

# Biological Prescience: The Role of Anticipation in Organismal Design

C. Deans<sup>1\*</sup>

<sup>1</sup>University of Minnesota, 219 Hodson Hall, 1980 Folwell Ave, St. Paul, MN 55108

\*Correspondence to: Carrie Deans, dean0179@umn.edu

## Abstract

Anticipation is the act of using information about the past and present to make predictions about future scenarios. As a concept, it is predominantly associated with the psychology of the human mind; however, there is accumulating evidence that diverse taxa without complex neural systems, and even biochemical networks themselves, can respond to perceived future conditions. Although predictive biological responses, such as circadian rhythms and stress preconditioning, have been extensively studied over the last century, newer research on predictive genetic networks in microbes have highlighted the existence of anticipatory processes in older, more primitive taxa. When taken together, these seemingly disparate processes can be connected based on a shared anticipatory design. This review discusses the important and often overlooked role of anticipation in the evolution of biological designs, and advocates for establishing a fundamental distinction between anticipatory and reactive processes as an effort to advance our understanding of complex biological processes.

## Keywords

predictive, cephalic responses, anticipatory regulation, feed-forward control, allostasis, microbes, Robert Rosen

## Introduction

The concept of anticipation is deeply rooted in the foundations of science. It lives at the intersection of ideas on time, change, perception, and causality, which encompass some of the oldest philosophical contemplations. As such, it has been addressed in a multitude of different fields, including gnoseology and epistemology (Burgers, 1975; Rosen, 1985a), mathematics (Rosen, 1978; Zadeh, 1978; Dubois, 2003), physics (Feynman, 1982; Whitehead, 1929), economics (King, 1938; Klir, 2002), computer science (Feynman, 1982; Svoboda, 1960; Zadeh, 2001), biology (Rosen, 1985a, 1985b; Libet et al., 1983; Libet, 1985), and psychology (Hoffmann et al., 2007; Butz, 2008; Bubic et al., 2010). Despite its recent popularity in the computer sciences, particularly in information theory, algorithm development, and machine learning, the concept of anticipation is perhaps most widely discussed in the field psychology. This is not surprising given that anticipation is an everyday feature of human thought and one with undeniable value. It is so highly regarded, in fact, that the ability to anticipate with a high degree of accuracy is widely regaled as a supernatural feat by cultures all over the world. Although anticipation has been studied in non-human organisms, perhaps most famously by Pavlov's work on classical conditioning in canines, the reality is that you would be hard-pressed to find a discussion of it in the broader biological sciences, particularly outside of research on cognition. Increasing evidence, however, shows that anticipatory processes are not simply reserved for those organisms with brains or complex neural systems. Anticipatory genetic networks have been documented in yeast and bacterial species (Shudo et al., 2003; Tagkopoulos et al., 2008; Mitchell et al., 2009), while other putative anticipatory processes can be found in plants (Karban et al., 2003; Engelberth et al., 2004; Kessler et al., 2006; Falik et al., 2011) and archaea (Whitehead et al., 2009). Even simple biochemical networks can themselves display anticipatory properties (Tagkopoulos et al., 2008; Mitchell et al., 2009; Freddolino and Tavazoie, 2012). These findings suggest that anticipation is not a pinnacle of evolutionary innovation but rather a fundamental biological mechanism that likely evolved very early on, and contrary to human superstition, is not at all supernatural but rather entirely natural (Westerhoff et al., 2014; Calvo and Baluška, 2015; Novoplansky, 2016).

If anticipatory processes, i.e., those possessing predictive regulatory structures, are so prevalent, why then, is anticipation such an elusive concept in the broader biological sciences? Despite many potential reasons for this, it isn't from a lack of consideration. The concept of anticipation, as it relates to biology, has indeed been studied by many academics. One of the most thoughtful and prolific writers on the subject is Robert Rosen. His seminal book *Anticipatory Systems* (Rosen, 1985) provided a rigorous epistemological and mathematical justification for the existence and importance of anticipatory systems. Rosen asserted that anticipation had been unfairly overlooked in modern science due to the field's preoccupation with reductionism and Newtonian physics; a paradigm that only allows present states to be dependent on past or current states, but never future states. Rosen believed that this restriction was empirically and philosophically erroneous and ultimately prevented modern science from ever being able to explain complex biology systems. As a student of relational biology, which would later produce the field of systems biology, he believed that focusing on biological organization was essential.

Despite these philosophical conflicts, a substantial body of work on the theoretical aspects of anticipation can be found in the literature. Unfortunately, explicit links between this work and empirical biological data are rare. That is not to say that examples of biological responses possessing anticipatory characteristics have gone unidentified. They have simply been

described without a clear association to anticipation. Evidence of circadian rhythms was first discovered in the early 1700s (Albrecht, 2010; Bhadra et al., 2017), the biphasic dose-response curve in the late 1800s (Calabrese, 2002), and cephalic digestive responses at the turn of the 20<sup>th</sup> century (Pavlov, 1902; Powley, 1977; Smith, 1995; Power and Schulkin, 2008). These, and many others putative anticipatory responses, have been known for quite some time but have been studied in isolation with little reason to connect them to each other based on a shared relationship to anticipation. This is perhaps not surprising, as forthcoming examples will show that it is very difficult to discern between purely reactive processes and those operating in an anticipatory context, particularly when a biologically-explicit framework does not exist. It requires rigorous experimentation and the integration of detailed ecological, physiological, and molecular information, which is often unavailable. In many cases, anticipatory responses are routinely misidentified as purely reactive processes based on logical, yet often circumstantial, evidence. For instance, one would never expect gastric enzymes to be secreted before the ingestion of food, as is done in cephalic responses, because their function is so clearly dependent on a dietary substrate? It would seem illogical to expect, let alone, test for such a phenomenon. Moreover, the propensity to search for such phenomena is made even less likely by the presiding and pervasive assumption that biochemical reactions are always unidirectional and sequential, which is why most of the responses that can now be described as anticipatory have been, up to this point, viewed as biological outliers. Despite this, recent empirical data are improving our understanding of these processes and making their identification easier.

This review unites new and old data on anticipatory processes to highlight the important role that they have played in the evolution of biological designs. This synthesis shows that anticipatory processes are more primitive and ubiquitous than previously thought; however, the objective of this article is not to simply show that anticipation exists in basic biological processes, but that it is common. Doing this requires establishing a biologically-explicit anticipatory framework that satisfies three basic criteria. First, anticipatory processes must be defined in a way that clearly distinguishes them from other processes (what will be termed reactive processes). Second, evidence for the existence of these processes must be demonstrated within an evolutionary context. Third, a justification for the incorporation of an anticipatory framework into the current biological paradigm must be given, including an appraisal of how it advances our understanding of complex biological processes. These three criteria closely follow the structure of this article. The first section discusses the causal aspects of anticipation and discusses key considerations at different scales. The second section provides examples of different types of anticipatory processes, particularly in non-cognitive organisms. The last section explores the potential mechanisms used in anticipatory responses, discusses the direction of future research, and highlight how an anticipatory framework can improve our understanding of complex biological processes.

## **What is Biological Anticipation?**

In his book *Anticipatory Systems* (1985), Robert Rosen defines an anticipatory system as one whose current state depends upon a future state, and which occurs when a system contains a model of itself that unfolds in faster than real time. Since then, many have made adjustments to Rosen's definition. Dubois and Resconi's (1992) and Nadin's (2010; 2012) amendments asserted that anticipatory systems can take into account past, current, and future states, as well as possible future states, which highlights the importance of predictive models with no contingencies.

Making predictions about future states does not require any information about the future, but instead can arise from predictive models that incorporate information about the past and present to form a probabilistic forecast of future scenarios. Because this predictive process occurs in real-time and is in no way contingent on the accuracy of the model, anticipation can clearly be established in the realm of deterministic science. It is important to note that the terms anticipation and prediction will be used synonymously throughout this article. Some of the phenomena discussed may be described by other authors as “predictive”, but there is certainly no need to create unnecessary semantical obstacles in the discussion of like topics. The term “anticipatory”, however, has been specifically selected instead of “predictive” to describe the aforementioned biological processes because its connotation emphasizes a purposiveness that has clear biological and evolutionary relevance. This language also emphasizes the importance of the predicted state over the potentially inconsequential act of making a prediction.

In the context of cognition, anticipation is not controversial, as human beings make predictions on a daily basis. However, the notion that non-cognitive organisms or even biochemical networks can operate in a predictive manner is arguably more provocative. This is especially true given that the language associated with anticipation and prediction is so closely tied to the process of thinking. But what is the process of thinking if not a network of biochemical reactions, subject to the same evolutionary constraints as any other process? Acknowledging the biochemical nature of cognition makes the prospect of more primitive anticipatory responses more acceptable. To truly understand the different facets of anticipation in a biological context, we must start with clear causal principles and identify how these principles operate at different levels of biological organization.

### *Causal Considerations*

If anticipatory systems are those whose current state is in some way dependent on a future state, then a more biologically-explicit definition would describe anticipatory biological processes as those operating in the present to provide an anticipated future function. Here, biological processes are viewed as systems, sometimes anticipatory systems, but it is important to note that living systems are different in many ways from the non-living systems. Most notably, biological processes are subject to natural selection, and this narrows the conditions under which anticipatory biological processes can develop, constrains their designs, and places conditions on the utility of these processes. The implications of these factors will be discussed in more detail below. On a broader scale, however, anticipatory processes can be defined by the role that time and information play in these responses. A fundamental aspect of anticipation is time; thus, any definition of anticipatory systems or processes must contain a temporal element. The definition above makes a causal connection between processes occurring in the present and their dependence on a predicted future state. This is in contrast to processes that occur in the present but are only dependent on events that have already happened. This temporal distinction provides a key division between anticipatory processes and non-anticipatory, or what will be termed reactive, processes.

Information provides another defining factor, as the prediction of a future state is ultimately dependent on it. Information theory describes the relationship between information and uncertainty. The greater the possible states a system is capable of being in, the greater the uncertainty surrounding its state at any given time. Acquiring information about the system or anything that affects it can reduce this uncertainty. In a reactive system that operates based on what has already happened, there is little uncertainty about what is required. However, an anticipatory system operates based on a potential future state that can only be inferred from past

and current information, which produces more inherent uncertainty. Why then, would anticipatory processes be advantageous? The answer may have something to do with the fact that uncertainty is an unavoidable feature of life due to the interaction that living things have with their environment. The majority of biological functions are geared toward maintaining growth and reproduction in variable environments, thus environmental variability represents a considerable amount of fitness-relevant uncertainty. There is, therefore, strong evolutionary pressure to reduce this uncertainty through the acquisition of information, which is evidenced by the existence of diverse sensory structures and signal transduction pathways.

Acquiring information is only one aspect, however, as processes are designed to utilize different types of information in different ways. Some signaling molecules convey more information than others, while some simply contain different types of information. Signal transduction pathways have evolved to respond to these different qualities, and although they operate in real-time, some pathways can respond to temporal information about forthcoming events. In reactive processes, little uncertainty exists about the utility of the response because there is a direct temporal relationship between the initiating signal indicating the need for the response and the response itself. In anticipatory processes, information about future scenarios can be transmitted through temporal correlations between a current signal and future events. When environmental factors change in predictable ways, the detection of a change in one variable can reliably predict a future change in another, reducing uncertainty, but these correlations are still probabilistic even if highly reliable. As such, the anticipatory processes contain an inherently higher level of uncertainty than reactive processes. For more information about information theory and cell signaling see Brennan et al. (2012).

### *Ecological Considerations*

In an unchanging environment it is difficult to imagine a use for anticipatory processes. All physiological functions would be carried out by reactively, as organisms would have nothing to anticipate. On Earth, however, environmental change is common across spatio-temporal scales and because of this, living things must be able to sense and react to these changes. Some organisms do this through reactive, sometimes called reflexive, processes that sense a change, then elicit a response. Organisms in erratic environments will sometimes switch randomly between responses when environmental fluctuations are quick and difficult to detect (Mitchell et al., 2009; Kussell and Leibler, 2005; Ben-Jacob and Schultz, 2010). When environmental patterns are reliable, however, the ability to predict environmental changes is more tenable and can be highly beneficial. The ability to anticipate change is adaptive under adverse environmental conditions, as advanced preparation can increase an organism's chances of survival, but also under favorable environmental conditions, as it can improve an organism's competitive edge. In general, anticipatory regulation is most favorable when response times are long and environmental uncertainty is high, as the ability to mitigate changes effectively can be limited (Kussell and Leibler, 2005; Freddolino and Tavazoie, 2012).

Ecosystems have distinct biogeochemical cycles that often produce reliable environmental patterns. Because of this, many qualitative and quantitative aspects of environmental factors are strongly correlated with each other. When these correlations have a temporal element, they contain information about the time-course of environmental events, and biological processes can be designed to take advantage of this information, i.e., anticipatory processes. Figure 1 provides a graphical representation of how these different signals can affect organismal responses. The time course of a theoretical environmental factor is shown in panel (a), and panel (b) shows the time course of Signal 1, a signal with the same temporal pattern as



the environmental factor, and Signal 2, which is a signal that is correlated with, but precedes, the environmental factor. Figure 1c shows that because the reactive response is regulated by Signal 1, there is a lag in the response-time. This delay is associated with the amount of time it takes to produce the response, such as time related to gene transcription and protein production/transport. The anticipatory response, on the other hand, has no lag in response-time because it is regulated by Signal 2, which precedes the change in environment.

Anticipatory processes are most likely to evolve when environmental associations are closely tied to the fitness of the organism. So, when they represent a significant constraint on survival or performance. Additionally, the costs of anticipatory, versus reactive, processes must be less than the fitness benefits obtained through them. In some cases, anticipation may justify the evolution of costly sensory structures, such as brains, but benefits can also be gleaned from simple inexpensive changes in the design of regulatory pathways. As the costs of anticipatory regulation increases, these processes are only favorable under high levels of environmental uncertainty (Kussell and Leibler, 2005; Freddolino and Tavazoie, 2012).

