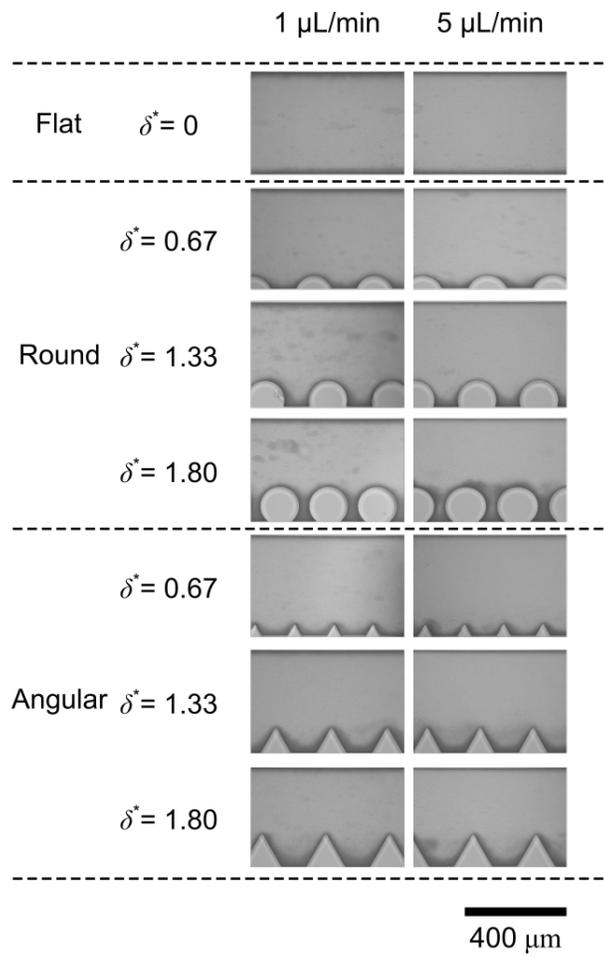
**Figure S3.** Cross-sectional images of the biofilms at 14-hour growth time in channels with flat boundaries and boundaries with round and angular roughness (both with roughness relative height  $\delta^* = 1.33$ ) at seven flow rates. Red dashed line represents cases with no biofilm growth.



**Figure S4.** At low-flow conditions ( $<5 \mu\text{L}/\text{min}$ ), biofilms in microfluidic channels continue growing after 14 hours and start to clog the channel after 24 hours. The scale bar is 100  $\mu\text{m}$ .



**Figure S5.** The biofilm growth follows exponential law under low flow rates:  $1 \mu\text{L}/\text{min}$  (a) and  $5 \mu\text{L}/\text{min}$  (b).



**Figure S6.** Cross-sectional images of the biofilms at 14-hour growth time in channels with flat boundaries and boundaries with round and angular roughness at two different flow rates and three roughness heights.

**Table S1.** Parameters used in COMSOL simulation.

Flow conditions	Laminar flow
	Incompressible flow
Density	995.6 kg/m <sup>3</sup>
Dynamic viscosity	0.001 Pa·s
Average inflow velocity	0.09 m/s
	0.05 m/s
	0.03 m/s
	0.02 m/s
	0.01 m/s
	0.003 m/s
	0.0007 m/s
Temperature	303.15 K

**Table S2.** The 90% confidence interval of critical shear stress  $\tau_{crit}$ ,  $p$  value, and  $R^2$  were calculated by regression analysis using MATLAB.

	90% confidence interval	$p$	$R^2$
Flat	0.19 ~ 0.48	$1.0 \times 10^{-3}$	0.95
Round	0.59 ~ 1.44	$1.1 \times 10^{-4}$	0.96
Angular	0.61 ~ 1.84	$2.2 \times 10^{-4}$	0.95

### References

(1) Drescher, K.; Shen, Y.; Bassler, B.L.; Stone, H.A. Biofilm streamers cause catastrophic disruption of flow with consequences for environmental and medical systems. *Proceedings of the National Academy of Sciences*. 2013, *110*(11), 4345-4350.