A good example of these ecological considerations can be found in Tagkopoulos et al.'s (2008) work with *Escherichia coli*. The life cycle of *E. coli* involves the movement of the bacteria from soil, into the mouth of a host, and eventually into the intestines, during which there are predictable fluctuations in environmental temperature and oxygen at each stage. Initial ingestion is accompanied by an increase in temperature, and as the bacteria travels to the intestines, it experiences a shift from an oxygen-rich to an oxygen-poor environment. Thus, there is a strong temporal association between the increase in temperature and an impending drop in oxygen availability. These factors have important ecological implications for *E. coli*, as switching from aerobic to anaerobic respiration is an energetic and time-consuming process, also one that if not timed properly, will result in death. This example demonstrates how a temporal correlation between two fitness-relevant factors could favor the evolution of anticipation. Tagkopoulos et al. (2008) did indeed show that *E. coli* transcriptionally regulate metabolic genes in an anticipatory fashion when exposed to increases in temperature. In fact, they showed experimentally that after temperature is increased, genes related to aerobic respiration were significantly repressed while those related to anaerobic respiration were significantly up-regulated, even when the current oxygen levels were still high.

### *Physiological Considerations*

While the ecological considerations outline the utility for anticipation in evolutionary terms, anticipatory processes are ultimately carried out by physiological responses. One of the defining features of life is its opposition to entropy. In a dynamic environment, organisms must maintain order within themselves, a feature epitomized by the concept of homeostasis, which refers to an organism's internal steady state. It is also often defined as the process by which this internal state and proper physiological function is maintained after perturbations occur. Inherent to the idea of homeostasis is physiological constancy and the existence of static biological optima, or setpoints (Cannon, 1929; Moore-Ede, 1959). Homeostasis, as a process, operates by detecting deviations away from these setpoints, termed error, and eliciting responses to return setpoints back to their original state. Homeostasis can be described as a reactive process that is only initiated after error occurs. However, the observation that many traits in humans and other animals, including blood pressure, insulin secretion, body temperature, etc., vary consistently throughout the day, challenges the notion of static setpoints. Additionally, the existence of physiological trade-offs contradicts the idea that setpoints must be unchanging for homeostasis to be reached. Over time, new terminology has been created to account for these dynamic aspects

of physiological regulation, including homeorhesis (Waddington, 1958, 1968; Bauman and Currie, 1980; Baumgard et al., 2017), heterostasis (Selye, 1975), homeokinetics (Soodak and Iberall, 1978), rheostasis (Mrosovsky, 1990), but few terms have caught on in popular literature.

The reactive nature of homeostasis was questioned by Moore-Ede (1986), after noticing that the diurnal cycles of many physiological processes, including body temperature, plasma cortisol levels, growth hormone levels, and urinary potassium excretion, began before changes in light cues occurred and/or ahead of the sleep-wake cycles that were assumed to be regulating them (Moore-Ede, 1982; Fuller et al., 1985). This led to the conclusion that these traits were regulated by a form of predictive homeostasis, a process that was more efficient because it minimized error. Sterling and Eyer (1988) built off of this idea to produce the concept of allostasis, which places fitness, rather than constancy, as the primary objective of physiological regulation (Sterling, 2004). In contrast to homeostasis, allostasis affirms that fitness is primarily maintained through physiological flexibility because optima are constantly changing across different environments. As a result, the best way to reduce error is to pre-set optima to predicted levels based on the expected physiological response drawn from past experience (Sterling and Eyer, 1988; Sterling, 2004; McEwen, 1998; Schulkin, 2009). Because environmental factors are often temporally correlated, using environmental information to adjust setpoints produces more efficient responses than keeping setpoints at some average level, making an allostatic model more evolutionarily advantageous, and thus more plausible. Although not a commonly discussed concept, the allostatic perspective has in fact improved our understanding of the regulation of several traits, in addition to providing a strong foundation for anticipatory regulation (Tagkopoulos et al., 2008; Mitchell et al., 2009; Freddolino and Tavazoie, 2012; Perkins et al., 2009).

Regulatory mechanisms are the most physiologically-relevant aspects of anticipatory processes and are delineated by stimulus-response (S-R) relationships. It is important to note that the accurate detection of stimulus-response relationships is imperative to discerning between reactive and anticipatory processes, but these relationships are not always easy to detect, especially in complex regulatory networks. Mitchell et al. (2009) provides a nice framework for conceptualizing the regulatory differences between reactive and anticipatory processes by distinguishing between four types of regulatory strategies: direct regulation, stochastic switching, symmetrical anticipatory regulation, and asymmetrical anticipatory regulation. Direct regulation occurs when one stimulus activates one response ( $S_1 \rightarrow R_1$  and  $S_2 \rightarrow R_2$ ) and is most similar to generalized reactive processes. In stochastic switching, one stimulus activates one of two responses randomly ( $S_1 \rightarrow R_1$  or  $R_2$ ). In symmetrical anticipatory regulation (SAR), two stimuli elicit their own responses and the response of each other ( $S_1 \rightarrow R_1$  or  $R_2$  and  $S_2 \rightarrow R_2$  or  $R_1$ ). In asymmetrical anticipatory regulation (AAR), one stimulus activates its own response and the response of another stimulus, while the other stimulus only activates its own response ( $S_1 \rightarrow R_1$  or  $R_2$  and  $S_2 \rightarrow R_s$ ).

A good example of these strategies can be seen in the regulation of sugar metabolism in *E. coli* (Mitchell et al., 2009; Freddolino and Tavazoie, 2012). *E. coli* can utilize a range of different sugars for energy acquisition, and throughout their life cycle they encounter different sugars in a particular order. As shown in Fig. 2a, *E. coli* bacteria encounter lactose at the proximal end of the small intestine, while maltose is not encountered until reaching the distal end of the small intestine (Mitchell et al., 2009; Savageau, 1998). Because of this, exposure to lactose always precedes exposure to maltose. Figure 2b shows how the *lac* operon, which controls the expression of lactase enzymes, operates in a reactive manner. The expression of lactase genes

only occurs in the presence of lactose, which serves as a direct stimulus. The *mal* operon, which controls the production of maltase enzymes, operates in an anticipatory manner, as the expression of maltase is stimulated by the presence of maltose (direct stimulus) but also lactose, which serves as an indirect stimulus. Because of this, maltase expression begins when bacteria encounter lactose in the proximal small intestine but before they are ever exposed to maltose. This way, maltase is already present when maltose becomes available, reducing the response-time associated with maltase production and increasing the efficiency of maltose catabolism. Although Mitchell et al. (2009) did not directly measure the response-time of maltase production under different sugar scenarios, they did show that pre-exposure to lactose significantly increased the fitness of *E. coli* subsequently reared on maltose. Some other examples of putative anticipatory regulation include the cross-protection of yeast to oxidative stress when pre-exposed to heat shock and/or ethanol shock (Mitchell et al., 2009), the high-risk foraging behaviors of the slime mold *Physarum polycephalum* on low quality food sources (Latty and Beekman, 2010), and the up-regulation of motility genes and metabolic genes for preferred carbon sources in *E. coli* reared on lower quality carbon sources (Liu et al., 2005; Zhao et al., 2007).

These generalized regulatory strategies show how qualitative differences in S-R relationships can produce reactive and anticipatory processes. Anticipatory processes, however, can also stem from alterations to the quantitative features of S-R relationships, such as stimulus thresholds and response dynamics. For instance, hormetic responses are characterized by a disproportionate response to low levels of stress, where low doses of stress cause an overcompensated response, hypothesized to better prepare the organisms for ensuing higher doses (see section on Hormesis). Priming, also called preconditioning, is another example of a dose-dependent response. Primed responses occur when organisms are exposed to low-moderate amounts of stress that initiate a partial response that is only completed upon exposure to higher levels of stress (see section on Priming). The primed state occurs in anticipation of higher levels of future stress, much in the same way that the overcompensated response in hormesis does. In both examples, a stimulus can produce a reactive or anticipatory response depending on the concentration of the stimulus.

### *Biochemical Considerations*

All biological processes are fundamentally mediated by biochemical reactions. As such, defining anticipatory processes in biochemical terms is essential for demonstrating their fundamental role in biology. This is unfortunately quite difficult to do, however, because our understanding of anticipatory mechanisms is still quite limited. Despite this, considerations at broader biological scales can be used to shape our understanding of biochemical criteria, even if only hypothetically. For instance, the causal aspects of anticipation highlight the importance of timing, while physiological considerations provide us with a S-R structure to build off of. These components and the constraints they entail allow for the development of hypotheses about the more ambiguous biochemical characteristics.

Every biological process has a beginning, a middle, and an end. The timing of the reactions involved at each of these stages has important impacts on the efficiency of response. Many physiological responses are only induced when needed, thus, they are initiated once homeostatic error occurs. Once error is detected, there is a preparatory phase where the required components (enzymes, substrates, and/or co-factors) must be produced, transported, and/or concentrated so that downstream reactions can take place. In fact, four important events can be demarcated in the time course of biological responses: the manifestation of error, the initiation of the preparatory phase, the initiation of downstream reactions, and the commencement of the



response. The organization of these events affect the overall duration of the response, which is the time between the initiation of the preparatory phase and commencement, as well as the response-time, which is the period of time between the manifestation of error and the initiation of downstream reactions; or when error mitigation actually begins. Figure 3 shows the time course for each of these events in a reactive response and a generalized anticipatory response. It's important to note that the duration of the preparatory phase can vary depending on the response, but regardless of whether it is long or short, it results in a delay in the downstream reactions and an increase in response-time. Anticipatory processes, however, are coordinated such that all or part of the preparatory phase occurs before the manifestation of error, thereby reducing response-times. This is evident in Fig. 3, which shows how the early initiation of the preparatory phase reduces the response-time for anticipatory processes relative to reactive processes. Using these events, we can then describe anticipatory processes in biochemical terms, as those processes that reduce response times by initiating the preparatory phase before the manifestation of error. These characteristics can be achieved in different ways; either through qualitative or quantitative alterations to the design of the process.

For instance, processes controlled by AAR represent good examples of qualitative alterations. In AAR there are two types of S-R relationships; two direct relationships ( $S_1 \rightarrow R_1$  and  $S_2 \rightarrow R_2$ ) and one indirect relationship ( $S_1 \rightarrow R_2$ ). The differences between direct and indirect stimuli can help produce useful hypotheses regarding the biochemical differences between reactive and anticipatory processes. Direct S-R relationship can be viewed as primary, while indirect stimuli are ancillary. A direct stimulus indicates that a physiological response is needed and elicits a response in real-time. To do this, a temporal relationship must exist between the presence of the stimulus and the manifestation of error, which suggests that direct stimuli are likely to be compounds or by-products of the process creating error, such as toxins or antigens. An indirect stimulus, on the other hand, need not have an intimate biochemical relationship with the process creating error or to the direct stimulus. It must simply be temporally correlated with the creation of error and/or the direct stimulus. This suggests that there are far fewer constraints on the chemical properties of indirect stimuli and their relationship to the downstream reactions they elicit relative to direct stimuli.

While some anticipatory processes utilize different stimuli to change the time course of responses, other processes simply alter the modulation of the response itself. Priming, which is discussed in more detail in the next section, is a good example of this. Sometimes called sensitization or preconditioning, priming occurs upon re-exposure to a prior stress. From a biochemical perspective, priming shortens response times by initiating and carrying out part of the preparatory phase at low levels of stress so that the response is primed and downstream reactions can occur more quickly upon re-exposure. While sensitization mediated by neurological responses is well understood as a form of neural plasticity, the mechanisms underlying other types of priming, particularly in non-cognitive species are more ambiguous. In plants, however, priming appears to be mediated by the accumulation of dormant signaling proteins, the storage of metabolite conjugates, and/or changes to epigenetic marks, such as gene methylation and histone modifications. For instance, in *Arabidopsis*, exposure to both salicylic acid analogues and bacterial infection leads to the accumulation of inactive mitogen-activated protein kinases, specifically MPK3 and MPK6 (Beckers and Jaskiewicz, 2009; Conrath, 2011), suggesting that they allow the plant to amplify stress signals faster upon subsequent exposure. The accumulation and storage of other compounds, such as reactive oxygen species and metabolite conjugates in vacuoles have also been documented in primed plants (Pastor et al.,

2013). Gene methylation has been observed in the priming of barley to powdery mildew exposure (Conrath, 2011; Kraska and Schönbeck, 1993), and histone modifications have been tied to increased expression of defense-related genes in primed responses due to the faster recruitment of transcription factors, co-activators, effector protein, and general transcriptional machinery (Alvarez-Venegas et al., 2007; March-Díaz et al., 2008; van den Burg and Takken, 2009; Jaskiewicz et al., 2011). In either case, the same stimulus triggers the response but response time is increased achieved by changes in the modulation of the response after it has been initiated.

### *Towards an Integrative Definition*

Figure 4 shows the criteria defining biological anticipation at different scales. These considerations can also be used to identify potential anticipatory processes based on different ecological, physiological, and biochemical criteria. Because reactive processes utilize simpler designs, and likely predate anticipatory processes, we can use them as a comparator to further understand the characteristics that make a process anticipatory. Moreover, examining processes that exist in both reactive and anticipatory forms can be useful.

If we consider the examples discussed above, we can see how considerations at different scales connect to provide an evolutionary justification for anticipation that is consistent with physiological and biochemical mechanisms. Using AAR as an example, we can imagine a scenario where a physiological response, denoted by a specific S-R relationship, evolves to be regulated by a new stimulus that is correlated with but predates a fitness-relevant environmental variable and that this alteration causes the preparatory phase of the response to be initiated before error occurs. This generalized scenario combines the criteria for anticipation at all level of biological organization into a cogent explanation. The next section takes things further and discusses real biological phenomena that meet some or all of these criteria.

## **Examples of Biological Anticipation**

Now that the relevant aspects of anticipation have been discussed at the causal, ecological, physiological, and biochemical levels, we can explore some examples. The following section outlines some key categories of anticipatory processes and provides specific examples, with a bias towards non-cognitive models. It should be noted that this is not an exhaustive list, and the categories may not necessarily be mutually exclusive, i.e., some examples may fall into more than one category. Despite this, the goal of this section is to show how different biological phenomena, studied using different model systems often in disparate fields, can be united under an anticipatory framework. Many of the examples discussed in this section, while well-established, have proved difficult to explain by reactive mechanisms. Because of this, many have been viewed, for at least a period of time, as biological outliers. They are increasingly explicable, however, by anticipatory mechanisms, highlighting the utility of creating firm distinctions between reactive and anticipatory processes.

### *Circadian Rhythms*

Circadian and circannual rhythms are arguably one of the oldest mechanisms of anticipation. Found in both prokaryotes and eukaryotes, these rhythms are controlled by an internal biological clock that is calibrated to the diurnal (day-night) and annual cycles of the Earth. Regulating physiological processes by an internal clock is more efficient than responding to individual environmental cues because so many environmental factors correlate with diurnal

and annual cycles. This not only streamlines regulatory processes and reduces the need for larger more costly networks, but, as argued by Moore-Ede (1986), it also allows for the anticipation of physiological needs. Possessing an internal clock that is calibrated to stable environmental periodicities allows organisms to schedule metabolic functions in the most efficient way, as shown in Fig. 5a with the example of photosynthetic compounds in plant increasing in anticipation of daylight. In humans, plasma cortisol and core body temperature up-cycle in the morning before any light stimulus is present (Moore-Ede, 1986). The diurnal regulation of insulin (Hara and Saito, 1980) digestive enzymes (Stevenson et al., 1975; Saito et al., 1976a, 1976b), and glucose transport (Stevenson and Fierstein, 1976) in rats have also been shown to be anticipatory, as well as circadian-controlled food seeking behavior in rats and squirrel monkeys (Moore-Ede, 1986; Boulos et al., 1980; Mistlberger, 1994; Stephan, 2001; Davidson et al., 2003). Circadian-based anticipatory mechanisms temporally coordinate metabolic activities to reduce the lag times associated with reactive responses. This strategy ensures that enzymes, proteins, nutrients, etc. are available at the instant they are needed, maximizing physiological efficiency.

Circadian rhythms are well documented in mammals, insects, fungi, and plants (Bhadra et al., 2017; Millar and Kay, 1996; Harmer et al., 2001), as well as prokaryotes (Johnson et al., 1996; Whitehead et al., 2009; Cohen and Golden, 2015; Nohales and Kay, 2016) and unicellular eukaryotes (Johnson and Kondo, 2001; Mittag and Wagner, 2003; O'Neill et al., 2011), suggesting that they evolved early on and also multiple times in different lineages (Johnson and Kondo, 2001). Traits exhibiting circadian rhythmicity vary widely across taxa and of course not all circadian-controlled traits demonstrate an anticipatory function. Despite this, the basic components of circadian processes are surprisingly similar, consisting of an endogenous central oscillator, input pathways that entrain the oscillator, and output pathways that manifest periodicity in specific traits. Some organisms, such as mammals, possess multiple clocks, including the master pacemaker in the suprachiasmatic nucleus (SCN) and peripheral clocks in other tissues. Despite differences in specific components, all circadian systems rely on delayed negative feedback loops at the transcriptional level (Harmer et al., 2001; Dunlap, 1999), as well as post-transcriptional regulation, to produce the 24-h oscillatory phase. While many clock components exhibit homology across taxa, primary differences in circadian mechanisms relate to variations in specific regulatory factors and complexity, which is more thoroughly reviewed by Harmer et al. (Harmer et al., 2001) and Bhadra et al. (2017). While the details of entrainment are well-understood for many taxa, our understanding of the circadian regulation of specific traits and its evolutionary history is limited. Elucidating the mechanistic relationship between the central oscillator and the regulation of specific physiological pathways in non-cognitive organisms will be essential for further understanding the evolution of circadian systems and this type of biological anticipation in general.

#### *Asymmetrical Anticipatory Regulation*

As previously discussed, when environmental factors show reliable temporal patterns the presence of one factor can be used to anticipate the future presence of another, much like associative learning. Regulatory pathways across diverse taxa have evolved to take advantage of these correlations through anticipatory processes (Tagkopoulos et al., 2008; Mitchell et al., 2009; Freddolino and Tavazoie, 2012). The regulation of these processes can be altered through changes in signaling, transcription, translation, and post-translational pathways. Asymmetrical anticipatory regulation, as defined by Mitchell et al. (2009), occurs when a response is triggered by an environmental signal that is correlated to and precedes the original stimulus. In this way,

the response can be initiated earlier, better preparing the organism for future conditions, as demonstrated in Fig. 5b.

Identifying cases of AAR can be incredibly difficult, as it requires having a detailed understanding of an organism's ecology and the regulatory mechanisms for specific processes. The outcomes of regulatory networks are easy to measure but the mechanisms that produces them are much more difficult to document. Uncharacterized processes are often assumed to possess reactive S-R relationships by default, thereby reducing the discovery of AAR. However, unusual correlations in transcriptional patterns or unintuitive phenotypes can be good indicators that AAR is at play, particularly if the relationship can be explained ecologically. For example, Tagkopoulos et al. (2008) found that *E. coli* genes associated with anaerobic respiration were up-regulated when populations were subjected to a spike in temperature. This relationship makes little physiological sense, particularly considering that anaerobic genes increased while oxygen levels were still high; but, when you consider that the ecology of *E. coli* necessitates a move from the mouth of the host, a high temperature environment, to the anoxic intestines of the host, a functional explanation emerges. The spike in environmental temperature reliably indicates an ensuing anoxic environment, so using a thermal signal to initiate the production of anaerobic cellular machinery better prepares the bacteria for their future environment. Tagkopoulos et al. (2008) also showed that the relationship between temperature and anoxia could be decoupled by rearing bacteria under the opposite conditions (where temperature and oxygen decrease) for less than 100 generations, further substantiating that the transcriptional relationship was an evolved response to an environmental pattern rather than biochemical constraints. Mitchell et al. (2009) reported similar cases of AAR in the sugar metabolism of *E. coli* (Fig. 2) and in the stress-response of *S. cerevisiae* during the wine-making process. Other putative examples of AAR have also been reported in *Vibrio cholerae* (Dunlap, 1999), *Candida albicans* (Rodaki et al., 2009; Mitchell and Pilpel, 2011), *Mycobacterium tuberculosis* (Mitchell and Pilpel, 2011), and other stress responses in *S. cerevisiae* (Eoh et al., 2017; Berry and Gasch, 2008).

Although these more recent microbial studies have provided extremely new data on AAR, evidence of AAR can be found in the scientific literature as early as the turn of the century. For instance, the existence of cephalic responses, which were discussed earlier, have been known since Pavlov's seminal work on the alimentary canal (it is interesting to note that Pavlov called these cephalic responses "psychic secretions") (Smith, 2000; Power and Schulkin, 2008). Another example of AAR from a non-cognitive organism, is plant eavesdropping. Plants have the ability to pick up on above- and below-ground cues from neighboring plants about potential threats, such as herbivory or drought, and use them to induce defensive responses before exposure to stress occurs (Karban et al., 2003; Engelberth et al., 2004; Kessler et al., 2006; Falik et al., 2011). The majority of work on plant eavesdropping has been done on herbivore-induced plant volatiles, but signals emitted by roots and microbes in the soil have also been studied (Falik et al., 2011, Morrell and Kessler, 2014; Blande, 2017; Rebolleda-Gómez and Wood, 2019). When plants are damaged, the composition of their volatiles change and can serve as reliable signals of impending pest presence. These cues can either elicit direct induction of defenses or prime defense pathways (Morrell and Kessler, 2014). Eavesdropping allows plants to respond to the volatile cues that precede stress rather than the physical (plant damage) and/or chemical (insect saliva) cues that are only present after damage has occurred.

### Priming

Priming, which was also discussed earlier, refers to the enhanced response to a stressful

stimulus that has been previously encountered. Initial exposure to a stressful agent can elicit a direct response but can also prime future responses to the same or different stimuli. Priming causes a response to be partially initiated, paused, and only completely carried out after a secondary exposure. In this way, stress responses can be used therapeutically and carried out faster and with greater efficacy. Figure 5c shows a hypothetical example of priming for resistance to herbivory in plants, where a plant that has experience herbivory in the past is better able to respond to subsequent pest pressure than plants that have never been exposed. Priming can also be seen as a fail-safe to ensure that costly processes are employed economically and only carried out when absolutely necessary. Primed traits can persist over the short-term or long-term, mediated by different mechanisms. Given that priming is implemented as a response to future scenarios and it increases biological efficiency by reducing response-time, it qualifies as an anticipatory process.

The most cohesive body of work on priming in a non-cognitive organism comes from the study of induced plant defenses. In plants, exposure to pathogenic and apathogenic bacteria (Jung et al., 2009; Balmer et al., 2015), fungi (Molitor et al., 2011; Gamir et al., 2012; Martínez-Medina et al., 2013), oomycetes (Massoud et al., 2012; Lim et al., 2013), nematodes (Hao et al., 2012; Vos et al., 2013), arthropod damage (Vos et al., 2013; Smith et al., 2012) egg oviposition (Kim and Felton, 2012), abiotic stressors (Tutegja and Sopory, 2008; Suzuki et al., 2012; Virilouvet et al., 2018), and synthetic effectors (Bektas and Eulgem, 2015; Conrath et al., 2015; Conrath, 2011) can prime plant defense pathways, conferring increased resistance to a wide range of stressors compared to unprimed plants. While constitutive and direct induction of plant defenses can be costly, primed plants exhibit full protection with fewer fitness costs due to flexibility that priming offers (van Hulten et al., 2006; Wang et al., 2015).

The pathways and processes responsible for priming in plants are diverse, as are the post-challenge responses of primed states. They depend on the species of plant, the type of stress, and other environmental factors that may constrain physiological responses. Short-term mechanisms of priming include the accumulation of signaling proteins or metabolite conjugates, while long-term priming results from changes to epigenetic markers. Plant defense metabolites, such as reactive oxygen species (ROS) and pathogenesis-related proteins (PRs), inactive conjugated plant signaling hormones (ethylene (Et), jasmonic acid (JA), abscisic acid (ABA), salicylic acid (SA)), microbe-associated molecular pattern (MAMPs) receptor abundance, and mRNA of other signaling components (mitogen-activated protein kinases (MAPKs, MEKs, and MEKKs)) have all been shown to accumulate in the tissue of primed plants, allowing for faster activation and reduced response-times upon subsequent exposure (Beckers et al., 2009; Conrath, 2011; Pastor et al., 2013). Additionally, changes in chromatin structure, via histone modifications and DNA methylation, have been correlated with the increased expression of defense genes due to enhanced accessibility and recruitment of transcription co-factors and general transcription machinery (RNAPII, TFIID) (Alvarez-Venegas et al., 2007; March-Díaz et al., 2008; van den Burg et al., 2009; Jaskiewicz et al., 2011; Conrath, 2011). Specifically, the methylation and acetylation of lysine residues on histones H3 and H4 (H3K4me3, H3K4me2, H3K9ac, H4K5ac, H4K8ac, H4K12ac) and the replacement of H2A with H2A.Z have been tied to increased expression of defense genes in *Arabidopsis* (Alvarez-Venegas et al., 2007; March-Díaz et al., 2008; van den Burg et al., 2009; Jaskiewicz et al., 2011). Importantly, these epigenetic marks can allow a primed state to be transferred to the next generation (see the Transgenerational Effects section).

Priming also occurs in animals, though it is more commonly referred to as



preconditioning. The most well-studied examples of priming relate to ischemic cardiac preconditioning, hypoxia, immunological challenges, and thermal stress. Sub-lethal ischemia, oxidative damage, volatile anesthetics, and pharmacological interventions have been shown to reduce subsequent myocardial infarction size and ischemia in various organs (Murry et al., 1986; Belhomme et al., 1999; Zaugg et al., 2002). Hypoxic and thermal preconditioning have also been widely shown to attenuate ischemic injury to different organ systems and other general stressors (Ahmad et al., 2003; Joyeux-Faure et al., 2005; Bernhardt et al., 2006; Lu et al., 2012). Additionally, exposure to bacteria, viruses, parasites, and tumor cells can prime monocyte and macrophage responses via interferon- $\gamma$  signaling, resulting in faster cytokine biosynthesis upon secondary detection of lipopolysaccharides (Hayes et al., 1995; Raetz et al., 1991; Ma et al., 2015). The specific mechanisms related to these types of priming vary, but like in plants, the primed state is mediated by enhanced signal transduction, as well as epigenetic effects, like DNA methylation, histone modifications, and microRNA activity (Meller et al., 2015; Stankewicz et al., 2013; Horowitz, 2016).

Priming is also common in more primitive taxa, such as bacteria and biofilms, where it is often described as tolerance. Priming in response to pH (Foster and Hall, 1991; Shah, 2000), temperature (Boutibonnes et al., 1993; Walker et al., 1999; Desmond et al., 2004), salinity (Vargas et al., 2008), oxygen stress (Ahn et al., 2001), hydrostatic pressure (Ananta and Knorr, 2003), and several other sub-lethal stressors have been shown to increase tolerance to a range of different stressors (Hartke et al., 1994; De Angelis and Gobetti, 2004; Duwat et al., 2000).

### *Hormesis*

Hormesis is an adaptive stress-response identifiable by a biphasic nonlinear dose-response curve that exhibits a stimulatory (positive) effect at low stress levels and an inhibitory (negative) effect at higher stress levels (Calabrese and Baldwin, 1997). It is used to describe a biological response to stress, but can also occur in response to other stimuli that are not inherently harmful, such as nutritional signals. A hormetic dose-response, shown in Fig. 5d, is represented by a J- or U-shaped curve depending on whether the y-axis represents a parameter of normal biological function or dysfunction (Calabrese and Baldwin, 2001). At its core, hormesis is an example of adaptive over-response, whereby acute or chronic exposure to low levels of stress can result in a positive increase, typically 30-60%, in various fitness-relevant endpoints. Hormesis was coined in 1943 by Southam and Ehrlich (1943) to describe the enhanced metabolism of fungi exposed to Red Cedar extract, however, the stimulatory effects of low doses of toxins have been acknowledged since the 1880s. Although widely accepted in plant and microbial systems, it took longer for hormetic effects to be recognized in the field of mammalian toxicology. This was largely due to a perceived association with homeopathy but also to traditional medicine's adoption of threshold dose-response models, which preclude the identification of responses in the low-dose zone (Calabrese, 2014). Today, hormesis has been documented in a diversity of taxa, from bacteria to humans, and documented for thousands of different compounds and physiological endpoints (Calabrese et al., 2008). As such, it is now considered to be a fundamental biological response. Although hormesis hasn't been explicitly associated with anticipation or described as an anticipatory process (to my knowledge), it deserves mention here as it meets many of the criteria.

Calabrese et al. (2007) describes three different types of hormesis: hormesis, which refers to the basic phenomena described above; conditioning hormesis, where prior exposure to a low-level stress reduces the effects of a subsequent higher exposure; and postexposure conditioning hormesis, where the negative effects of exposure to a high-level stressor is reduced by the

subsequent exposure to a low-level stressor. As conditioning hormesis and postexposure conditioning hormesis relate more strongly to priming, this section will focus on the basic form of hormesis, i.e., the low-dose stress response in a naïve organism or cell. Hormesis is characterized as a type of general overcompensation to a homeostatic disturbance. The stimulatory effects observed at low stress exposure are presumed to stem from the unintended benefits of over-activated defense pathways in the absence of higher stress levels. Hormesis, for instance, can result in increased survival due to the up-regulation of general defense pathways that inadvertently protect the organism from non-target sources of morbidity/mortality, as seen in dietary restriction. Even though, the stimulatory effects of hormesis are quite moderate, exhibiting a 30-60% change relative to control groups, it is difficult to derive an evolutionarily-sound explanation for this sort of response, given that an over-reaction is an inherently inefficient response. One would expect natural selection to optimize stress-responses so that they are adequate for mitigation but not excessive. Then again, one wouldn't expect selection against any process that increases fitness. However, if we view hormesis as an anticipatory process, things begin to make more sense.

Just as biological processes can evolve to respond to temporal patterns in qualitative environmental signals, as in AAR, they can also evolve to respond to temporal patterns in the quantitative aspects of environmental signals. Probabilistic relationships can exist between different events and also between different degrees of the same event. For instance, exposure to a specific amount of toxin could either indicate that a higher or lower dose is forthcoming. The relative probability of a higher or lower future dose depends on the shape of dose-time curve for the stressor in the environment and the position of the present dose along this trajectory. Like most environmental factors, the quantitative aspect of different stressors can vary through time in predictable ways, and this information can be integrated into stress-response pathways and used to modulate responses in an anticipatory manner. In this way, the over-response observed at low doses of stress can be viewed as an attempt to mitigate the damage caused by higher future doses; but, interestingly, the benefits of hormesis do not derive from achieving this goal of enhanced preparedness. They instead originate from the preparation itself. Hormesis describes a situation where the benefits of an anticipatory response are evident in both the presence and absence of a future stressor. This makes hormesis a very unique type of anticipatory response, as its utility appears to be dependent on the inaccurate prediction of a future state; in other words, on mounting a stress response for a stressor that never comes.

While this may sound quite controversial, one could easily imagine a scenario where a stress-response could prove beneficial under control conditions, particularly if it was only induced under very specific circumstances. Additionally, evolutionary theory doesn't predict strong selection against any phenotype that proves to be beneficial, even if it is the result of an erroneous prediction. A possible explanation for the maintenance of hormesis may lie in the generality of the response. While many stress responses are very specific, hormesis appears to elicit a systemic generalized response that up-regulates a suite of defensive pathways regardless of the causal stressor. The up-regulation of specific stress-response pathways during control conditions would be of little benefit in the absence of that specific stress, but a more generalized response, like that observed in hormesis, could be beneficial, particularly since organisms must routinely contend with multiple stressors simultaneously. For instance, the activation of detoxification pathways would be a waste of resources in the absence of toxins because detoxification enzymes serve no other purpose than to aid in the excretion of toxic compounds, but the up-regulation of processes protecting against oxidative stress, which is a natural

byproduct of energy metabolism, would likely have positive effects under normal conditions.

In general, the nature of hormetic responses is not well understood. It is hypothesized by some to be an example of the universal cellular response (UCR), sometimes called protoreaction, a highly conserved phenomenon characterized by dose-dependent physio-chemical changes in the intracellular environment (Nasonov, 1962; Matveev, 2005; Matveev, 2010) and mediated by changes in cytoplasmic factors upon exposure to a wide variety of different stressors. At low levels of stress, an initial decrease in cytoplasmic turbidity and viscosity is observed, as well as membrane hyperpolarization. At higher stress levels, a simultaneous increase in intracellular turbidity and viscosity and membrane depolarization are seen (Matveev, 2005; Matveev, 2010; Suzdalskaya, 1977). The UCR has been shown to produce increased resistance to stress at low initial stress levels and reductions in this resistance as stress level increases, similar to the effects of hormesis. These characteristics are hypothesized to stem from changes in protein folding, cellular hydrophobicity, and intracellular  $K^+$  gradients but the specific mechanisms are not known. Unfortunately, investigations into the UCR have largely ceased in the wake of new molecular advancements that prioritize fine-scale quantitative mechanisms over more broad qualitative phenomena (Agutter, 2007).

Whatever the exact mechanism, it is important to note, as Forbes (2000) points out, that hormesis is not a phenomenon that acts directly on fitness, but instead is a homeostatic/allostatic mechanism that aims to maintain fitness across stressful conditions. As a result, the stimulatory effects of hormesis are likely subject to physiological trade-offs with other traits. An excellent example of this trade-off can be seen in dietary restriction (DR), a phenomenon where an increase in maximal and median lifespan is observed for organisms fed slightly deficient but not malnourishing diets (McCay et al., 1935; Piper et al., 2011). Dietary restriction has been documented across diverse taxa, from yeast to mammals (Mair and Dillin, 2008; Fontana et al., 2010), and represents a hormetic response where changes in diet, either a reduction in calories or specific macronutrients depending on the model organism, cause a systemic up-regulation of cellular defense pathways that reduce a variety of age-related pathologies and lead to increased longevity (Masoro, 2007; Mair et al., 2005; Gems and Partridge, 2008; Simpson and Raubenheimer, 2009). It is a completely reversible response and, in most organisms, is associated with a concomitant reduction in reproductive output, indicating a trade-off between survival and reproduction (Lee et al., 2008; Moatt et al., 2016). The overriding evolutionary hypothesis for DR is that nutritional signals can act as indicators of environmental suitability, and when these signals change in a specific way, they indicate the presence of stressful conditions and elicit a biological response that prioritizes defense rather than reproduction. In other words, DR is a biological program to “ride out the storm”. Different types of DR exist, with effects varying depending on whether the signal comes from a change in diet composition (protein or calories) or the timing of feeding (intermittent feeding, fasting, etc.) (Fontana et al., 2010). Several different nutrient-sensing pathways have been implicated in this response, including the insulin-like signaling pathway, the target or rapamycin pathway, and the AMP-kinase signaling pathways, with many downstream effects mediated by FOXO transcription factors (Fontana et al., 2010; Solon-Biet et al., 2015). Despite the many advancements in understanding DR, the mechanisms coordinating various defense pathways remain unknown.

### *Transgenerational Effects*

Lastly, I want to discuss how transgenerational effects can serve as a unique type of anticipatory processes; one whose purpose is to benefit the next generation. Up to this point, we have only discussed anticipation as it relates to individual organisms, particularly processes that

maintain or increase an individual's survival and/or performance across variable environments. Anticipation, however, can also be used to better prepare offspring for future events, via maternal/paternal effects and epigenetic marks (Angers et al., 2010; Bollati and Baccarelli, 2020; Leung et al., 2020). Transgenerational effects broadly refer to offspring phenotypes that are not dependent on changes in DNA sequence, and can occur when the conditions experienced by the parent either directly affect the cells that will become future offspring (maternal/paternal effects), or cause changes to epigenetic markers that can persist for multiple generations (epigenetic effects). Both processes are capable of impacting gene expression and protein production in offspring in adaptive ways. Although, this section focuses on generational impacts at the organismal level, the same mechanisms are relevant at the cellular level as well.

Parental effects refer to heritable phenotypes that are set in offspring before birth, either through the persistence of expressional states initiated in pre-zygotic cells (primordial germs cells or gametes), or through the passage of cytoplasmic signaling molecules, transcription factors, or RNAs from either parent during fertilization (Badyaev and Uller, 2009). Messenger RNAs, often preserved by polyadenylation (Winata et al., 2018), can be passed from the parent to the offspring to jumpstart gene expression (Ménézo, 2006; Stoeckius et al., 2014; Binder et al., 2015), in addition to various kinds of small RNAs (miRNA, siRNA, piRNA). Other cytoplasmic factors can also impact gene expression in the offspring through direct effects on transcription or by mediating changes to gene methylation or chromatin structure (Kappeler and Meaney, 2010; Moreno-Romero et al., 2017; Tikhodeyev, 2018). Figure 5e shows a hypothetical example in plants where pest pressure experienced by the parent can cause an up-regulation of resistance genes in their offspring via transgenerational effects.

Epigenetic effects are defined here as those mediated by alterations to epigenetic marks. There are many different types of epigenetic effects, however, not all of them are transgenerational or likely serve anticipatory functions. For example, paramutation, genomic imprinting, Polycomb silencing, and position effect variegation are epigenetic phenomena that show no specific relationship to environmental factors but instead are either randomly determined or operate only during development and differentiation (Martin et al., 2007; Golbabapour et al., 2013). Other epigenetic marks, such as gene methylation and histone modifications, can carry information about past expressional states and persist across both mitosis and meiosis. If these marks are maintained in offspring or new cells, they can direct gene expression in adaptive ways. Because these marks serve as a sort of memory of expressional states, they are better candidates for carrying out anticipatory functions (Angers et al., 2010; Bollati and Baccarelli, 2020; Jaenisch and Bird, 2003; Leung and Gaudin, 2020). Gene methylation and histone modifications are the two most well-studied epigenetic marks, and while the causal relationship between gene expression and alterations to these marks are still ambiguous, the correlative patterns between them are reasonably consistent (Holliday, 1990, 2006; Karlič et al., 2010; Deans and Maggert, 2015). It is important to note that the distinction between parental and epigenetic effects hinges on the relationship between the past expressional state and how long it can be transmitted or “remembered”. When a parent is exposed to a particular environment, so too are their germ cells, hence it is impossible to separate the direct effects of gametic or embryonic exposure from the transfer of cytoplasmic compounds during fertilization. In mammals, exposure of an adult male to a particular environment also leads to the exposure of the F0 sperm cells within him. If a pregnant female is exposed to a particular environment, her eggs (F0), embryo (F1), and the primordial germ cells within the embryo (F2) are simultaneously exposed (up to three generations) (Deans and Maggert, 2015; Heard and

Martienssen, 2014). Parental effects, as defined here, are limited to the extent of this exposure, while epigenetic effects can persist in beyond three generations.

Gene methylation refers to the attachment of a methyl group to specific DNA bases. In plants and mammals, methylation most frequently occurs at the C<sup>5</sup>-position of cytosine residues, however, bacteria primarily utilize the methylation of N<sup>6</sup>-position of adenine residues for epigenetic regulation (Sánchez-Romero et al., 2015; Kumar et al., 2018). Gene methylation and demethylation are reversible covalent modifications carried out by methyltransferases. Typically, gene methylation results in the silencing of gene expression via the attraction of repressors, the inhibition of transcription factors, and/or alterations made to chromatin structure (Gibney and Nolan, 2010; Moore et al., 2013). The exact mechanisms that mediate methylation in response to environmental changes is not known, though *de novo* methylation in the parent and the selective maintenance of methylation marks during reprogramming are likely key factors (Angers et al., 2010; Bollati and Baccarelli, 2020). Much more research on the relationship between environmental factors and gene methylation is needed to better establish the role it may play in anticipatory processes, particularly as it relates to causal mechanisms.

Expressional memory can also be maintained and passed on through histone modifications that impact chromatin structure. DNA is packaged into chromatin by being wrapped around histone proteins. Chromatin structure has important implications for gene expression, as transcription can only occur when the chromatin structure is loose enough to allow transcription machinery to interact with the DNA. Modifications on histone tails ultimately control chromatin structure, thus potentially impacting gene expression. For instance, as mentioned in the Priming section, the addition of various methyl and acetyl groups on the lysine residues of histones H3 and H4 are associated with a loosening of chromatin structure and the upregulation of certain genes (Tse et al., 1998; Wang et al., 2001). Similarly to gene methylation, these modifications may be maintained across meiosis and/or mitosis to impact expression in progeny cells and/or offspring (Holeski et al., 2012; Klosin et al., 2017); however, the connection between environmental variables and heritable histone modifications is ambiguous in many taxa, including mammals (Blake and Watson, 2016; Miska and Ferguson-Smith, 2016; van Otterdijk and Michels, 2016).

Although much is still unknown about how and if environmental factors impact parental effects and epigenetic markers, transgenerational effects represent a potential anticipatory mechanism for information about parental environment to be stored and passed on to offspring to better prepare them for similar conditions. These mechanisms meet the criteria of anticipatory processes, as they increase the efficiency with which offspring can respond to their environment. Parental effects and epigenetic markers are also not permanent and can be erased, or simply not maintained, if environmental conditions change, making this strategy flexible enough to respond to a dynamic environment.

### Summary

The anticipatory processes described in this section were selected because they meet at least some of the criteria outlined in the first section, however, this is far from an exhaustive list. There is also considerable overlap between many of these processes. For instance, plant eavesdropping often leads to priming in plants (Kessler, 2006; Morrell and Kessler, 2014). There are also many similarities between hormesis and priming (particularly priming and conditioning hormesis), with the main differences relating to the timeline and generality of the response. One must wonder if these processes represent uniquely evolved responses, different stages of a common evolutionary trajectory, or a shared underlying mechanism. Given that underlying



mechanisms for most of these processes are not well understood, it is also possible that some of these responses are redundant and represent the same mechanism. A good example of this ambiguity can be seen in the phenomenon of DR, as it has been suggested that DR represents a hormetic response but also bears the hallmarks of AAR, with defensive pathways responding to nutritional cues as a general indicator of future stress. Much more work will be needed to understand the relationship between these responses, which for too long have been study in isolation. Perhaps the greatest obstacle will be characterizing the mechanisms at work; a topic which is discussed in the following section.

## **Mechanisms of Anticipation**

A preeminent goal in systems biology is to understand how living systems remain robust to environmental perturbations. We have already discussed the adaptive role that anticipatory processes play in homeostasis/allostasis but have yet to discuss the specific mechanisms that mediate these responses. While information on anticipatory mechanisms is still quite limited, control systems theory, network analyses, and dynamical systems modeling have been broadly employed to understand different types of biological mechanisms. In particular, control system theory has been useful for understanding homeostatic regulation, through the acknowledgement that optimization in both living and manmade systems operates to reduce error. Given that we have already defined anticipatory systems in terms of error, we can use these same principles to discuss potential anticipatory mechanisms. It has also been shown that certain network topologies and designs can be reliably associated with specific system properties, i.e., signal sorting, filtering, delays, synchronization (201-202). While the role that these properties and other types of system dynamics play in anticipatory mechanisms is still unknown, theoretical work has shown that anticipation can be produced under certain designs. For instance, master-slave systems have been shown to produce anticipatory synchronization under certain parameters. The following sections summarize what is currently known and discusses the potential link between existing designs and biological anticipatory mechanisms.

### *Feed-Forward Control*

The primary application of control systems theory in biology acts to understand how biological systems mitigate homeostatic error; particularly how they maintain function in the face of fluctuations (robustness), and how precisely and quickly they can reset back to an undisturbed state (adaptation). Generally speaking, there are two main types of control systems: feed-back and feed-forward control. In a feed-back control system, the output is quantitatively compared to the input to generate a measure of error. This error signal is then filtered out by a feed-back controller that adjusts the input going back into the system. Feed-back control represents a closed-loop design that mitigates error after it occurs, much in the same way that traditional models of homeostasis operate. Alternatively, feed-forward control systems are open designs with controllers that adjust for error before the input enters the system. In this setup, environmental error is predicted or predetermined and applied to the input before it enters the systems to avoid or reduce system error, similarly to the process described in allostasis. There is an obvious connection between feed-forward control and anticipatory processes, as both rely on predictions about future states to minimize error in adaptive ways.

There are a multitude of examples showing how feed-forward control is used in biological processes to deal with environmental fluctuations, particularly in cognitive taxa (203-204) but fewer examples available for non-cognitive organisms (18, 21, 204). Control systems

were discussed by Rosen (2) and later Poli (206-207) in reference to anticipation, yet a strong empirical connection between feed-forward control and biological anticipation remains to be substantiated, although, examples do exist. El-Samad (21) and Shudo (18) describe how the heat-shock response in *E. coli* uses feed-forward control to anticipate the need for heat-shock protein (HSP) production. In the *E. coli*, the transcription of HSP genes is regulated by the  $\sigma^{32}$  protein. The mRNA of the *rpoH* gene, which encodes the  $\sigma^{32}$  protein, is bound to itself at low temperatures but unfolds at high temperatures as the secondary structure relaxes. This allows RNA polymerase to bind to the promoter and transcription/translation of HSPs to take place. The translation of the  $\sigma^{32}$  protein happens before any heat-induced damage to cellular proteins occurs. In this way the  $\sigma^{32}$  protein serves as a temperature sensor and an anticipatory stimulus, and because it modulates the production of HSPs based on an anticipated future need, it represents a feed-forward control system. In addition to HSPs,  $\sigma^{32}$  also up-regulates the expression of the chaperones *DnaK* and *DnaJ*, which control  $\sigma^{32}$  activity through a sequestration feed-back loop by binding  $\sigma^{32}$  at low concentrations of mis-folded proteins. This binding not only halts the expression of HSPs but leads to the degradation of  $\sigma^{32}$  by *Lon* and *FtsH* proteases.

Another example of feed-forward control in *E. coli* is the anticipatory expression of alkalization defense genes by trimethylamine *N*-oxide (TMAO). Bordi et al. (208) describe how the TorS/TorR regulatory system modulates the activity of the anaerobic TMAO reductase respiratory system, which utilizes methylamines as substrates. TMAO is reduced to TMA by TMAO reductase as part of this process and TMA acts as weak base that increases intracellular pH. They found that the presence of TMAO up-regulated the expression of TMAO reductase respiratory machinery but also defense genes related to extreme pH. Again, in this system TMAO acts as an anticipatory stimulus and is used as a predictor of impending pH stress to modulate the expression alkalization defense genes through a feed-forward control design.

I want to note that feed-forward control and feed-forward motifs, which are used to describe regulatory network topologies in biochemical circuits, are not the same thing. In the literature, these concepts are unfortunately easily conflated. Feed-forward loops are common regulatory network motifs found largely in microbial transcriptional networks (28, 201-202, 209-212). They describe three-node networks where one transcription factor directly regulates a target gene and also indirectly regulates the same gene through a secondary transcription factor. There are eight possible configurations of FFLs, which depend on whether the interactions between the nodes are positive or negative and whether they display AND or OR logic (202, 209-210). While incoherent FFL designs are associated with accelerated response-times and have been implicated in perfect adaptation (28), these network motifs do not possess an inherent connection to feed-forward control systems or anticipation and have only been implicated in reactive processes. Although FFLs are likely present in both anticipatory and reactive regulatory circuits, and thus do play a role in relating stimuli with their respective responses, there is currently no evidence for their role in anticipatory functions. In fact, no specific network motif has been implicated in anticipatory mechanisms. As discussed in Lipshtat et al. (213), in order to understand the function of different network motifs they must be placed in the context of other motifs because the overall structure and organization ultimately shapes input-output relationships. This notion is also imperative for understanding anticipatory mechanisms, as the entire system has to be accounted for, from the foremost stimulus to the endpoint of the response, in order to characterize a process as reactive or anticipatory. This is rarely done when studying fine-scale mechanisms, which highlights the importance of understanding biological phenomena on multiple levels; from the evolutionary scale to the biochemical scale. That being said,

determining whether specific network motifs or circuit designs are more prevalent in anticipatory processes will be useful for further elucidating the mechanistic details of these processes.

### *Anticipatory Synchronization*

The specific components of biological systems, including network topology, regulatory motifs, sign-specific interactions, coherence, and logic gating, shape the dynamic properties of biological processes. They impact robustness, gain, bistability, and a multitude of temporal aspects, such as the presence of pulses, delays, oscillations, and response-time. In dynamical systems modeling, the goal is to understand how different system properties impact input-output relationships, particularly differences in reaction kinetics. Although system designs can alter the timing of reactions, thereby impacting the dynamics of the system, it is unclear how or if changes in system dynamics alone can be used to make systems anticipatory. Of the five different types of anticipatory processes discussed in the last section, only two clearly implicate a role for system dynamics in producing the anticipatory quality of the response. Circadian rhythms employ an internal clock to time responses in an anticipatory manner and hormesis uses low-response thresholds to produce compensatory responses. Priming and transgenerational effects, on the other hand, rely primarily on long-term memory mechanisms. Although the role of this memory could arguably be incorporated into a dynamic systems model, the system is ultimately dependent on multiple events controlled by environmental factors, making the internal dynamics more difficult to accurately model. Processes controlled by AAR, as discussed earlier, are more likely the result of feed-forward control than changes in system kinetics, hence their anticipatory nature stems from changes in the stimulus controlling the reaction rather than system dynamics. Much more work is needed to accurately model anticipatory processes and determine whether they possess unique dynamics, but progress has been made in showing that alterations to system dynamics can at least theoretically produce anticipation.

The production of anticipatory synchronized oscillations is an example of one way that system dynamics can mediate anticipation. Anticipatory synchronization (AS) can be achieved by coupling two identical dynamic systems in a master-slave configuration (214-215). By subjecting the slave to a certain degree of delayed negative feedback, a stable state emerges where the state of the slave at time  $t$  is in the same state that the master will be in at a future time. In this way, the slave anticipates the future state of the master through AS. Anticipatory synchronization has been demonstrated in diverse models, including discrete and continuous systems, as well as noisy, chaotic, excitable, and quasi-periodic dynamic systems (214, 216-220), producing both short- and long-term anticipation times (221). Anticipatory synchronization has been experimentally demonstrated using electronic circuits (219, 222-223) and lasers (224-228). Matias et al. (215) showed that AS could be produced in a 3-neuron Hodgkin-Huxley type model under certain parameters, and Ciszak et al. (223) even showed how AS could be used to suppress spikes in a perturbed FitzHugh-Nagumo neuronal system by incorporating AS into a feed-forward design. Although oscillations are common in living things, including circadian rhythms, cytoplasmic oscillators such as glycolytic cycles, cell cycles, neuronal membrane oscillators, and transcriptional oscillations, it isn't yet established whether the AS found in physical systems are also present in biological systems.

### **Conclusions**

An unavoidable disadvantage of working within any paradigm, is that we are only able to see those things that fit inside of it. It is an age-old epistemological problem. Rosen's view was

that modern science, being largely shaped by the fields of physics and mathematics, leads us to view biology in a purely “reactive” way that has marginalized the role of anticipation in biological systems. Whether you agree with Rosen’s supposition or not, the topics of anticipation and predictive processes have been curiously limited in the greater biological sciences, despite a history of evidence that spans over 100 years. One could invoke the law of parsimony to justify or explain this, as anticipatory designs do seem less intuitive and perhaps more complex than reactive ones; but are they really? Are feed-forward systems more complex than feed-back systems? I would argue that there is no inherent difference in complexity, simply a difference in organization. In any case, if a primary goal of biology is to understand the diversity of biological processes and functions, the argument is largely moot, for anticipatory processes most definitely exist and therefore must have a place in the field of biology.

It may seem bombastic to suggest that the accounting for anticipatory processes requires a reworking of fundamental biological principles; however, the dichotomy between reactive and anticipatory processes is foundational. The differences between them occur at a causal level and, as such, must affect all successive qualities. Making this change doesn’t require the destruction of any standing principles, simply the addition of new considerations. Much like the acknowledgement of epigenetics has not erased Mendelian principles but simply expanded the field in a way that has helped explain many aspects of gene function that would otherwise remain inexplicable. Anticipatory mechanisms need not defy Newtonian principles. They don’t even require additional parts; simply the reorganized of existing parts.

Much more work is needed to further understand anticipatory processes, including their evolution and role in biological function. This effort will require identifying new anticipatory processes in new organisms and further characterization of anticipatory mechanisms, but this can only be accomplished if we know what to look for. It is my hope that the criteria and examples provided in this review will help inform and direct future research and ultimately expand our understanding of complex biological processes.

### Data Accessibility Statement

Given that this is a review, there is no original data presented in this article except for the figures. If accepted for publication, all figures will be published on figshare (<https://figshare.com/>) and made available to the public.

### References

- Agutter, P. S. (2007) Cell mechanics and stress: from molecular details to the ‘universal cell reaction’ and hormesis. *BioEssays* **29**, 324-333.
- Ahlers, V., Parlitz, U., & Lauterborn, W. (1998) Hyperchaotic dynamics and synchronization of external-cavity semiconductor lasers. *Physical Rev E*, **58**, 7208.
- Ahmad, S., Ahmad, A., Gerasimovskaya, E., Stenmark, K. R., Allen, C. B., & White, C. W. (2003) Hypoxia protects human lung microvascular endothelial and epithelial-like cells against oxygen toxicity: role of phosphatidylinositol 3-kinase. *American Journal of Respiratory Cell and Molecular Biology*, **28**, 179-187.

Ahn, J. B., Hwang, H. J., & Park, J. H. (2001) Physiological responses of oxygen-tolerant anaerobic *Bifidobacterium longum* under oxygen. *Journal of Microbiology and Biotechnology*, **11**, 443-451.

Albanesi, D., Reh, G., Guerin, M. F., Schaeffer, F., Debarbouille, M., Buschiazzi, A., Schujman, G. E., De Mendoza, D., & Alzari, P. M. (2013) Structural basis for feed-forward transcriptional regulation of membrane lipid homeostasis in *Staphylococcus aureus*. *PLoS Pathogens*, **9**, e1003108.

Albrecht, U. (2012) *The Circadian Clock* (Vol. 12). Springer Science & Business Media, New York, NY.

Alon, U. (2007) Network motifs: theory and experimental approaches. *Nature Reviews Genetics* **8**, 450-461.

Alvarez-Venegas, R., Al Abdallat, A., Guo, M., Alfano, J. R., & Avramova, Z. (2007) Epigenetic control of a transcription factor at the cross section of two antagonistic pathways. *Epigenetics*, **2**, 106-113.

Ananta, E. and Knorr, D. (2003) Pressure-induced thermotolerance of *Lactobacillus rhamnosus* GG. *International Food Research Journal*, **36**, 991-997.

Angers, B., Castonguay, E., & Massicotte, R. (2010) Environmentally induced phenotypes and DNA methylation: how to deal with unpredictable conditions until the next generation and after. *Molecular Ecology*, **19**, 1283-1295.

Badyaev, A. V. & Uller, T. (2009) Parental effects in ecology and evolution: mechanisms, processes and implications. *Philosophical Transactions of the Royal Society B: Biological Sciences*, **364**, 1169-1177.

Balmer, A., Pastor, V., Gamir, J., Flors, V., & Mauch-Mani, B. (2015) The ‘prime-ome’: towards a holistic approach to priming. *Trends in Plant Science*, **20**, 443-452.

Bauman, D. E. & Currie, W. B. (1980) Partitioning of nutrients during pregnancy and lactation: a review of mechanisms involving homeostasis and homeorhesis. *Journal of Dairy Science*, **63**, 1514-1529.

Baumgard, L. H., Collier, R. J., & Bauman, D. E. (2017) A 100-Year Review: Regulation of nutrient partitioning to support lactation. *Journal of Dairy Science*, **100**, 10353-10366.

Beckers, G. J., Jaskiewicz, M., Liu, Y., Underwood, W. R., He, S. Y., Zhang, S., & Conrath, U. (2009) Mitogen-activated protein kinases 3 and 6 are required for full priming of stress responses in *Arabidopsis thaliana*. *Plant Cell*, **21**, 944-953.

Bektas, Y. & Eulgem, T. (2015) Synthetic plant defense elicitors. *Frontiers in Plant Science*, **5**, 804. Belhomme, D., Peynet, J., Louzy, M., Launay, J. M., Kitakaze, M., & Menasché, P. (1999)



Evidence for preconditioning by isoflurane in coronary artery bypass graft surgery. *Circulation*, 100, 11340-11344.

Ben-Jacob, E. & Schultz, D. (2010) Bacteria determine fate by playing dice with controlled odds. *Proceedings of the National Academy of Sciences*, **107**, 13197-13198.

Bernhardt, W. M., Câmpean, V., Kany, S., Jürgensen, J. S., Weidemann, A., Warnecke, C., Arend, M., Klaus, S., Günzler, V., Amann, K., & Willam, C. (2006) Preconditional activation of hypoxia-inducible factors ameliorates ischemic acute renal failure. *Journal of the American Society of Nephrology* **17**, 1970-1978.

Berry, D. B. & Gasch, A. P. (2008) Stress-activated genomic expression changes serve a preparative role for impending stress in yeast. *Molecular Biology of the Cell*, **19**, 4580-4587.

Binder, A. M., LaRocca, J., Lesseur, C., Marsit, C. J., & Michels, K. B. (2015) Epigenome-wide and transcriptome-wide analyses reveal gestational diabetes is associated with alterations in the human leukocyte antigen complex. *Clinical Epigenetics*, **7**, 79.

Blake, G. E. & Watson, E. D. (2016) Unravelling the complex mechanisms of transgenerational epigenetic inheritance. *Current Opinion in Chemical Biology*, **33**, 101-107.

Blande, J. D. (2017) Plant Communication with Herbivores In *Advances in Botanical Research* (Vol. 82, 281-304). Academic Press, Cambridge, MA.

Bollati, V. & Baccarelli, A. (2020) Environmental epigenetics. *Heredity*, **105**, 105-112.

Bordi, C., Théraulaz, L., Méjean, V., & Jourlin-Castelli, C. (2003) Anticipating an alkaline stress through the Tor phosphorelay system in *Escherichia coli*. *Molecular Microbiology*, **48**, 211-223.

Boulos, Z., Rosenwasser, A. M., & Terman, M. (1980) Feeding schedules and the circadian organization of behavior in the rat. *Behavioural Brain Research*, **1**, 39-65.

Boutibonnes, P., Giard, J. C., Hartke, A., Thamavongs, B., & Auffray, Y. (1993) Characterization of the heat shock response in *Enterococcus faecalis*. *Antonie Van Leeuwenhoek International Journal of General and Molecular Microbiology*, **64**, 47-55.

Bhadra, U., Thakkar, N., Das, P., & Bhadra, M. P. (2017) Evolution of circadian rhythms: from bacteria to human. *Sleep Medicine*, **35**, 49-61.

Brennan, M. D., Cheong, R., & Levchenko, A. (2012) How information theory handles cell signaling and uncertainty. *Science*, **338**, 334-335.

Bubic, A., Von Cramon, D. Y., & Schubotz, R. I. (2010) Prediction, cognition and the brain. *Frontiers in Human Neuroscience*, **4**, 25-24.

Buldú, J. M., Rogister, F., Trull, J., Serrat, C., Torrent, M. C., García-Ojalvo, J., & Mirasso, C. R. (2002) Asymmetric and delayed activation of side modes in multimode semiconductor lasers with optical feedback. *Journal of Optics B: Quantum and Semiclassical Optics*, **4**, 415.

5 van den Burg, H. A. & Takken, F. L. W. (2009) Does chromatin remodeling mark systemic acquired resistance? *Trends Plant Science*, **14**, 286–294.

Burgers, J. M. (1975) Causality and Anticipation: Analysis of the concept of anticipation can contribute to the philosophy of biology. *Science* **189**, 194-198.

10 Butz, M. V. (2008) How and why the brain lays the foundations for a conscious self. *Constructivist Foundations*, **4**, 1-14.

Calabrese, E. J. (2014) Hormesis: a fundamental concept in biology. *Microbial Cell*, **1**, 145.

15 Calabrese, E. J. (2002) Hormesis: changing view of the dose-response, a personal account of the history and current status. *Mutation Research/Reviews*, **511**, 181-189.

20 Calabrese, E. J., Bachmann, K. A., Bailer, A. J., Bolger, P. M., Borak, J., Cai, L., Cedergreen, Cherian, N. M. G., Chiueh, C. C., Clarkson, T. W., & Cook, R. R. (2007) Biological stress response terminology: integrating the concepts of adaptive response and preconditioning stress within a hormetic dose–response framework. *Toxicology and Applied Pharmacology*, **222**, 122-128.

25 Calabrese, E. J. & Baldwin, L. A. (1997) The dose determines the stimulation (and poison): development of a chemical hormesis database. *International Journal of Toxicology*, **16**, 545-559.

Calabrese, E. J. & Baldwin, L. A. (2001) Hormesis: U-shaped dose responses and their centrality in toxicology. *Trends Pharmacological Science*, **22**, 285-291.

30 Calabrese, E. J., Stanek III, E. J., Nascarella, M. A., & Hoffmann, G. R. (2008) Hormesis predicts low-dose responses better than threshold models. *International Journal of Toxicology*, **27**, 369-378.

35 Calvo, P. & Baluška, F. (2015) Conditions for minimal intelligence across eukaryota: a cognitive science perspective. *Frontiers in Psychology*, **6**, 1329.

Calvo, O., Chialvo, D. R., Eguíluz, V. M., Mirasso, C., & Toral, R. (2004) Anticipated synchronization: A metaphorical linear view. *Chaos*, **14**, 7-13.

40 Cannon, W. B. (1929) Organization for physiological homeostasis. *Physiological Reviews*, **9**, 399–431,

45 Cizak, M., Calvo, O., Masoller, C., Mirasso, C. R., & Toral, R. (2003) Anticipating the response of excitable systems driven by random forcing. *Physical Review Letters*, **90**, 204102.

Ciszak, M., Mirasso, C. R., Toral, R., & Calvo, O. (2009) Predict-prevent control method for perturbed excitable systems. *Physical Review E*, **79**, 046203.

5 Cohen, S. E. & Golden, S. S. (2015) Circadian rhythms in cyanobacteria. *Microbiology and Molecular Biology Reviews*, **79**, 373-385.

Conrath, U. (2011) Molecular aspects of defence priming. *Trends Plant Science*, **16**, 524-531.

10 Conrath, U., Beckers, G. J., Langenbach, C. J., & Jaskiewicz, M. R. (2015) Priming for enhanced defense. *Annual Review of Phytopathology*, **53**, 97-119.

Davidson, A. J., Poole, A. S., Yamazaki, S., & Menaker, M. (2003) Is the food-entrainable circadian oscillator in the digestive system? *Genes Brain Behavior*, **2**, 32-39.

15 De Angelis, M. & Gobetti, M. (2004) Environmental stress responses in *Lactobacillus*: a review. *Proteomics*, **4**, 106-122.

20 Desmond, C., Fitzgerald, G. F., Stanton, C., & Ross, R. P. (2004) Improved stress tolerance of GroESL-overproducing *Lactococcus lactis* and probiotic *Lactobacillus paracasei* NFBC 338. *Applied Environmental Microbiology*, **70**, 5929-5936.

Deans, C. & Maggert, K. A. (2015) What do you mean, “epigenetic”? *Genetics*, **199**, 887-896.

25 Dhar, R., Sägesser, R., Weikert, C., & Wagner, A. (2013) Yeast adapts to a changing stressful environment by evolving cross-protection and anticipatory gene regulation. *Molecular Biology and Evolution*, **30**, 573-588.

Dubois, D. & Resconi, G. (1992) Hyperincursivity: A New Mathematical Theory. Presses Universitaires de Liège, Liège.

30 Dubois, D. M. (2003) Mathematical foundations of discrete and functional systems with strong and weak anticipations In *Anticipatory Behavior in Adaptive Learning Systems* (110-132). Springer, Berlin, Heidelberg.

35 Dunlap, J. C. (1999) Molecular bases for circadian clocks. *Cell*, **96**, 271-290.

Duwat, P., Cesselin, B., Sourice, S., & Gruss, A. (2000) *Lactococcus lactis*, a bacterial model for stress responses and survival. *International Journal of Food Microbiology*, **55**, 83-86.

40 El-Samad, H. (2010) Control strategies in times of adversity: how organisms survive stressful conditions In *Control Theory and System Biology* (85-99), MIT Press, Cambridge, MA.

45 Engelberth, J., Alborn, H. T., Schmelz, E. A., & Tumlinson, J. H. (2004) Airborne signals prime plants against insect herbivore attack. *Proceedings of the National Academy of Sciences*, **101**, 1781-1785.

Eoh, H., Wang, Z., Layre, E., Rath, P., Morris, R., Moody, D. B., & Rhee, K. Y. (2017) Metabolic anticipation in *Mycobacterium tuberculosis*. *Nature Microbiology*, **2**, 17084.

5 Fajas, L., Fruchart, J. C., & Auwerx, J. (1998) Transcriptional control of adipogenesis. *Current Opinions in Cell Biology*, **10**, 165-173.

Falik, O., Mordoch, Y., Quansah, L., Fait, A., & Novoplansky, A. (2011) Rumor has it...: relay communication of stress cues in plants. *PLoS One*, **6**, e23625.

10 Feynman, R. P. (1982) Simulating physics with computers. *International Journal of Theoretical Physics*, **21**, 467– 488.

Fontana, L., Partridge, L., & Longo, V. D. (2010) Extending healthy life span—from yeast to humans. *Science*, **328**, 321-326.

15 Forbes, V. E. (2000) Is hormesis an evolutionary expectation? *Functional Ecology*, **14**, 12-24.

Foster, J. W. & Hall, H. K. (1991) Inducible pH homeostasis and the acid tolerance response of *Salmonella typhimurium*. *Journal of Bacteriology*, **173**, 5129-5135.

20 Franks, P. J., Cowan, I. R., & Farquhar, G. D. (1997) The apparent feedforward response of stomata to air vapour pressure deficit: information revealed by different experimental procedures with two rainforest trees. *Plant, Cell, & Environment*, **20**, 142-145.

25 Freddolino, P. L. & Tavazoie, S. (2012) Beyond homeostasis: a predictive-dynamic framework for understanding cellular behavior. *Annual Review of Cell and Developmental Biology*, **28**, 363-384.

30 Fuller, C. A., Sulzman, F. M., & Moore-Ede, M. C. (1985) Role of heat loss and heat production in generation of the circadian temperature rhythm of the squirrel monkey. *Physiology & Behavior*, **34**, 543-546.

35 Hoffmann, J., Berner, M., Butz, M. V., Herbort, O., Kiesel, A., Kunde, W., & Lenhard, A. (2007) Explorations of anticipatory behavioral control (ABC): a report from the Cognitive Psychology Unit of the University of Wurzburg. *Cognitive Processing*, **8**, 133–142.

40 Hao, Z., Fayolle, L., van Tuinen, D., Chatagnier, O., Li, X., Gianinazzi, S., & Gianinazzi-Pearson, V. (2012) Local and systemic mycorrhiza-induced protection against the ectoparasitic nematode *Xiphinema index* involves priming of defence gene responses in grapevine. *Journal of Experimental Botany*, **63**, 3657-3672.

Hartke, A., Bouche, S., Gansel, X., Boutibonnes, P., & Auffray, Y. (1994) Starvation-induced stress resistance in *Lactococcus lactis* subsp. *lactis* IL1403. *Applied and Environmental Microbiology*, **60**, 3474-3478.

Holeski, L. M., Jander, G., & Agrawal, A. A. (2012) Transgenerational defense induction and epigenetic inheritance in plants. *Trends in Ecology & Evolution*, **27**, 618-626.

Horowitz, M. (2016) Epigenetics and cytoprotection with heat acclimation. *Journal of Applied Physiology*, **120**, 702-710.

van Hulten, M., Pelser, M., Van Loon, L. C., Pieterse, C. M., & Ton, J. (2006) Costs and benefits of priming for defense in Arabidopsis. *Proceedings of the National Academy of Sciences*, **103**, 5602-5607.

Gamir, J., Pastor, V., Cerezo, M., & Flors, V. (2012) Identification of indole-3-carboxylic acid as mediator of priming against *Plectosphaerella cucumerina*. *Plant Physiology and Biochemistry*, **61**, 169-179.

Gems, D. & Partridge, L. (2008) Stress-response hormesis and aging: “that which does not kill us makes us stronger”. *Cell Metabolism*, **7**, 200-203.

Gibney, E. R. & Nolan, C. M. (2010) Epigenetics and gene expression. *Heredity*, **105**, 4-13.

Golbabapour, S., Majid, N. A., Hassandarvish, P., Hajrezaie, M., Abdulla, M. A., & Hadi, A. H. A. (2013) Gene silencing and Polycomb group proteins: an overview of their structure, mechanisms and phylogenetics. *OMICS*, **17**, 283-296.

Hara, E. & Saito, M. (1980) Diurnal changes in plasma glucose and insulin responses to oral glucose load in rats. *American Journal of Physiology*, **238**, E463-E466.

Harmer, S. L., Panda, S., & Kay, S. A. (2001) Molecular bases of circadian rhythms. *Annual Review of Cell and Developmental Biology*, **17**, 215-253.

Hayes, M. P., Freeman, S. L., & Donnelly, R. P. (1995) IFN- $\gamma$  priming of monocytes enhances LPS-induced TNF production by augmenting both transcription and mRNA stability. *Cytokine*, **7**, 427-435.

Heard, E. & Martienssen, R. A. (2014) Transgenerational epigenetic inheritance: myths and mechanisms. *Cell*, **157**, 95-109.

Heil, T., Fischer, I., Elsässer, W., Mulet, J., & Mirasso, C. R. (2001) Chaos synchronization and spontaneous symmetry-breaking in symmetrically delay-coupled semiconductor lasers. *Physical Review Letters*, **86**, 795.

Hernández-García, C. Masoller, and Mirasso, C. R. (2002) Anticipating the dynamics of chaotic maps. *Physica A*, **295**, 39-43.

Holliday, R. (1990) DNA methylation and epigenetic inheritance. *Philosophical Transactions of the Royal Society B: Biological Sciences*, **326**, 329-338.



Holliday, R. (2006) Epigenetics: a historical overview. *Epigenetics*, **1**, 76-80.

Jaenisch, R. & Bird, A. (2003) Epigenetic regulation of gene expression: how the genome integrates intrinsic and environmental signals. *Nature Genetics*, **33**, 245-254.

Jaskiewicz, M., Conrath, U., & Peterhänzel, C. (2011) Chromatin modification acts as a memory for systemic acquired resistance in the plant stress response. *EMBO Reports*, **12**, 50-55.

Johnson, C. H., Golden, S. S., Ishiura, M., & Kondo, T. (1996) Circadian clocks in prokaryotes. *Molecular Microbiology*, **21**, 5-11.

Johnson, C. H. & Kondo, T. (2011) Circadian Rhythms in Unicellular Organisms In *Circadian Clocks* (61-77). Springer, Boston, MA.

Joyeux-Faure, M., Stanke-Labesque, F., Lefebvre, B., Beguin, P., Godin-Ribuot, D., Ribaut, C., Launois, S. H., Bessard, G., & Levy, P. (2005) Chronic intermittent hypoxia increases infarction in the isolated rat heart. *Journal of Applied Physiology*, **98**, 1691-1696.

Jung, H. W., Tschaplinski, T. J., Wang, L., Glazebrook, J., & Greenberg, J. T. (2009) Priming in systemic plant immunity. *Science*, **324**, 89-91.

Kappeler, L. & Meaney, M. J. (2010) Epigenetics and parental effects. *BioEssays*, **32**, 818-827.

Karban, R., Maron, J., Felton, G. W., Ervin, G., & Eichenseer, H. (2003) Herbivore damage to sagebrush induces resistance in wild tobacco: evidence for eavesdropping between plants. *Oikos*, **100**, 325-332.

Karlič, R., Chung, H. R., Lasserre, J., Vlahoviček, K., & Vingron, M. (2010) Histone modification levels are predictive for gene expression. *Proceedings of the National Academy of Sciences*, **107**, 2926-2931.

Kessler, A., Halitschke, R., Diezel, C., & Baldwin, I. T. (2006) Priming of plant defense responses in nature by airborne signaling between *Artemisia tridentata* and *Nicotiana attenuata*. *Oecologia*, **148**, 280-292.

King, W. I. (1938) The causes of economic fluctuations: possibilities of anticipation and control. Ronald Press Co, New York.

Kim, J. & Felton, G. W. (2013) Priming of antiherbivore defensive responses in plants. *Insect Science*, **20**, 273-285.

Klir, G. J. (2002) Uncertainty in economics: the heritage of G.L.S. Shackle. *Fuzzy Economic Review*, **7**, 3-21.

Klosin, A., Casas, E., Hidalgo-Carcedo, C., Vavouri, T., & Lehner, B. (2017) Transgenerational transmission of environmental information in *C. elegans*. *Science*, **356**, 320-323.

Kumar, S., Chinnusamy, V., & Mohapatra, T. (2018) Epigenetics of modified DNA bases: 5-methylcytosine and beyond. *Frontiers in Genetics*, **9**, 640.

5 Kussell, E. & Leibler, S. (2005) Phenotypic diversity, population growth, and information in fluctuating environments. *Science*, **309**, 2075-2078.

10 Kraska, T. & Schönbeck, F. (1993) About changes in the chromatin structure after resistance induction in *Hordeum vulgare* L. *J. Phytopathology*, **137**, 10-14.

Libet, B., Gleason, C. A., Wright, E. W., & Pearl, D. K. (1983) Time of conscious intention to act in relation to onset of cerebral-activity (readiness-potential)—The unconscious initiation of a freely voluntary act. *Brain*, **10**, 623-42.

15 Libet, B. (1985) Unconscious cerebral initiative and the role of conscious will in voluntary action. *Behavioral and Brain Sciences*, **8**, 529–39.

Latty, T. & Beekman, M. (2010) Food quality and the risk of light exposure affect patch-choice decisions in the slime mold *Physarum polycephalum*. *Ecology* **91**, 22-27.

20 Lee, K. P., Simpson, S. J., Clissold, F. J., Brooks, R., Ballard, J. W. O., Taylor, P. W., Soran, N., & Raubenheimer, D. (2008) Lifespan and reproduction in *Drosophila*: new insights from nutritional geometry. *Proceedings of the National Academy of Sciences*, **105**, 2498-2503.

25 Leung, J. & Gaudin, V. (2020) Who rules the cell? an epi-tale of histone, DNA, RNA, and the metabolic deep state. *Frontiers in Plant Science*, **11**, 181.

30 Lim, S., Borza, T., Peters, R. D., Coffin, R. H., Al-Mughrabi, K. I., Pinto, D. M., & Wang-Pruski, G. (2013) Proteomics analysis suggests broad functional changes in potato leaves triggered by phosphites and a complex indirect mode of action against *Phytophthora infestans*. *Journal of Proteomics*, **93**, 207-223.

Lipshtat, A., Purushothaman, S. P., Iyengar, R., & Ma'ayan, A. (2008) Functions of bifans in context of multiple regulatory motifs in signaling networks. *Biophysical Journal*, **94**, 2566-2579.

35 Liu, Y., Takiguchi, Y., Davis, P., Aida, T., Saito, S., & Liu, J. M. (2002) Experimental observation of complete chaos synchronization in semiconductor lasers. *Applied Physics Letters*, **80**, 4306-4308.

40 Liu, M., Durfee, T., Cabrera, J. E., Zhao, K., Jin, D. J., & Blattner, F. R. (2005) Global transcriptional programs reveal a carbon source foraging strategy by *Escherichia coli*. *Journal of Biological Chemistry*, **280**, 15921-15927.

45 Leung, C., Angers, B., & Bergeron, P. (2020) Epigenetic anticipation for food and reproduction. *Environmental Epigenetics*, **6**, 1-8.

Lu, H., Wang, Y., He, X., Yuan, F., Lin, X., Xie, B., Tang, G., Huang, J., Tang, Y., Jin, K., & Chen, S. (2012) Netrin-1 hyperexpression in mouse brain promotes angiogenesis and long-term neurological recovery after transient focal ischemia. *Stroke*, **43**, 838-843.

5 Ma, J., Chen, T., Mandelin, J., Ceponis, A., Miller, N. E., Hukkanen, M., Ma, G. F., & Konttinen, Y. T. (2003) Regulation of macrophage activation. *Cell Mol Life Sci*, **60**, 2334-2346.

Mair, W. & Dillin, A. (2008) Aging and survival: the genetics of life span extension by dietary restriction. *Annual Review of Biochemistry*, **77**, 727-754.

10 Mair, W., Piper, M. D., & Partridge, L. (2005) Calories do not explain extension of life span by dietary restriction in *Drosophila*. *PLoS Biology*, **3**, e223.

15 Mangan, S. & Alon, U. (2003) Structure and function of the feed-forward loop network motif. *Proceedings of the National Academy of Sciences*, **100**, 11980-11985.

Mangan, S., Zaslaver, A., & Alon, U. (2003) The coherent feedforward loop serves as a sign-sensitive delay element in transcription networks. *Journal of Molecular Biology*, **334**, 197-204.

20 Mank, N. N., Berghoff, B. A., & Klug, G. (2013) A mixed incoherent feed-forward loop contributes to the regulation of bacterial photosynthesis genes. *RNA Biology*, **10**, 347-352.

25 March-Díaz, R., García-Domínguez, M., Lozano-Juste, J., León, J., Florencio, F. J., Reyes, J. C. (2008) Histone H2A.Z and homologues of components of the SWR1 complex are required to control immunity in *Arabidopsis*. *The Plant Journal*, **53**, 475–487.

Martin, C. & Zhang, Y. (2007) Mechanisms of epigenetic inheritance. *Current Opinions in Cell Biology*, **19**, 266-272.

30 Martínez-Medina, A., Fernández, I., Sánchez-Guzmán, M. J., Jung, S. C., Pascual, J. A., & Pozo, M. J. (2013) Deciphering the hormonal signalling network behind the systemic resistance induced by *Trichoderma harzianum* in tomato. *Frontiers in Plant Science*, **4**, 206.

35 Masoro, E. J. (2007) The Role of Hormesis in Life Extension by Dietary Restriction In *Mechanisms of Dietary Restriction in Aging and Disease* (Vol. 35, 1-17). Karger Publishers, Basel, CH.

40 Masoller, C. (2001) Anticipation in the synchronization of chaotic semiconductor lasers with optical feedback. *Physical Review Letters*, **86**, 2782.

Masoller, C. & Zanette, D. H. (2003) Different regimes of synchronization in nonidentical time-delayed maps. *Physica A*, **325**, 361-370.

45 Massoud, K., Barchietto, T., Le Rudulier, T., Pallandre, L., Didierlaurent, L., Garmier, M., Ambard-Bretteville, F., Seng, J. M., & Saindrenan, P. (2012) Dissecting phosphite-induced

priming in *Arabidopsis* infected with *Hyaloperonospora arabidopsidis*. *Plant Physiology*, **159**, 286-298.

Matias, F.S., Carelli, P.V., Mirasso, C.R., Copelli, M. (2011) Anticipated synchronization in a biologically plausible model of neuronal motifs. *Physical Reviews E*, **84**, 021922.

Matveev, V. V. (2005) Protoreaction of protoplasm. *Cellular and Molecular Biology*, **51**, 715-23.

Matveev, V. V. (2010) Native aggregation as a cause of origin of temporary cellular structures needed for all forms of cellular activity, signaling and transformations. *Theoretical Biology and Medical Modelling*, **7**, 19.

McCay, C.M., Crowell, M.F., Maynard, L.A. (1935) The effect of retarded growth upon the length of life and upon the ultimate body size. *Journal of Nutrition*, **10**, 63-79.

McEwen, B. S. (1998) Stress, adaptation, and disease: allostasis and allostatic load. *Annals of the New York Academy of Sciences*, **840**, 33-44.

Ménézo, Y. J. (2006) Paternal and maternal factors in preimplantation embryogenesis: interaction with the biochemical environment. *Reproductive BioMedicine Online*, **12**, 616-621.

Meller, R., Pearson, A., & Simon, R. P. (2015) Dynamic changes in DNA methylation in ischemic tolerance. *Frontiers in Neurology*, **6**, 102.

Mrosovsky, N. (1990) Rheostasis: The Physiology of Change. Oxford University Press.

Millar, A. J. & Kay, S. A. (1996) Integration of circadian and phototransduction pathways in the network controlling CAB gene transcription in *Arabidopsis*. *Proceedings of the National Academy of Sciences*, **93**, 15491-15496.

Milo, R., Itzkovitz, S., Kashtan, N., Levitt, R., Shen-Orr, S., Ayzenshtat, I., Sheffer, M., & Alon, U. (2004) Superfamilies of evolved and designed networks. *Science*, **303**, 1538-1542.

Miska, E. A. & Ferguson-Smith, A. C. (2016) Transgenerational inheritance: models and mechanisms of non-DNA sequence-based inheritance. *Science*, **354**, 59-63.

Mistlberger, R. E. (1994) Circadian food-anticipatory activity: formal models and physiological mechanisms. *Neuroscience & Biobehavior Reviews*, **18**, 171-195.

Mitchell, I. A. & Pilpel, Y. (2011) A mathematical model for adaptive prediction of environmental changes by microorganisms. *Proceedings of the National Academy of Sciences*, **108**, 7271-7276.

Mitchell, A., Romano, G. H., Groisman, B., Yona, A., Dekel, E., Kupiec, M., Dahan, O., & Pilpel, Y. (2009) Adaptive prediction of environmental changes by microorganisms. *Nature*, **460**, 220-224.

Mittag, M. & Wagner, V. (2003) The circadian clock of the unicellular eukaryotic model organism *Chlamydomonas reinhardtii*. *Biological Chemistry*, **384**, 689-695.

Moatt, J. P., Nakagawa, S., Lagisz, M., & Walling, C. A. (2016) The effect of dietary restriction on reproduction: a meta-analytic perspective. *BMC Evolutionary Biology*, **16**, 199.

Molitor, A., Zajic, D., Voll, L. M., Pons-Kühnemann, J., Samans, B., Kogel, K. H., & Waller, F. (2011) Barley leaf transcriptome and metabolite analysis reveals new aspects of compatibility and *Piriformospora indica*-mediated systemic induced resistance to powdery mildew. *Molecular Plant-Microbe Interactions*, **24**, 1427-1439.

Moore-Ede, M. C. (1986) Physiology of the circadian timing system: predictive versus reactive homeostasis. *American Journal of Physiology-Regulatory Integrative and Comparative Physiology*, **250**, 735-752.

Moore-Ede, M. C., Sulzman, F. M., & Fuller, C. A. (1982) *Clocks That Time Us: Physiology of the Circadian Timing System*. Cambridge, Harvard University Press, Cambridge, MA.

Moore, L. D., Le, T., & Fan, G. (2013) DNA methylation and its basic function. *Neuropsychopharmacology*, **38**, 23-38.

Moreno-Romero, J., Santos-González, J., Hennig, L., & Köhler, C. (2017) Applying the INTACT method to purify endosperm nuclei and to generate parental-specific epigenome profiles. *Nature Protocols*, **12**, 238-254.

Morrell, K. & Kessler, A. (2014) The scent of danger: Volatile-mediated information transfer and defence priming in plants. *The Biochemist*, **36**, 26-31.

Murry, C. E., Jennings, R. B., & Reimer, K. A. (1986) Preconditioning with ischemia: a delay of lethal cell injury in ischemic myocardium. *Circulation*, **74**, 1124-1136.

Nadin, M. (2010) Anticipation and dynamics: Rosen's anticipation in the perspective of time. *International Journal of General Systems*, **39**, 3-33.

Nadin, M. (2016) *Anticipation Across Disciplines*. Springer International Publishing, New York, NY.

Nasonov, D. N. (1962) *Local Reaction of Protoplasm and Gradual Excitation* [in Russian], USSR Academy of Sciences Publishing, Moscow-Leningrad.

Nohales, M. A. & Kay, S. A. (2016) Molecular mechanisms at the core of the plant circadian oscillator. *Nature Structural & Molecular Biology*, **23**, 1061-1069.

Novoplansky, A. (2016) Future perception in plants In *Anticipation Across Disciplines* (57-70). Springer, Cham.



O'Neill, J. S., Van Ooijen, G., Dixon, L. E., Troein, C., Corellou, F., Bouget, F. Y., Reddy, A. B., & Millar, A. J. (2011) Circadian rhythms persist without transcription in a eukaryote. *Nature*, **469**, 554-558.

5 van Otterdijk, S. D. & Michels, K. B. (2016) Transgenerational epigenetic inheritance in mammals: how good is the evidence? *The FASEB Journal*, **30**, 2457-2465.

Pastor, V., Luna, E., Mauch-Mani, B., Ton, J., & Flors, V. (2013) Primed plants do not forget. *Environmental and Experimental Biology*, **94**, 46-56.

10 Pavlov, I. P. (1902) The Work of the Digestive Glands. Charles Griffin.

Piper, M. D., Partridge, L., Raubenheimer, D., & Simpson, S. J. (2011) Dietary restriction and aging: a unifying perspective. *Cell Metabolism*, **14**, 154-160.

15 Poli, R. (2009) The complexity of anticipation. *Balkan Journal of Philosophy*, **1**, 19-29.

Poli, R. (2010) The many aspects of anticipation. *Foresight*, **12**, 7-17.

20 Power, M. L. & Schulkin, J. (2008) Anticipatory physiological regulation in feeding biology: cephalic phase responses. *Appetite*, **50**, 194-206.

Powley, T. L. (1977) The ventromedial hypothalamic syndrome, satiety, and a cephalic phase hypothesis. *Psychological Review*, **84**, 89.

25 Pyragas K. & Pyragienė, T. (2008) Coupling design for a long-term anticipating synchronization of chaos. *Physical Reviews E*, **78**, 046217.

30 Raetz, C. R., Ulevitch, R. I., Wright, S. D., Sibley, C. H., Ding, A., & Nathan, C. F. (1991) Gram-negative endotoxin: an extraordinary lipid with profound effects on eukaryotic signal transduction 1. *FASEB Journal*, **5**, 2652-2660.

35 Rebolledo-Gómez, M. & Wood, C. W. (2019) Unclear intentions: eavesdropping in microbial and plant systems. *Frontiers in Ecology and Evolution*, **7**, 1-13.

Rodaki, A., Bohovych, I. M., Enjalbert, B., Young, T., Odds, F. C., Gow, N. A., & Brown, A. J. (2009) Glucose promotes stress resistance in the fungal pathogen *Candida albicans*. *Molecular Biology of the Cell*, **20**, 4845-4855.

40 Rosen, R. (1985a) Anticipatory systems: Philosophical, Mathematical, and Methodological Foundations. Springer, New York, NY.

Rosen, R. (1978) Fundamentals of measurement and representation of natural systems (Vol. 1). North Holland.

Rosen, R. (1985b) Organisms as causal systems which are not mechanisms: an essay into the nature of complexity In *Theoretical Biology and Complexity* (165-203). Academic Press, Cambridge, MA.

5 Saito, M. E., Murakami, E., & Sudo, M. (1976) Circadian rhythms in disaccharidases of rat small intestine and its relation to food intake. *Biochemical and Biophysical Research Communications*, **421**, 177-179.

10 Saito, M. E., Nishida, T., Fuhisawa, Y., & Suda, M. (1976) Circadian rhythms of digestive enzymes in the small intestine of the rat. *Journal of Biochemistry*, **80**, 563-568.

Sánchez-Romero, M. A., Cota, I., & Casadesús, J. (2015) DNA methylation in bacteria: from the methyl group to the methylome. *Current Opinion in Microbiology*, **25**, 9-16.

15 Savageau, M. A. (1998) Demand theory of gene regulation. II. Quantitative application to the lactose and maltose operons of *Escherichia coli*. *Genetics*, **149**, 1677-1691.

20 Schild, S., Tamayo, R., Nelson, E. J., Qadri, F., Calderwood, S. B., & Camilli, A. (2007) Genes induced late in infection increase fitness of *Vibrio cholerae* after release into the environment. *Cell Host & Microbe*, **2**, 264-277.

Schulkin, J. (2009) Allostasis: a neural behavioral perspective. *Horm Behav*, **43**, 21-27 (2003).  
59 T. J. Perkins, Theodore and P. S. Swain. Strategies for cellular decision-making. *Molecular Systems Biology*, **5**, 1-15.

25 Selye, H. (1975) Homeostasis and heterostasis In *Trauma* (25-29). Springer, Boston, MA.

Shah, N. P. (2000) Probiotic bacteria: selective enumeration and survival in dairy foods. *Journal of Dairy Science*, **83**, 894-907.

30 Shudo, E., Haccou, P., & Iwasa, Y. (2003) Optimal choice between feedforward and feedback control in gene expression to cope with unpredictable danger. *Journal of Theoretical Biology*, **223**, 149-160.

35 Simpson, S. J. and Raubenheimer, D. (2009) Macronutrient balance and lifespan. *Aging*, **1**, 875-880.

Smith, G. P. (1995) Pavlov and appetite. *Integrative Psychology and Behavioral Science*, **30**, 169-74.

40 Smith, G. P. (2000) The controls of eating: a shift from nutritional homeostasis to behavioural neuroscience. *Nutrition*, **16**, 814-820.

Smith, C. M. & Clement, S. L. (2012) Molecular bases of plant resistance to arthropods. *Annual Review of Entomology*, **57**, 309-328.

Sivaprakasam, S., Shahverdiev, E. M., Spencer, P. S., & Shore, K. A. (2001) Experimental demonstration of anticipating synchronization in chaotic semiconductor lasers with optical feedback. *Physical Review Letters*, **87**, 154101.

5 Solon-Biet, S. M., Mitchell, S. J., de Cabo, R., Raubenheimer, D., Le Couteur, D. G., & Simpson, S. J. (2015) Macronutrients and caloric intake in health and longevity. *Journal of Endocrinology*, **226**, R17.

10 Soodak, H. and Iberall, A. (1978) Homeokinetics: A physical science for complex systems. *Science*, **201**, 579-582.

Southam, C. M. & Ehrlich, J. (1943) Effects of extract of western red-cedar heartwood on certain wood-decaying fungi in culture. *Phytopathology*, **33**, 517-524.

15 Stankiewicz, A. M., Swiergiel, A. H., & Lisowski, P. (2013) Epigenetics of stress adaptations in the brain. *Brain Research Bulletin*, **98**, 76-92.

20 Stephan, F. K. (2001) Food-Entrainable Oscillators in Mammals In *Circadian Clocks* (223-246). Springer, Boston, MA.

Sterling, P. (2004) Principles of Allostasis: Optimal Design, Predictive Regulation, Pathophysiology, and Rational In *Allostasis, Homeostasis, and the Costs of Physiological Adaptation*, (1-36) J. Schulkin (Eds.), Cambridge University Press, Cambridge, UK.

25 Sterling, P. & Eyer, J. (1988) Allostasis: A New Paradigm to Explain Arousal Pathology In, *Handbook of Life Stress, Cognition, and Health* (629-649), S. Fisher, J.T. Reason (Eds.), Wiley, Chichester.

30 Stevenson, N. R., Ferrigni, F., Parnicky, K., Day, S., & Fierstein, J. S. (1975) Effect of changes in feeding schedule on the diurnal rhythms and daily activity levels of intestinal brush border enzymes and transport systems. *Biochemica et Biophysica Acta*, **406**, 131-145.

35 Stevenson, N. R. & Fierstein, J. (1976) Circadian rhythms of intestinal sucrase and glucose transport: cued by time of feeding. *American Journal of Physiology*, **230**, 731-735.

Stoeckius, M., Grün, D., & Rajewsky, N. (2014) Paternal RNA contributions in the *Caenorhabditis elegans* zygote. *EMBO Journal*, **33**, 1740-1750.

40 Suzdalskaya, I. P. (1977) Increase in stability of isolated tissues and proteins caused by action of chemical stimulants. *Tsitologiya*, 19, 839-849. (in Russian)

Suzuki, N., Koussevitzky, S., Mittler, R., & Miller, G. (2012) ROS and redox signalling in the response of plants to abiotic stress. *Plant, Cell & Environment*, **35**, 259-270.

45 Svoboda, A. (1960) Un mode'le d'instinct de conservation (A model of the self-preservation instinct) In *Information Processing Machine. 7*. Prague: Czechoslovak Academy of Sciences

(147 –155). [English translation in Memorable Ideas of a Computer School: The Life and Work of Antonín Svoboda, ed. By G. J Klir, 2007, Prague: Czech Technical University Publishing House] (1960).

5 Tagkopoulos, I., Liu, Y. C., & Tavazoie, S. (2008) Predictive behavior within microbial genetic networks. *Science*, **320**, 1313-1317.

Tikhodeyev, O. N. (2018) The mechanisms of epigenetic inheritance: how diverse are they? *Biological Reviews*, **93**, 1987-2005.

10 Tse, C., Sera, T., Wolffe, A. P., & Hansen, J. C. (1998) Disruption of higher-order folding by core histone acetylation dramatically enhances transcription of nucleosomal arrays by RNA polymerase III. *Molecular and Cellular Biology*, **18**, 4629-4638.

15 Tuteja, N. & Sopory, S. K. (2008) Chemical signaling under abiotic stress environment in plants. *Plant Signaling & Behavior*, **3**, 525-536.

20 Vargas, C., Argandoña, M., Reina-Bueno, M., Rodríguez-Moya, J., Fernández-Aunión, C., & Nieto, J. J. (2008) Unravelling the adaptation responses to osmotic and temperature stress in *Chromohalobacter salexigens*, a bacterium with broad salinity tolerance. *Saline Systems*, **4**, 14.

25 Virilouvet, L., Avenson, T. J., Du, Q., Zhang, C., Liu, N., Fromm, M., Avramova, Z., & Russo, S. E. (2018) Dehydration stress memory: gene networks linked to physiological responses during repeated stresses of *Zea mays*. *Frontiers in Plant Science*, **9**, 1058.172.

Voss, H. U. (2000) Anticipating chaotic synchronization. *Physical Reviews E*, **61**, 5115.

30 Voss, H. U. (2002) Real-time anticipation of chaotic states of an electronic circuit. *International Journal of Bifurcation and Chaos*, **12**, 1619-1625.

Vos, I. A., Verhage, A., Schuurink, R. C., Watt, L. G., Pieterse, C. M., & Van Wees, S. (2013) Onset of herbivore-induced resistance in systemic tissue primed for jasmonate-dependent defenses is activated by abscisic acid. *Frontiers in Plant Science*, **4**, 539.

35 Waddington, C. H. (1958) The Strategy of the Genes: A Discussion of Some Aspects of Theoretical Biology. Allen & Unwin, London.

40 Waddington, C.H. (1968) Letter to Thom. Towards a Theoretical Biology 1 Prolegomena, Adline, Chicago.

Walker, S. L., Sojka, M., Dibb-Fuller, M., & Woodward, M. J. (1999) Effect of pH, temperature and surface contact on the elaboration of fimbriae and flagella by *Salmonella* serotype *Enteritidis*. *Journal of Medical Microbiology*, **48**, 253-261.

Wang, H., Cao, R., Xia, L., Erdjument-Bromage, H., Borchers, C., Tempst, P., & Zhang, Y. (2001) Purification and functional characterization of a histone H3-lysine 4-specific methyltransferase. *Molecular Cell*, **8**, 1207-1217.

Wang, X., Vignjevic, M., Liu, F., Jacobsen, S., Jiang, D., & Wollenweber, B. (2015) Drought priming at vegetative growth stages improves tolerance to drought and heat stresses occurring during grain filling in spring wheat. *Journal of Plant Growth Regulation*, **75**, 677-687.

Westerhoff, H. V., Brooks, A. N., Simeonidis, E., García-Contreras, R., He, F., Boogerd, F. C., Jackson, V. J., Goncharuk, V., & Kolodkin, A. (2014) Macromolecular networks and intelligence in microorganisms. *Frontiers in Microbiology*, **5**, 379.

Whitehead, A. N. (1929) Process and reality, an essay in cosmology. University Press, Cambridge, UK.

Whitehead, K., Pan, M., Masumura, K. I., Bonneau, R., & Baliga, N. S. (2009) Diurnally entrained anticipatory behavior in archaea. *PLoS One*, **4**, e5485.

Winata, C. L., Łapiński, M., Prysycz, L., Vaz, C., bin Ismail, M. H., Nama, S., Hajan, H. S., Lee, S. G. P., Korzh, V., Sampath, P., & Tanavde, V. (2018) Cytoplasmic polyadenylation-mediated translational control of maternal mRNAs directs maternal-to-zygotic transition. *Development*, **145**, 1-12.

Yeo, S. H., Franklin, D. W., & Wolpert, D. M. (2006) When optimal feedback control is not enough: feedforward strategies are required for optimal control with active sensing. *PLoS Computational Biology*, **12**, e1005190 (2006).

Zadeh, L. A. (1978) Fuzzy sets as the basis for a theory of possibility. *Fuzzy Sets and Systems*, **1**, 9-34.

Zadeh, L. A. (2001) A new direction in AI: toward a computational theory of perceptions. *Accuracy International Magazines*, **22**, 73–84.

Zaugg, M., Lucchinetti, E., Spahn, D. R., Pasch, T., & Schaub, M. C. (2002) Volatile anesthetics mimic cardiac preconditioning by priming the activation of mitochondrial KATPChannels via multiple signaling pathways. *Anesthesiology*, **97**, 4-14.

Zhao, K., Liu, M., & Burgess, R. R. (2007) Adaptation in bacterial flagellar and motility systems: from regulon members to ‘foraging’-like behavior in *E. coli*. *Nucleic Acids Research*, **35**, 4441-4452.



**Fig. 1.** Example showing the time course of a hypothetical environmental factor (a), the time course of two signals that are correlated with this variable (b), and a demonstration of how the temporal relationship between the environmental variable, signal, and response can be used to distinguish between reactive and anticipatory processes (c).

**Fig. 2.** Anticipatory regulation of sugar metabolism in *E. coli*. The movement of *E. coli* from the proximal end of the small intestine toward the distal end is associated with a predictable sequence of exposure, first to lactose followed by maltose (a). Expression of the lac operon occurs in a reactive manner once lactose is present, while the expression of specific maltase genes exhibits anticipatory regulation. Maltase genes are up-regulated in the presence of maltose but also significantly expressed upon exposure to lactose in the absence of maltose (b). This regulatory structure induces maltase production in anticipation of future maltose availability and is associated with increased fitness. The data shown here are from Mitchell et al. (2009). Only the expression specific promoters were measured (data for lacY and lacA and only five of the ten maltase gene promoters are shown).

**Fig. 3.** Diagram showing the key events in a reactive response and a generalized anticipatory response, including the manifestation of error, the initiation of the preparatory phase, downstream reactions, and commencement. Notice that although the duration of each response ( $D_r$  and  $D_a$ ) is the same, the anticipatory response has a shorter response time ( $RT_a$ ) than the reaction response ( $RT_r$ ) due to the initiation of the preparatory phase occurring before the manifestation of error.

**Fig. 4.** Summary of key criteria for biological anticipation at different levels of organization, including causal, ecological, physiological, and biochemical considerations.

**Fig. 5.** Graphical representation of the different types of anticipatory processes. Panel (a) shows how circadian rhythms can regulate the timing of physiological processes in an anticipatory manner. This figure is adapted from Millar and Kay (1996) and shows the up-regulation of chlorophyll a/b-binding protein (CAB) expression in *Arabidopsis* seedlings before the morning increase in light availability. Panel (b) shows asymmetrical anticipatory regulation (AAR), which occurs when a response is regulated by a stimulus that is correlated to but predates an environmental variable, allowing the response to be carried out in anticipation of the environmental change. Panel (c) shows priming in a plant-insect model system, where previously attacked, or primed, plants are more resistant than naïve plants to future attack. Panel (d) shows the J-response curve characteristic in hormesis, where an over-response to a specific stressor can provide beneficial protection from other more general stressors. Panel (e) demonstrates how parental expressional states can be passed on to offspring to increase fitness in predicted future conditions via transgenerational effects, shown here by heritable gene methylation and histone modifications.